DISSERTATION ON INSULIN RESISTANCE IN NON DIABETIC PATIENTS WITH CORONARY ARTERY DISEASE

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CERTIFICATE

This is to certify that the dissertation entitled "INSULIN RESISTANCE IN NON DIABETIC PATIENTS WITH CORONARY ARTERY DISEASE" is the bonafide original work of Dr. M. Santni, post graduate student, Department of General Medicine, Kilpauk Medical College, Chennai under my guidance in partial fulfillment of the requirements for MD (General Medicine) branch I of the Tamil Nadu Dr. M.G.R Medical university. The period of postgraduate study and training was from May 2007 to March 2010.

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INTRODUCTION

INTRODUCTION

Cardiovascular disease accounts for approximately 12 million deaths annually & is the commonest cause of death globally. Previously considered a disease of the affluent, the past three decades have seen considerable decline in the incidence and prevalence of atherosclerotic coronary disease in the industrialized western world; whereas at the same time this problem is assuming epidemic proportions in the developing world.

Asian Indians, whether living in their own country or elsewhere, have higher rate of coronary artery disease than any other ethnic groups studied (1-3). Not only CAD is more prevalent, it is more severe and occurs at a younger age in Asian Indians. (3) Even within India, the prevalence of CAD is not homogeneous and is two fold higher in southern parts. The higher prevalence of CAD in Asian Indians is accompanied by paradoxically, a lower prevalence of conventional risk factors such as hypertension, diabetes mellitus, hyperlipidemia and cigarette smoking (4,5). This suggests the prevalence of other risk factors.

Hyperinsulinemia and Insulin resistance have been associated with obesity, Type 2 diabetes mellitus, hypertension, hyperlipidemia and CAD, however the link between endogenous hyperinsulinemia and CAD in nondiabetic adults is at best weak with conflicting results, perhaps suggesting ethnic differences. Recent physiological studies have shown that many characteristics of insulin resistance syndrome are more prevalent in Asian Indians compared to Caucasians (13,14,15). Impaired glucose tolerance, elevated fasting plasma glucose and insulin clamp and other techniques, have all been observed in Asian Indian populations (16,17,18,19). In addition to decreased insulin sensitivity, Asian Indians have lipid abnormalities, including increased triglycerides, low HDL and increased LDL, all may contribute to CAD. The present study was designed to examine the relationship between Insulin sensitivity and coronary artery disease in non diabetic patients.

AIM OF THE STUDY

AIM OF STUDY

To evaluate the association of plasma insulin and insulin resistance (IR) measured by Homeostasis Model Assessment (HOMA-IR) with coronary artery disease (CAD) in non diabetic subjects.

To estimate whether insulin resistance alone without diabetes is an independent risk factor for Coronary artery disease.

REVIEW OF LITERATURE

REVIEW OF LITERATURE

INSULIN ACTIONS:

Insulin has unique properties as a hormone when compared to other hormones. Insulin is an anabolic hormone, apart from glucose homeostasis, induces protein synthesis, lipogenesis and regulation of various genes. Insulin also has anti inflammatory and anti apoptotic actions & protects endothelial functions

Insulin regulates diverse physiological processes in mammals, including membrane transport, intermediary metabolism and cell growth and differentiation. The most conspicuous metabolic effects of insulin are associated with skeletal muscle, cardiac muscle, adipose tissue and liver either having stimulatory or inhibitory effect. A surprising feature is that to date the only signalling component known to be unique to insulin action is the insulin Receptor (IR) itself, which is widely expressed in mammalian cells, although levels vary greatly between cell types. Thus normal insulin action influences wide range of tissues and insulin resistance can adversely affect diverse tissue functions, though not all to same extent.

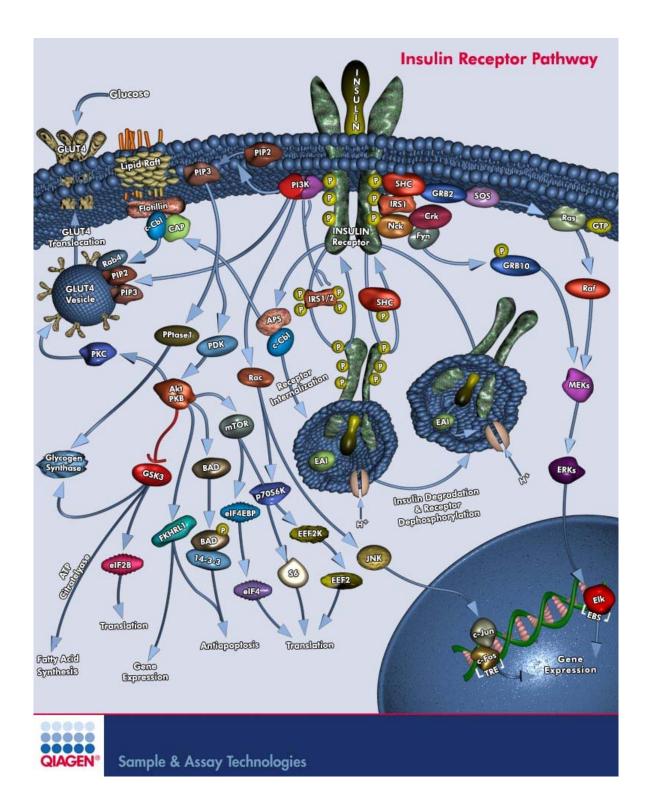
THE INSULIN RECEPTOR FAMILY

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The insulin receptor is a large, hetero tetrameric, transmembrane glycoprotein containing two types of subunits designated alpha

(Mr.140.kDa) and beta (95 k Da), linked by disulphide bonds in a beta alpha - alpha- beta configuration (7). The IR binds insulin with high affinity and specificity and transmits a signal to the cytosol via its intrinsic tyrosine - specific protein kinase activity (8) This phosphorylates a number of intracellular substrates, especiall most y the so-called insulin receptor substrates (IRS), which recruit and activate an array of signalling proteins containing Src homology -2 (SH2) domains. (9). Two signals have been shown to play major role in insulin action, namely those transmitted by enzyme phospho-inositide 3 kinase (PI 3kinase), which generates ptd Ins (3,4,5) tris-phosphate at the cytosolic face of membrane and the guanine nucleotide exchange factor Grb2/sos which activates the small G-protein Ras. These act as switch mechanism to change the "currency" of signalling from tyrosine phosphorylation to serine / threonine phosphorylation of target proteins. However these signals and the downstream signalling cascade involving protein kinase B and mitogen activated protein kinases (MAPKs) have been implicated in the specific actions of Insulin.

Metabolic and antiapoptotic effects of insulin are mediated by signalling pathway involving IRS proteins, (PI-3k), protein kinase B and mToR (Mamalian target of Rapamycin). In contrast, non metabolic,



proliferative and mitogenic effects are mediated largely via activation of Ras, Raf and mitogen activated protein kinase Erk -1, Erk-2.

There is accumulating evidence that obesity associated insulin resistance reflects inhibitory influences on this pathway at the level of insulin receptor substrates (IRS). Insulin resistance syndromes described in adults are either type A IR due to defect in insulin signalling pathways or type B IR due to auto antibodies directed at insulin receptors.

INSULIN RESISTANCE:

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Insulin Resistance is defined as an inability of a known quantity of exogenous or endogenous insulin to increase glucose uptake & utilization in an individual as such as it does in a normal person. Both insulin resistance and decreased insulin secretion are genetically programmed. This program is modified by a variety of environmental factors especially diet and activity (11).

It was in 1988, at the Banting lecture of the American Diabetes Association Annual meeting, that "Dr Gerald Reaven" first proposed the concept that insulin resistance and compensatory hyperinsulinemia were the cornerstones of a plurimetabolic syndrome that included hyper triglyceridemia, reduced plasma HDL cholesterol levels, essential hypertension and some degree of glucose intolerance (12). The concept was first referred to as "syndrome X", implied that the Insulin Resistance syndrome might be associated with significant elevation in the risk of cardiovascular disease. It was also suggested that a significant proportion of non-diabetic individuals in the general population perhaps as many as 2.5 percent, might display some features of the syndrome. Both IR and decreased insulin secretion are genetically programmed which is modified by a number of environmental factors especially diet & activity(16).

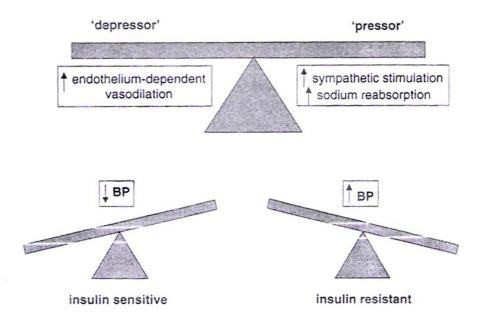
INSULIN RESISTANCE AND HYPERTENSION

The direct association between hyperinsulinemia, insulin resistance and essential hypertension that could not be attributed to confounding obesity was most convincingly demonstrated by Ferrannini and colleagues in 1987 (20). A number of epidemiological studies have also been demonstrated on the association between insulin levels and blood pressure (21).

It is unlikely that hyperinsulinemia directly causes hypertension and the relationship between insulin resistance and vascular dysfunction is not direct and simple. Patients with insulinoma do not tend to have hypertension (22). Nevertheless prospective studies have shown that individuals with hyperinsulinemia have a risk of developing both hypertension and coronary events (23).

Insulin Resistance is absent in secondary hypertension patients, but present in normotensive offsprings of essential hypertensive patients(24) suggesting that it may precede the development of high blood pressure.

ENDOTHELIAL DYSFUNCTION AND ATHEROTHROMBOTIC DISEASE



The relation between IR and hypertension relates to several different mechanisms. First it is important to note that insulin is a vasodilator, that is mediated by nitric oxide release from endothelial cells, when given intravenously to people of normal weight (25) with secondary effects on sodium absorption in the kidney (26). Evidence indicate that sodium reabsorption is increased in white people but not in Africans or Asians. (27). In the setting of Insulin Resistance, the vasodilator effect of insulin can be lost (28), but renal effect is preserved (29). Because nitric oxide accounts for all of insulin induced vasodilation, it is likely that hypertension is result of partial nitric oxide deficiency or resistance (30).

Insulin also increases the activity of sympathetic nervous system (31) an effect that might also be preserved in the setting of insulin resistance (32) due to hyperinsulinemia. In pathophysiological states such as obesity, the balance may be disrupted by enhanced sympathetic activation in response to hyperinsulinemia together with "blunting" of insulin mediated vasodilatation (Vascular insulin resistance).

Insulin Resistance and Dyslipidemia

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Insulin Resistance is associated with many lipid abnormalities, including increased triglycerides, low high density lipoprotein (HDL) cholesterol and increased lipoprotein (a), all of which may contribute to excess coronary artery disease (33,34).

In general, with increases in free fatty acid flux to the liver, increased production of apo B containing triglyceride rich VLDL occurs (35). The effect of insulin on this process is complex. In the setting of insulin resistance, increased flux of free fatty acid to the liver increases hepatic triglyceride synthesis but under physiological conditions, insulin inhibits VLDL secretion in systemic circulation (36). This response in part is an effect of insulin on the degradation of apo B. Yet insulin is also lipogenic, increasing transcription of many genes related to triglyceride biosynthesis.

Additionally, Insulin Resistance could also reduce the concentration of lipoprotein lipase in peripheral tissues (ie in adipose tissue more than muscle). This alteration in lipoprotein lipase, however contributes less to hypertriglyceridemia than does overproduction of VLDL. Nevertheless hypertriglyceridemia is an excellent reflection of insulin resistance condition.

In the presence of hypertriglyceridemia, a decrease in the cholesterol content of HDL results from decreases in cholesteryl ester content of the lipoprotein core, with variable increase in triglyceride, making the particle small and dense, a function in part of cholesteryl ester transfer protein (38). This change in lipoprotein composition also results in an increased clearance of HDL from circulation (39). The relation of these changes in HDL to insulin resistance is probably indirect, arising in concert with changes in triglyceride rich lipoprotein metabolism.

In addition to HDL, the composition of LDL is also modified in similar way. In fact, with fasting serum triglycerides >2mmol/L, almost all patients have a predominance of small dense LDL (40). This change in LDL composition is attributable to relative depletion of unesterified and esterified cholesterol and phospholipids, with either no change or an increase in LDL Triglyceride (41).

Small dense LDL is more atherogenic than buoyant LDL because

- (i) it is more toxic to the endothelium,
- (ii) it is more able to transit through the endothelial basement membrane,
- (iii) it adheres well to glycosaminoglycans,
- (iv) it has increased susceptibility to oxidation,
- (v) it is more selectively bound to scavenger receptors in macrophages(42).

Hypertriglyceridemia is a major determinant of the distribution of LDL particles. The higher the fasting triglyceride level, the greater the preponderance of small dense LDL in total LDL concentration (43).

Insulin Resistance and Obesity

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Recent studies of body fat distribution have shown that insulin resistance is associated with central obesity and visceral fat (13,14) importantly. The increase in central obesity often is not apparent from measurement of body mass index (BMI), which may be in the normal range as defined by standard weight tables and other readily available criteria (44,45).

International Association for the study of obesity and the international obesity task force defined central obesity as waist circumference > 80cm for women and >90cm for men (46).

A distinction between a large waist due to increase in subcutaneous adipose tissue versus visceral fat is debated. This distinction can be made with computed tomography or MRI (47).

With increases in intra-abdominal or visceral adipose tissue, a higher rate of flux of adipose tissue derived free fatty acids to liver through splanchinc circulation would be expected, whereas increases in abdominal subcutaneous fat would release lipolysis products into the systemic circulation and avoid more direct effects on hepatic metabolism (ie glucose production, lipid synthesis, and secretion of prothrombotic proteins such as fibrinogen and plasminogen activator inhibitor - 1 (48)).

Despite these potential differences in mechanisms related to excessive abdominal adipose tissue distribution, the clinical diagnosis of metabolic

syndrome does not distinguish between increases in subcutaneous and visceral fat.

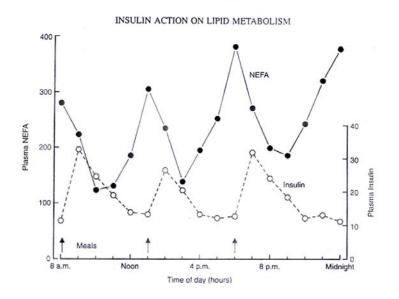
Yet perhaps by a mechanism related to free fatty acid flux and metabolism, the relative predominance of visceral rather than increase in adipose tissue with increasing waist circumference in Asians and Asian Indians (49) renders the relative prevalence of the syndrome higher than in African American men in whom subcutaneous fat predominate (50).

However, there is evidence that the elevated post prandial free fatty acid release in upper body obese women originates from the non-splanchnic upper body fat, and not from the visceral depot (51). These results suggest that visceral fat might be a marker for, but not the source of excess post prandial free fatty acid in obesity.

The effect of insulin to reduce circulating Non Esterified fatty acid (NEFA) concentrations is an important part of coordination of metabolic processes that occurs after a meal. At that time, glucose becomes the major oxidative fuel for skeletal muscle and it is appropriate that "substrate competition" from fatty acids is minimized.

Also, plasma NEFAs are potent stimulus for hepatic gluconeogenesis and glucose output and again this stimulus is not appropriate in the postprandial period when hepatic glucose output needs to be suppressed to maintain glucose homeostasis

The plasma NEFA concentrations display a marked diurnal variation, the reverse of insulin concentration, with troughs after meals and peaks before the next meal.



Randle's hypothesis

The glucose fatty acid hypothesis proposed by Randle et al in 1963 attempted to delineate the relationship between increased non - esterified fatty Acids (NEFAs) and insulin resistance. Randle and co-workers conducted a series of invitro experiments in rat cardiac muscle that suggested substrate competition between NEFAs and glucose as an energy source for muscle. These studies observed a relative increase in the rate of fat oxidation compared with carbohydrate metabolism in response to increase in NEFAs. In addition, studies revealed a reduction in insulin stimulated glucose uptake and utilization by the cardiac muscle.

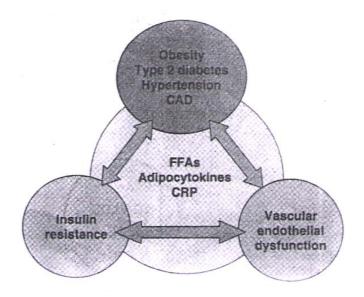
The mechanism by which NEFAs bring about insulin resistance may be explained by recent studies that reveal NEFAs, induce different isoforms of protein kinase C. These isoforms can interfere with the intracellular signalling pathway of insulin and ultimately block glucose transport activity.

Insulin resistance and increased NEFA supply to liver are associated with increased prevalence of Non – alcoholic steatohepatitis (NASH), which contributes to liver fibrosis and cirrhosis in a proportion of patients.

Insulin Resistance and Atherosclerosis

The risk of coronary artery disease is greatly increased in the type 2 diabetes. In a study examining the seven year incidence of myocardial infarction, Haffner et al (54) reported that the 10 year risk of a non-diabetic subject without previous MI was 3.5 percent, if a subject had previously had an MI, the risk of a further event was 18.8 percent. In contrast in diabetic subjects without previous MI, the risk was comparable to that of a non diabetic post MI subject (20.2 percent), while the risk of second MI in a

diabetic subject was 45 percent. Thus, diabetes that is characterised by insulin resistance and hyperinsulinemia is associated with an accelerated risk of atherosclerosis.



However, in patients without overt type 2 DM, the independent association between insulin resistance and atherosclerosis risk has been less easy to demonstrate. Nevertheless, there are now several good epidemiological studies that suggest that insulin resistance is an independent risk factor for cardiovascular disease. For example, surrogate measures of insulin resistance are associated with carotid artery intima-media thickness, a measure of atherosclerosis (55,56).

As yet there are no prospective data on the predictive role of insulin resistance on cardiovascular event rate, although the European group for the study of insulin resistance (EGIR) is currently conducting a large multicenter study to address the issue.

Adipokines and Insulin Resistance

The adipose tissue is now regarded as a major endocrine organ and a variety of factors released by adipose cells potentially mediate insulin resistance (57). Evidence suggest that one or more of these adipokines could impair insulin signalling and cause insulin resistance early in pre diabetics. (58) These factors include tumour necrosis factor TNF α , leptin, Interleukin 6 (IL-6) and recently resistin and adiponectin: HSL : hormone sensitive lipase, IL-6: interleukin - 6, LPL: lipoprotein lipase, NEFA non esterified fatly acids, PAI-1, Plasminogen activator Inhibitor -1; SR: soluble receptor, TNF α tumour necrosis factor – alpha, VLDL -TG : very low density lipoprotein - triglyceride. However the precise role of these factors and the molecular mechanisms whereby they generate insulin resistance have remained elusive (59). Recently adiponectin has gained significance as a mediator of insulin sensitivity. Many studies reported lower levels of adiponectin in insulin resistant states and plasma adiponectin is inversely associated with overall and central fat distribution.

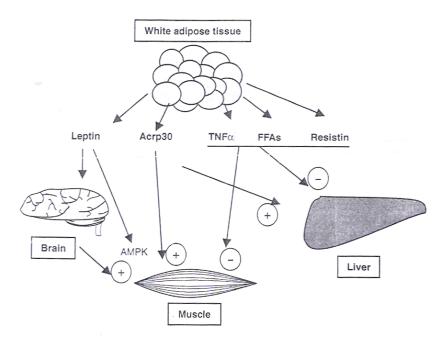


Figure 5.2 Adipose tissue (AT) as an endocrine organ

Dietary Factors in Insulin Resistance

Obesity is strongly linked to impaired insulin sensitivity, but acute changes in total energy intake influence insulin independently of changes in body weight or fat mass. (59).

A high intake of saturated fat reduces insulin sensitivity, but that of mono unsaturated and poly unsaturated fat are neutral or beneficial at least in the setting of moderate total fat intake

The effect of carbohydrate are less clear. However, unrefined carbohydrate with a low glycemic index, whole grain and high fibre foods

appear to have beneficial effects on insulin sensitivity compared with more refined carbohydrates.

There may also be an influence of specific micronutrients such as magnesium, chromium and vitamin E, however, the evidence is limited

Exercise and Physical Activity

Physical activity has a positive effect on risk factors and diseases, in obese population. Though the exact mechanism is not known, exercise has many benefits for health and CAD risk factors, and reduces cardiovascular morbidity and mortality.

Exercise has a favorable action on modifying blood lipids, particularly HDL cholesterol in overweight individuals. Other studies have shown beneficial effect on blood pressure. Exercise has been shown to increase insulin sensitivity, GLUT-4 concentration and glucose disposal (66). Regular physical exercise improves endothelial function, by increasing vasculature shear stress and by increased production of nitric oxide. (67).

In overweight and insulin resistant subjects, exercise has been shown to reduce the non traditional cardiovascular risk factor homocysteine. It is known that regular exercise, even in overweight individuals leads to an improvement in fibrinolytic system. (69). However little is gained unless long term changes are established and this is known to be difficult to acheive in the majority of individuals. The compliance is generally poor.

Measurement of Insulin Resistance

The concept of insulin resistance is relatively easy to understand, but determining precisely who is insulin resistant is more complicated (70).

A World Health Organization consensus group recently concluded that the insulin sensitivity index (SI) of the lowest 25 percent of a general population can be considered insulin resistant (71). The European group for the study of insulin resistance took a more restricted view, defining insulin resistance as the SI of the lowest 10 percent of a non-obese, non-diabetic, normotensive Caucasian population. Richard Legro and his associates also used the SI of the lowest 10 percent of an obese, non PCOS population to define insulin resistance (72).

Hyperinsulinemic Euglycemic Clamp

The gold standard for evaluating insulin sensitivity, this clamp technique requires a steady intravenous infusion of insulin to be administered in one arm. The serum glucose level is "clamped" at a normal fasting concentration by administering a variable intravenous glucose infusion ((73) in the other arm. Numerous blood samplings are taken to monitor serum glucose so that a steady "fasting" level can be maintained. (In theory the intravenous infusion should completely suppress hepatic glucose production and not interfere with the test's ability to determine how sensitive target tissues are to the hormone).

The degree of insulin resistance should be inversely proportional to the glucose uptake by target tissues during the procedure. In other words, the less glucose that is taken up by tissues during the procedure, the more insulin resistant the patient is.

The hyperinsulinemic - euglycemic clamp technique is most scientifically sound technique for measuring insulin sensitivity and it is against this standard that all other tests are usually compared Because this and similar "clamp" techniques are expensive, time consuming, and labour intensive, they are not very practical in an office setting.

To overcome these obstacles, alternative tests have been developed, including the frequently sampled intravenous glucose tolerance test (FSIVGTT), insulin tolerance test (ITT), insulin sensitivity test (IST), and continuous infusion of glucose with model assessment (CIGMA). Unfortunately all of these methods require intravenous access and multiple venipunctures, making them relatively impractical for office assessment. The oral glucose tolerance test (OGTT) does not require intravenous access but does involve several venipunctures and 2 to 4 hours of patient and technician time. Each of these tests have been shown to correlate reasonably well with dynamic clamp techniques.

Non Dynamic Measurements of Insulin Sensitivity

These tests, which include fasting insulin alone, are the HOMA test (homeostasis assessment model) (78 and 79) and the QUICKI (Quantitative Insulin Sensitivity Check Index). These tests are simple to perform, being based on fasting insulin and glucose measurements alone.

HOMA has been widely employed in clinical research to assess insulin sensitivity. Rather than using fasting insulin or a G/I ratio, the product of the fasting values of glucose (GO) expressed as mg/dl and insulin (IO) expressed as mcU/ml is divided by a constant.

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The constant 405 should be replaced by 22.5 if glucose is expressed in S.I. Units. Unlike IO and the G/I ratio, the HOMA calculation compensates for fasting hyperglycemia. The HOMA value correlates well with clamp techniques and has been frequently used to assess changes in insulin

sensitivity after treatment. HOMA has also been used to study insulin resistance among PCOS patients of differing ethnic origins.

Like HOMA, QUICKI can be applied to normoglycemic and hyperglycemic patients. It is derived by calculating the inverse of the sum of logarithmically expressed values of fasting glucose and insulin:

Both HOMA and QUICKI values increase in the insulin – resistant patients.

Ischemic Heart Disease

Ischemic heart disease is defined as myocardial impairment due to imbalance between coronary blood flow and myocardial requirement. The commonest cause of IHD is atherosclerotic coronary artery disease

Spectrum of myocardial ischemia.

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Traditionally, ischemic heart disease has been divided into several seperate syndromes such as stable effort angina, unstable angina, non -Q wave MI and Q wave MI. The recent understanding of the conversion of a stable atherosclerotic lesion to a plaque rupture with thrombosis has

provoked a unifying hypothesis for the aetiology of acute coronary syndrome.

| Old | Present |
|----------------------|---|
| Asymptomatic | Asymptomatic |
| Stable effort angina | Stable effort angina |
| Unstable angina | Acute coronary syndrome, unstable angina without necrosis |
| Non Q wave MI | Non ST elevation MI (evidence of myocardial necrosis) |
| Q wave MI | ST elevation MI |
| Sudden death | Sudden death |
| Silent ischemia | Ischemic cardiomyopathy |

Nomenclature of myocardial ischemia

The concept of myocardial ischemia as a spectrum provides framework for understanding the pathogenesis, clinical features and outcome of patients

Pathophysiology

Atherosclerotic obstruction of the coronary arteries leads to a spectrum of clinical syndromes varying between effort or stable angina to acute coronary syndrome (ACS) where an atheromatous plaque becomes vulnerable to rupture and produces sudden, complete or partial occlusion of the coronary artery. A complete occlusion leads to ST elevation myocardial infarction (STEMI) whereas partial occlusion leads to non – ST elevation myocardial infarction (NSTEMI)

Site & size of MI

| Left anterior descending (LAD) | - Antroseptal MI | |
|--------------------------------|---------------------------|--|
| Left circumflex artery | - Anterolateral MI | |
| Right coronary artery | - Inferoposterior wall MI | |
| | with or without RVMI | |

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MATERIALS AND METHODS

MATERIALS AND METHODS

The study was conducted at the out patient department of cardiology at Government Royapettah Hospital.

Selection of Cases

A total of fifty patients with coronary artery disease were chosen from the outpatient department of cardiology.

Fifty healthy controls matched for age and glycemia were chosen from the master health check up at Government Royapettah hospital.

Women chosen for the study were in the post menopausal age group or in follicular phase of their menstrual cycle to decrease the potential influence of gonadal steroids on insulin action.

Inclusion criteria

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Coronary artery disease in the study was defined as:

- Documented myocardial infarction, substantiated by Q waves in electrocardiogram (ECG) and / or regional wall motion abnormality on echocardiogram,
- Angina supported by (i) ST depression or T wave inversion and/or
 (ii) coronary angiographic evidence of >70% stenosis of one or more vessels.

A control was defined as an induvidual without a history of angina and with a normal ECG

Exclusion criteria

- (i) Diabetes Mellitus, which was diagnosed according to American diabetes association (80)
- (ii) Patients with fasting plasma glucose over 110 mg/dl.
- (iii) Acute heart failure
- (iv) Hepatic dysfunction
- (v) Serum creatinine>1.4 mg/dl
- (vi) Patients receiving hormone replacement therapy
- (vii) Patients on antidiabetic agents or steroid therapy
- (viii) Patients who had an acute coronary event in the past 6 weeks.

A detailed history and physical examination was carried out in each patient including measurement of blood pressure, height and weight. All the candidates were screened for blood chemistry abnormalities.

Anthropometric measurements

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Height and weight were measured by standard procedures. Body mass index was calculated by using the formula

$$BMI = \frac{Weight in kg}{(Height in m)^2}$$

The waist to hip circumference ratio (WHR) was measured using a flexible tape. The waist circumference was measured in the horizontal plane above the iliac crest. The hip circumference was measured at the maximum circumference at the level of the femoral trochanters.

Biochemical Analysis

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In our patients, samples were collected from venous blood after overnight fasting and blood chemistry was performed. Fasting blood sugar was measured by enzymatic oxidase method. The concentrations of serum lipids including total cholesterol, triglyceride, HDL cholesterol &LDL cholesterol were measured. LDL cholesterol was calculated using Fridewald formula. Insulin was estimated by ELISA assay.

Insulin resistance was calculated by the homeostasis model assessment of insulin resistance (HOMA-IR), proposed by Mattews et al., whose formula was (78).

HOMA -IR =
$$\frac{\text{Fasting glucose (mgldl) x fasting insulin (µU/ml)}}{405}$$

Fasting insulin and HOMA –IR were used as surrogate markers of insulin resistance in our study.

STATISTICAL ANALYSIS

STATISTICAL ANALYSIS

Data were expressed as mean \pm SD or geometric mean (95% confidence interval). The case and control group were analyzed using unpaired t tests. The categorical variables were analyzed using x² or Fisher exact test. The association between insulin resistance and all other parameters were first analyzed by univariate analysis. Multivariate analysis to evaluate associations of CAD was performed by using multivariate logistic regression tests. A p value <0.05 was considered statistically significant.

OBSERVATIONS AND ANALYSIS

OBSERVATIONS AND ANALYSIS

Clinical characteristics of the study subject and baseline laboratory results are given in Tables I and 2.

Group = Control Group

Table -1

| | Ν | Mean | Std. Deviation |
|--------------------|----|----------|----------------|
| | 1 | Witcum | Stu: Deviation |
| AGE | 50 | 51.1800 | 5.95798 |
| BMI | 50 | 27.4908 | 3.124834 |
| WHR | 50 | .7390 | .08229 |
| SBP | 50 | 125.1600 | 15.05956 |
| DBP | 50 | 79.0000 | 6.46813 |
| HDL | 50 | 41.6600 | 3.05467 |
| LDL | 50 | 97.5200 | 12.09021 |
| TGL | 50 | 143.9400 | 23.70861 |
| ТС | 50 | 189.4800 | 30.79109 |
| FBS | 50 | 91.7800 | 8.62907 |
| SI | 50 | 11.4140 | 2.49399 |
| HOMA-IR | 50 | 2.6974 | .55524 |
| Valid N (listwsie) | 50 | | |

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Descriptive statistics

Group = Case Group

Table – 2

DESCRIPTIVE STATISTICS

| | Ν | Mean | Std. Deviation |
|-------------------|----|----------|----------------|
| AGE | 50 | 51.600 | 6.91641 |
| BMI | 50 | 24.4288 | 3.03031 |
| WHR | 50 | .8452 | .09943 |
| SBP | 50 | 126.0000 | 13.09307 |
| DBP | 50 | 78.6000 | 5.34907 |
| HDL | 50 | 32.2800 | 3.72548 |
| LDL | 50 | 100.5600 | 20.26272 |
| TGL | 50 | 174.8600 | 27.86557 |
| ТС | 50 | 204.3400 | 41.26352 |
| FBS | 50 | 100.5600 | 6.07827 |
| SI | 50 | 21.6040 | 6.23892 |
| HOMA-IR | 50 | 5.3952 | 1.75469 |
| Valid N(listwsie) | 50 | | |

In the studied subjects age wise distribution of patients were as follows

Table - 3

Frequency table

| Age | Cases | Control |
|-------|-------|---------|
| 40-50 | 24 | 21 |
| 50-60 | 20 | 26 |
| >60 | 6 | 3 |

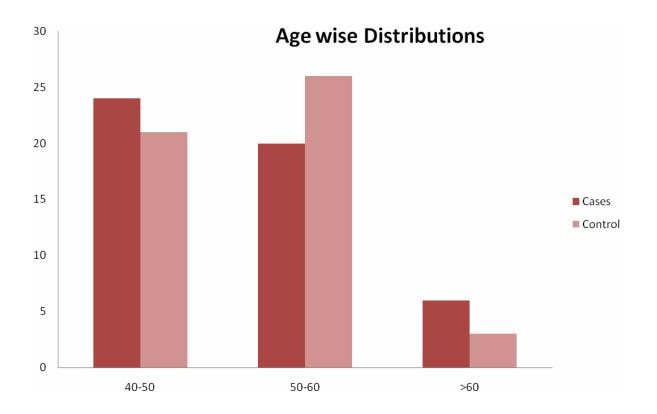
COMPARISON OF AGE: T TEST

GROUP STATISTICS

| | Group | Ν | Mean | Std. Deviation | Std. Error Mean |
|-----|---------------|----|---------|----------------|-----------------|
| AGE | Case Group | 50 | 51.6000 | 6.91641 | .97813 |
| | Control Group | 50 | 51.1800 | 5.95798 | .84259 |

p value : 0.746

p-Value of 0.746 indicates the difference in age across case and Control groups is not significant. In other words both Case and Control groups have subjects with similar age group.



Comparison of BMI (T test)

Table - 5

Group Statistics

| BMI Case Group 50 24.4288 3.03031 .42855 Control Group 50 27.4908 3.124834 4.41918 | Mean | Std. Error | Std. Deviation | Mean | N | Group | |
|--|------|------------|----------------|---------|----|---------------|-----|
| Control Group 50 27.4908 3.124834 4.41918 | | .42855 | 3.03031 | 24.4288 | 50 | II Case Group | BMI |
| | | 4.41918 | 3.124834 | 27.4908 | 50 | Control Group | |

p. Value : 0.494

p-Value of 0.494 indicates the difference in BMI across Case and Control groups is not significant (i.e.,) the BMI of subjects in Case and Control groups is the same.

Comparison of WHR (T test)

Table 6

Group Statistics

| (| Group | N | Mean | Std. Deviation | Std.Error Mean |
|-----|---------------|----|-------|----------------|----------------|
| WHR | Case Group | 50 | .8152 | 0.09943 | .01406 |
| | Control Group | 50 | .7390 | 0.8229 | .01164 |

p-Value < 0.01

p- Value less than 0.01 indicates the difference in WHR across Case and Control groups is highly significant (Significant at 1%). The group statistics table shows Case group is associated with higher waist to hip ratio than control group inspite of having normal BMI, indicating a tendency towards central obesity.

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Comparison of SBP (T test)

Table – 7

GROUP STATISTICS

| | Group | N | Mean | Std. Deviation | Std. Error Mean |
|-----|---------------|----|----------|----------------|-----------------|
| SBP | Case Group | 50 | 126.0000 | 13.09307 | 1.85164 |
| | Control Group | 50 | 125.1600 | 15.05956 | 2.12974 |

p-Value: 0.767

p-Value of 0.767 indicates the difference in SBP across Case and Control group is not significant.

Comparison of DBP (T test)

Table – 8

GROUP STATISTICS

| | Group | N | Mean | Std. Deviation | Std. Error Mean |
|-----|---------------|----|---------|----------------|-----------------|
| DBP | Case Group | 50 | 78.600 | 5.34904 | .75647 |
| | Control Group | 50 | 79.0000 | 6.46813 | .9473 |

p - Value : 0.737

p- Value of 0.737 indicates the difference in DBP across Case and Control groups is not significant.

Comparison of HDL (T test)

Table – 9

GROUP STATISTICS

| | Group | Ν | Mean | Std. Deviation | Std. Error Mean |
|-----|---------------|----|---------|----------------|-----------------|
| HDL | Case Group | 50 | 32.2800 | 3.72548 | .52686 |
| | Control Group | 50 | 41.660 | 3.05467 | .43200 |

p value : <0.01

p-Value less than 0.01 indicates the difference in HDL across Case and Control groups is highly significant (Significant at 1%). The Group statistics table shows Case group is associated with lower HDL than control group.

Comparison of LDL (T test)

Table – 10

GROUP STATISTICS

| | Group | Ν | Mean | Std. Deviation | Std. Error Mean |
|-----|---------------|----|----------|----------------|-----------------|
| LDL | Case Group | 50 | 100.5600 | 20.26272 | 2.86558 |
| | Control Group | 50 | 97.5200 | 12.09021 | 1.70981 |

p - Value : 0.365

p-Value of 0.365 indicates the difference in LDL across Case and Control groups is not significant.

Comparison of TGL (T test)

Table -11

| Group | | Ν | Mean | Std. Deviation | Std.Error Mean |
|-------|---------------|----|----------|----------------|----------------|
| TGL | Case Group | 50 | 174.8600 | 27.86557 | 3.94079 |
| | Control Group | 50 | 143.9400 | 23.70861 | 3.35290 |

p - Value < 0.01

p-Value less than 0.01 indicates the difference in TGL across Case and Control groups is highly significant (Significant at 1%). The Group statistics table shows Case group is associated with higher TGL than control group.

Comparison of TC (T test)

Table – 12

GROUP STATISTICS

| Grou | ър | Ν | Mean | Std. Deviation | Std. Error Mean |
|------|---------------|----|----------|----------------|-----------------|
| ТС | Case Group | 50 | 204.3400 | 41.26352 | 5.83554 |
| | Control Group | 50 | 189.4800 | 30.79109 | 4.35452 |

p -Value : 0.044

p-Value less than 0.044 indicates the difference in TC across Case and Control groups is significant (Significant at 5%). The Group statistics table shows Case group is associated with higher TC than controls.

Comparison of FBS (T test)

Table - 13

| Group | | N | Mean | Std. Deviation | Std.Error Mean | |
|-------|---------------|----|----------|----------------|----------------|--|
| FBS | Case Group | 50 | 100.5600 | 6.07827 | .85960 | |
| | Control Group | 50 | 91.7800 | 8.62907 | 1.22033 | |

P value < 0.01

P-Value less than 0.01 indicates the difference in FBS across Case and Control groups is highly significant (Significant at 1%); The Group statistics table shows Case group is associated with higher FBS than control group though it was lower than in diabetics.

Comparison of SI (T test)

Table – 14

GROUP STATISTICS

| Group | | Ν | Mean | Std. Deviation | Std.Error Mean | |
|-------|----|---------------|------|----------------|----------------|--------|
| ľ | SI | Case Group | 50 | 21.6040 | 6.23892 | .88232 |
| | | Control Group | 50 | 11.4140 | 2.49399 | .35270 |

p -Value :< 0.01

p-Value less than 0.01 indicates the difference in SI across Case and Control groups is highly significant (Significant at 1%). The Group statistics table shows Case group is associated with higher SI than control group.

Comparison of HOMA – IR (T test)

Table - 15

| Group | N | Mean | Std. Deviation | Std.Error Mean |
|--------------------|----|--------|----------------|----------------|
| HOMA-IR Case Group | 50 | 5.3952 | 1.75469 | .24815 |
| Control Group | 50 | 2.6974 | .55524 | 0.7852 |

p value : <0.01

p-Value less than 0.01 indicates the difference in HOMA-IR across Case and Control groups is highly significant (Significant at 1%). The Group statistics table shows Case group is associated with higher HOMA-IR than control group.

Categorical Variables Analysed By Chi Square Test

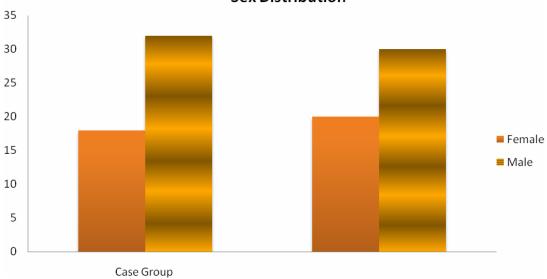
| Table - | 16 |
|---------|----|
|---------|----|

| | | | Total | |
|--------|----------------|------------------|------------|--------|
| | | Control Group | Case Group | 1 otur |
| Gender | Female Count | 18 | 20 | 38 |
| | Expected count | 19.0 | 19.0 | 38.0 |
| | Male Count | 32 | 30 | 62 |
| | Expected count | 31.0 | 31.0 | 62.0 |
| Total | Count | 50 | 50 | 100 |
| | Expected Count | 50.0 | 50.30 | 100.0 |

Table - 17

| | Value | dt | Asymp. Sig (2 sided) | Exact sig (2sdides) | Extract sig (1sides) |
|-----------------------|-------|----|--------------------------|----------------------|-------------------------|
| Pearson chi-square | .170 | 1 | .680 | | |
| Continuity correction | .042 | 1 | .837 | | |
| Likelihood Ratio | .170 | 1 | .680 | | |
| Fihser's exact test | | | | .837 | .418 |
| Linear- by – Linear | .168 | 1 | .682 | | |
| association | | | | | |
| N of valid cases | 100 | | | | |

Based on Pearson Chi-Square test, p-Value of 0.680 indicates the difference in gender across case and control groups is statistically not Significant



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Sex Distribution

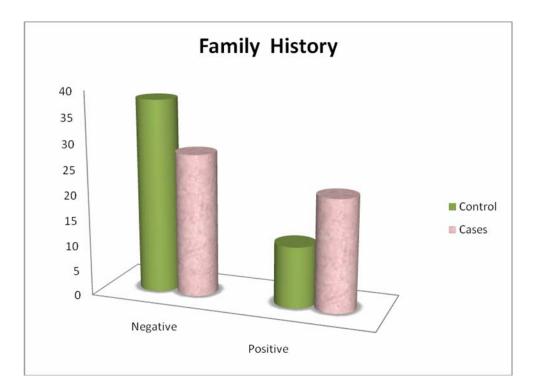
| Table - | - 18 |
|---------|------|
|---------|------|

| | Gro | Group | | |
|-------------------------|------------------|---------------|-------|--|
| | Control Group | Case Group | Total | |
| FAM-HIST Negative Count | 38 | 28 | 66 | |
| Expected Cour | it 33.0 | 33.0 | 66.0 | |
| Positive Count | 12 | 22 | 34 | |
| Expected Cour | nt 17.0 | 17.0 | 34.0 | |
| Total Count | 50 | 50 | 100 | |
| Expected Cour | nt 50.0 | 50.0 | 100.0 | |

Table - 19

| | Valu | Dt. | Asymp. Sig | Exact sig | Extract sig |
|-----------------------|-------|-----|------------|-----------|-------------|
| | e | | (2 sided) | (2sdides) | (1sides) |
| Pearson chi-square | 4.456 | 1 | .035 | | |
| Continuity correction | 3.610 | 1 | .057 | | |
| Likelihood Ratio | 4.506 | 1 | .034 | | |
| Fihser's exact test | | | | .057 | .028 |
| Linear- by – Linear | 4.412 | 1 | .036 | | |
| association | | | | | |
| N of valid cases | 100 | | | | |

Based on Pearson Chi-Square test, p-Value of 0.035 indicates the difference in distribution of Family History across Case and control groups is statistically significant (at 5%), from the chart of distribution we can conclude that the case group has higher cases of positive family history than controls.



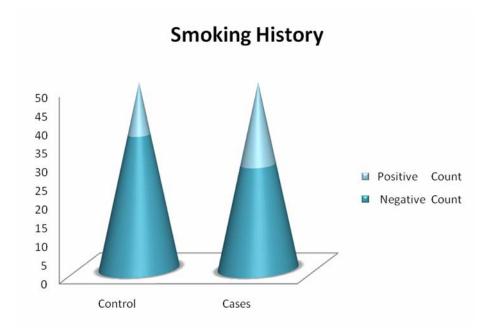
| Table | - 20 |
|-------|------|
|-------|------|

| | | Gro | Group | |
|----------------|----------------|------------------|---------------|-------|
| | | Control Group | Case Group | Total |
| SMOKING Negati | ve Count | 36 | 28 | 64 |
| | Expected Count | 32.0 | 32.0 | 64.0 |
| Positive | e Count | 14 | 22 | 36 |
| | Expected Count | 18.0 | 18.0 | 36.0 |
| Total | Count | 50 | 50 | 100 |
| | Expected Count | 50.0 | 50.30 | 100.0 |

Table - 21

| | Value | dt | Asymp. Sig (2 sided) | Exact sig (2sdides) | Extract sig (1sides) |
|-----------------------|-------|----|--------------------------|----------------------|----------------------|
| Pearson chi-square | 2.778 | 1 | .096 | | |
| Continuity Correction | 23127 | 1 | .145 | | |
| Likelihood Ratio | 2.795 | 1 | .095 | | |
| Fisher's Exact Test | | | | .144 | .072 |
| Linear- by – Linear | 2.750 | 1 | .097 | | |
| association | | | | | |
| N of valid cases | 100 | | | | |

Based on Pearson Chi-Square test, p-Value of 0.096 indicates the difference in smoking across case and control groups is statistically not significant.



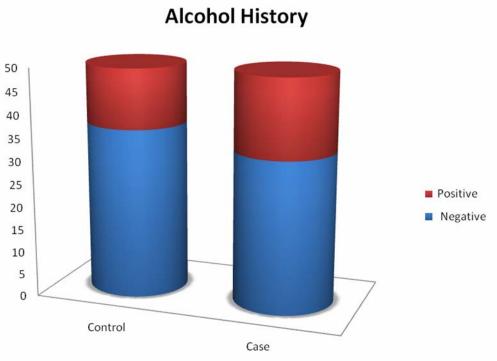
| Table - | 22 |
|---------|----|
|---------|----|

| | | Group | | | |
|-----------------|----------------|------------------|---------------|-------|--|
| | | Control Group | Case Group | Total | |
| ALCOHOL Negativ | e Count | 37 | 33 | 70 | |
| | Expected Count | 35.0 | 35.0 | 70.0 | |
| Positive | Count | 13 | 17 | 30 | |
| | Expected Count | 15.0 | 15.0 | 30.0 | |
| Total | Count | 50 | 50 | 100 | |
| | Expected Count | 50.0 | 50.30 | 100.0 | |

Table - 21

| | Value | dt | Asymp. Sig (2 sided) | Exact sig (2sdides) | Extract sig (1sides) |
|-----------------------|-------------------|----|--------------------------|----------------------|----------------------|
| Pearson Chi-square | .762 ^b | 1 | .383 | | |
| Continuity Correction | .429 | 1 | .513 | | |
| Likelihood Ratio | .764 | 1 | .382 | | |
| Fisher's exact test | | | | .513 | .257 |
| Linear- by - Linear | .754 | 1 | .385 | | |
| association | | | | | |
| N of valid cases | 100 | | | | |

Based on Pearson Chi – Square test, p- Value of 0.383 indicates alcohol consumption do not differ across case and control group.



Multivariate Analysis – Multiple Linear Regression

A multiple linear regression analysis was performed to study the association between coronary artery disease case group and HOMA-IR by controlling for all other factors.

Table - 24

| | Unstandardized Coefficients | | Standardized coefficients | | <i>c</i> : |
|-------------|--------------------------------|---------------|---------------------------|--------|------------|
| Model | В | Std. Error | Beta | t | Sig. |
| 1 constant) | -7.941 | 1.984 | | -4.003 | .000 |
| AGE | .032 | .018 | .111 | 1.795 | .076 |
| GENDER | 246 | .258 | 064 | 954 | .343 |
| FAM-HIST | 073 | .235 | 018 | 309 | .758 |
| SMOKING | .122 | .257 | .031 | .473 | .637 |
| ALCOHOL | .004 | 262. | .001 | .015 | .988 |
| BMI | .002 | .005 | .025 | 406 | .686 |
| WHR | 3.077 | 1.398 | 0.162 | 2.201 | .030 |
| SBP | .005 | .008 | 0.37 | 609 | .544 |
| HDL | 058 | .034 | 180 | -1.695 | .094 |
| LDL | .021 | .010 | .188 | 2.128 | .036 |
| TGL | .006 | .008 | .088 | .670 | .504 |
| ТС | 005 | .005 | 107 | -1.070 | .287 |
| FBS | .074 | .016 | .340 | 4.511 | .000 |
| GROUP | 1.092 | .465 | .293 | 2.348 | .021 |

COEFFICIENTS

a. Dependent Variable: HOMA -IR

Variable named as "Group" is the indicator for coronary artery disease. Coding of Group is given by Group = 1 for case Group and Group = 0 for control group.

Based on the multivariate controlled study, after controlling for all other factors the difference in HOMA-IR between the case and control group is significant. Based on the p-value of 0.021 for Group variable and a positive regression coefficients of 1.092 suggests that we can conclude at 5% level of significance that the Coronary artery disease (case group) is associated with higher HOMA-IR and the case group has on an average 1.092 units of HOMA-IR higher than the control group.

DISCUSSION

DISCUSSION

Insulin resistance plays an important role in promoting CAD and the degree of insulin resistance correlates with the severity of CAD. (81). Insulin resistance has been considered to promote atherosclerosis by directly affecting blood vessels. The relationship between insulin resistance and atherosclerosis has been studied from various aspects. Hyperinsulinemia is an only indirect indicator of insulin sensitivity. Several large scale studies have revealed that hyperinsulinemia is closely associated with the mortality due to cardiovascular disease (82).

To date, however, few large scale studies have been conducted to examine the relationship between insulin resistance and coronary heart disease. Recently, IRAS (Insulin Resistance Atherosclerosis study) group has conducted large-scale epidemiological studies, and has reported that insulin resistance rather than insulin concentration is an independent powerful risk factor for coronary heart disease.(83).

There are different theories as to how insulin resistance affects coronary artery disease (84). Insulin resistance is related to the vasomotor dysfunction of the endothelial cells that line the coronary artery (85). It is well known that endothelial function is impaired in patients with coronary artery disease. In our study, we evaluated the effect of impaired glucose metabolism and insulin resistance that may play some role in the pathophysiology of CAD. We found increased fasting insulin levels in patients with CAD. Increased insulin resistance may play at least some role in patients with CAD.

Recent evidences indicate that insulin resistance is linked to untraditional CAD risk factors and possibly pro-atherosclerotic inflammatory state. Study conducted by Prof. Sidhartha Das and colleagues in angiographically established patients with CAD revealed the prevalence of IR in 40% and hyperinsulinemia in 50% of subjects having angiographically proven IHD who were normoglycemic.

Risk factors for CAD in Angiographically proved cases with IHD

| Obesity (WHR) | 100% |
|-----------------------------------|------|
| BMI | 40% |
| HTN | 60% |
| TC > 200 | 20% |
| LDL > 100 | 70% |
| TGL > 150 | 40% |
| HDL < 40 | 60% |
| Smoking | 20% |
| Insulin resistance (HOMA IR >2.5) | 40% |

| Mircoalbuminuria | 20% |
|------------------------------|-----|
| Hyper insulinemia (>9 µu/ml) | 50% |

In another study, Yazici et al (86) investigated the relationship between the degree of slow coronary flow (SCF) and serum insulin, glucose and lipid levels. As a result, they found no correlations between corrected TIMI frame count and serum insulin, glucose and lipid levels. However they measured only basal serum insulin, not insulin resistance, without homeostatic models. On the other hand, evaluating insulin resistance by homeostatic modelling is more reliable than the standard method because both fasting glucose and insulin levels are integrated (87).

According to Take zako et al; the profile of insulin resistance based on HOMA – IR model was correlated with severity of coronary atherosclerosis based on Gensini's score (88).

Other studies have reported that the incidence of major adverse cardiac events (MACE) following Percutaneous Coronary Intervention was higher in non –diabetic patients with high concentration of HbA₁C than those with low concentration of HbA₁C although these studies did not evaluate using insulin resistance (89). Here, the concentration of HbA₁C was lower in non-diabetic patients than in diabetic patients although it was high. Some studies have reported that hyperinsulinemia and insulin resistance measured by HOMA are closely associated with restenosis following stenting in non diabetic patients (90, 91). A study from Finland found a significantly increased risk of ischemic loop to loop pdf disease in individuals who had hyperinsulinemia.

The Rancho-Bernardo study found no association between insulin levels and cardiovascular disease in women and an inverse association between post challenge insulin levels and cardiovascular disease in men (higher insulin levels were protective). Likewise, no association was found in the 3 studies that presented data on men and women combined; the study from Finland, the study of Pima Indians, and the Sand Luis valley study. It is possible that some of these differences reflect ethnic or geographic variation between the studies.

Another preliminary study from North India by Rajiv Mahajan & colleagues showed no significant association between fasting insulin and CAD. Similarly there was no significant relationship between HOMA – IR and CAD. However in this study, in addition to excluding diabetes, they excluded hypertension and obesity (BMI>25), as both are also independently associated with hyperinsulinemia. They did not find a significant association between hyperinsulinema and CAD.

SUMMARY

SUMMARY

Age and sex distribution between the two groups (cases and controls) were similar.

There was a significant difference in the distribution of family history across cases and control group suggesting a higher prevalence of family history of coronary artery disease in cases.

There was no difference in the prevalence of smoking between the cases and controls group.

Although there was no difference in body mass index (BMI) between the two groups, waist to hip circumference ratio was higher in case group than in control group suggesting higher prevalence of central obesity in cases with coronary artery disease.

Case group had on an average high total cholesterol and triglyceride and lower HDL cholesterol which was statistically significant than control groups. Though LDL was higher in cases it was not statistically significant.

Fasting Blood sugar was significantly higher in cases than in controls although it was lower than in patients with diabetes.

Fasting insulin and Insulin Resistance (HOMA –IR) were significantly higher in cases than in controls.

Although family history, HDL, TGL, TC, FBS attained significance in univariate analysis, only WHR, LDL cholesterol and fasting blood sugar were significantly correlated with insulin resistance in multivariate analysis.

CONCLUSION

CONCLUSION

Thus fasting insulin levels and insulin resistance measured by homeostatic model assessment (HOMA-IR) are higher in patients with coronary artery disease compared to controls.

The mechanism by which insulin resistance provokes cardiovascular disease is mainly associated with the development of metabolic syndrome.

Insulin resistance is significantly associated with cardiovascular disease risk, even in the absence of diabetes, probably insulin resistance alone without diabetes can be considered as a coronary heart disease – risk equivalent in future.

Our study proves that non-diabetic patients with insulin resistance exhibit high concentration of serum triglyceride and low concentration of HDL cholesterol. The present study has shown that the value of HOMA- IR was correlated with levels of triglyceride and inversely correlated with that of HDL cholesterol

LIMITATIONS OF THE STUDY

LIMITATIONS OF THE STUDY

The limitation of the present study is that it examined only a small number of patients. To date only few studies have been conducted to examine whether insulin resistance is correlated with coronary artery disease. Thus it can be taken as a preliminary study for further large scale studies.

The present study failed to testify the reproducibility since it did not measure the value of HOMA –IR in a repetitive manner. The concentration of serum glucose and insulin can be altered at each different measuring time, although the present study measured them only once. However HOMA-IR index is well reflected in euglycemic hyperinsulinemic clamp test, a standard test and the clinical usefulness has been well established.

BIBLIOGRAPHY

BIBLIOGRAPHY

- Enas EA, Yusuf S, Mehta JL. Prevalence of coronary artery disease in Asian Indians Am. J.Cardiol 1992; 70: 945-949.
- Jha P, Enas E, Yusuf S.Coronary artery disease in Asian Indians: Prevalence and Risk factors. Am Pac. Isl J Health 1993;1:163-175.
- Enas EA, Mehta J. Malignant Coronary artery disease in young Asian Indians thoughts on pathogenesis, risk factors of coronary artery disease Am.J.Cardiol 1994;4:174-178.
- Mc Keigue PM 1996 Metabolic consequences of obesity and body fat pattern: lessons from migrants studies, Ciba Found symp 201:54-64; discussion 64-67, 188-193.
- MC keigu PM, Shah B,Marmot MG 1991 Relation of central obesity and insulin resistance with high diabetes prevalence and cardiovascular risk in south Asians. Lancet 337:382-386.
- 6. Nakae J, Kido Y and Accili D (2001) Distinct and overlapping functions of Insulin and IGF –I receptors Endocrine Rev 22, 818-835
- White , M.F. (2002) IRS proteins and the common path to diabetes Am J physiol Endocrinol metab 283, E413-422.
- 8. Van obberghen E Baron V Delahyes L Emanceelli B, Filippa, N Giorgetti- Peraldi S., Lebrun I., Mothe-Satney I., Peraldi P Rocchi

۰.

S,Saioka- Verhelle D., Tartare- Deckert S-and Guidicett.J J.(2001) Surfing the insulin signaling web. Eur J clin Invest 31, 966-977.

- Siddle K., Urso. B., Niesler A., Cope, D.L., Molina. L., Suriyana, K.H. and Soos., M.A., (2001) Specificity in Ligand binding and intracellular signalling insulin and insulin like growth factor receptors. Biochem sec Trans 29, 513-525.
- Saltiel A.R and kahn C.R (2001) insulin signaling and the regulation of glucose and lipid metabolism Nature 414,799-806.
- 11.Reaven G.M (1988). Role of insulin resistance in human disease.Diabetes 37, 1495-1507.
- 12.Kahn CR insulin action, diabetogens , and the cause of type 2 diabetes Diabetes 1994;43:1066-1084.
- 13.Banerji MA., Faridi N, Atlun R, Chaiken RL, Lebovitz H.E 1999.Body composition, Visceral fat, Leptin and insulin resistance in Asian Indians men J.Clin Endoorinol. Metab 84:137-144.
- 14.Raji A, Seely EW, Arky RA, Simonson DC 2001 Body fat distribution and insulin resistance in healthy Asian Indian and Caucasians J.Clin Endocrinol Metab 86: 5366-5371.
- 15.Ramachandran A, Snehalatha C, Latha E, Satyavani K, Vijay V 1998. Clustering of cardiovascular risk factors in urban Asian indians Diabetes care 21:967-971.

- 16. MC keigue PM, Faridi N, Siddle K, 1990 Relation of central obesity and insulin Resistance with high diabetes prevalence and cardiovascular risk in south Asian Lancet – 337-382-386.
- 17.Dcrose GK, Zimmet PZ, Alberti KG, Brighan L, Carlin JB, Tuomilehto J, Knight LT, Gareeboo H 1993. Serum insulin distribution and reproducibility of the relationship between 2 hour insulin and plasma glucose levels in Asian Indian, Creole and Chinese Mauritians Mauritius NCD study group. Metabolism 42:1232-1241.
- 18.Omar MA, Seedat MA, Dyer RB, Motala AA, knight LT, Becker PJ. 1994.South African Indians shows a high prevalence of NIDDM and bimodality in Plasma glucose distribution patterns. Diabetes care 17:70-73.
- 19.Snehalatha C, Ramachandran A, satyavani K, vallabi MY, Viswanathan V1997 Computed axial tomographical scan measurement of abdominal fat distribution and its correlation with anthropometry and insulin secretion in healthy Asian Indians. Metabolism 46:1220-1224.
- 20. Feskens E.J.M., Tuomileh to, J., Stengaard J.H., Pekkanen J., Nissinen, A. and kromhout D (1995) Hypertension and overweight associated with hyperinsulinemia and glucose tolerance: a

longituditinal study of the Finnish and Dutch cohorts of the seven countries study. Diabetologia 38,839-847.

- 21.Brands M.W., Hildebrandt D.A., Mizelle H.L and Mall J.E (1991) Sustained hyperinsulinemia increase arterial pressure in conscious rats. Am. J.Physiol 260, R 764-768.
- 22. Haffner S.M., Valde R.A., Hazuda HP., Mitchell B.D., Morales P.A and stern M.P(1992) . Prospective analysis of insulin Resistance syndrome (syndrome X). Diabetes 41, 715-722.
- 23.Despres J.P., Lamarche, B and Mauriege, Petal (1996)Hyperinsulinemia as an independent risk factor for ischemic heart disease, NEJM 334, 952-957.
- 24.Beatty O.L., Harper R., Sheriden B., Atkinsion A.B. and Bell P.M(1993) Insulin resistance in offspring of hypertensive parents BMJ 307, 92-96.
- 25.Steinberg H.O., Brechtel G. Insulin mediated muscle vasodilation is nitic oxide dependent. A novel action of insulin to increase nitric oxide release. J clin invest 1994; 94:1172-79.
- 26.De fron Zo R.A., Cooke C.R. The effect of insulin on renal handling of Na. K, ca and phosphate in man . J.clin invest 1975;55 845-55

- 27.Barbato , A, Folker, EJ., et al Metabolic syndrome and renal sodium handling in three ethnic groups living in England Diabetologia 2004;47:40-46.
- 28.Tooke J.E., Hannemann, M.M. Adverse endothelial function and the insulin resistance syndrome J. intern Med 2000; 247:425-31
- 29.Kuroda, S, Fuji T, et al. Role of Insulin Resistance in the genesis of sodium sensitivity in essential hypertension, J.Hum Hypertens 1999, 13:257-62.
- 30.Baron , AD.1994 Hemodynamic actions of insulin Am.J. Physiol 267;E187- E 202
- 31.Anderson, E.A, Hoffman R.P., Mark A.L HyperInsulinemia produces both sympathetic neural activation and vasodilatation in normal humans J.Clin Invest 1991; 87;2246-52
- 32.Egan, B M., Insulin resistance and the sympathetic nervous system curr hypertens Rep 2002, 5: 247-54
- 33.Mc. Keigue PM 1996. Metabolic consequences of obesity and body fat pattern: lessons from migrant studies Ciba Found Symp 201:54-64: discussion 64-67, 188-193.
- 34. Mohan V, Deepa R, Haranath S.P, Premalatha G, Rema M, Sastry N.G., Enas E.A. 1998 Lipoprotein (a) is an independent risk factor for

۰.

coronary artery disease in NIDDM patients in South India. Diabetes care 21:1819-1823.

- 35.Lewis G.F., Uffelman K.D. Interaction between FFA and insulin in acute control of VLDL production in human J. Clin invest 1995;95: 158-66.
- 36.Lewis G.F, Steiner G.Acute effects of insulin in the control of VLDL production in humans. Diabetes care 1996; 19: 390-93.
- 37. Eckel, R.M., Yost T.J., Alteration in lipoprotein lipase in insulin resistance. Int J.obes Relat Met disorder 1995; 19 (suppl); S 16-S21
- 38. Murakami T, Longhi R, et al Triglycerides are major determinants of cholesterol esterification / transfer and HDL remodelling in human plasma Arterioscle Thromb vase Biol 1995; 15:1819-28.
- 39.Brinton EA, Eisenberg Increased apo A1 and A2 fractional catabolic rate in patients with low HDL levels with or without hypertriglyceidemia J Clin invest 1991; 87:536-44.
- 40.Manzato E, Zambon Level and physiochemical properties of lipoprotein subclass in moderate hypertriglyceridemia Clin Acta 1993, 219:57-65.
- Kwiterovich Pojr. Clinical relevance of the biochemical, metabolic and genetic factors that influence LDL heterogenecity Am.J. cardiol 2002; 90; 30i-47i

- 42. Packard C.J. LDL subfractions and atherogenecity an hypothesis from the university of glascow Curr med. Opin 1996; 13:379-90
- 43.Abdel Maksoud M.F., H.O Kanson J.E. The complex role of triglycerides in cardiovascular disease Semin vase Med 2003; 2: 325-34.
- 44.Dudya V, Misra A, Pandey RM, Devina G., kumar G, vikram N.K.2001 BMI does not accurately predict overweight in Asian Indians in northern India Br, J. Nutr. 86:105-112.
- 45. Misra A 2003 Body composition and the metabolic syndrome inAsian Indians: A saga of multiple adversities Natl med. J. India. 16:3-7
- 46.WHO. Western pacific region. The Asia pacific perspective Redefining obesity and its treatment WHO/IASD/IVIF, 2000.
- 47.Lee S, Ross R. Inter individual variation in abdominal subcutaneous and visceral adipose tissue: influence of measurement site. J Appl physiology 2004; 97: 948-54.
- 48.Aubest H, Alessi MC. Weak and non independent association between plasma TAF I antigen levels and the insulin resistance syndrome J.Thromb Haemost 2003; 1: 791 – 97.

- 49.Bajaj M, Banerji M.A. Type 2 Diabetes in south Asians; a pathophysiological focus on the Asian Indians epidemic. Curr Diab Rep 2 004; 4:231-218.
- 50.Tanaka S, Horimai C.Ethnic differences in abdominal visceral fat accumulation between Japanese, African Americans and Caucasians : a meta analysis Acta Diabetol 2003; 40 (Suppl) S 302- S 304.
- 51.Guo Z, Jensen M.D, Johnson C.M. Regional post prandial FFA metabolism in different obesity phenotypes. Diabetes 1999; 48: 1586-92.
- 52.Randle P.J., Hales C.N. Garland P.B and Newson E.A (1963) The glucose fatty acid cycle, its role in insulin sensitivity and metabolic disturbances of diabetes mellitus Lancet 1, 785-789.
- 53.Griffin M.E., Marcucci M.J., Cline G.W., Bell K. Barucci, N., Lee D Good year, L.J., Kraegen, E.W., white M.F., and shulman G.I (1999)
 .Free fatty acid induced insulin resitance is associated with activation of protein kinase C theta and alterations in the insulin signalling cascade, Diabetes 48; 1270-74.
- 54.Haffner S.M., Lehto S., Ronnemaa T., Pyorala K.and Laasko M (1998) Morlality from coronary heart disease in subject with type 2 diabetes and in non – diabetic subjects with and without prior myocardial infarction. NEJM 339: 229-234.

- 55. Bokemark L, Wikstrand J., Attavalli S, Huthe J., Wedel H.and Fagerberg B (2001) Insulin resistance and intima - media thickness in carotid and femoral arteries of clinically healthy 58 years old men. The atherosclerosis and insulin resistance (AIR) study J. Intern Med 249, 59-67.
- 56.Howard G, O' Leary D.H., Zaccaroo D, Rewers H, Hamman R, Selby J.V Saad M.F., Savage P and Bergman (1996) Insulin sensitivity and arthrosclerosis. Circulation 93, 1809-1817
- 57.Sandholzer C, Hallman DM, Saha N, Sigurdsson G, Lackner C, Csanzar A, Boerwinkle E, Utermann G (1991) Effects of the apolipoprotein (a) Size, polymorphism on the lipoprotein (a) concentration in 7 ethnic groups. Hum Genet 86: 607-614.
- 58.Markovitz J.H. Kulkarni K, Goldschmidt dermont P, kiefe C.I, Rustagi P, Sekar P, Nanda N. 1998 Increased platelet activation and fibrinogen in Asian Indians. Potential implications for coronary risk . Eur Heart J19 720-726.
- 59.Nagi D.K, Knowler W.C, Hanson R.L., Ali V.M 1996 Plasminogen activator inhibitor (PAI-1) and non insulin dependent diabetes in Pima Indians, south Asians and Europeans Thromb Haemost 75: 921-927.

- 60.Chan .J.M., Rimm E.B., Colditz G.A., stampfer M. J., and Willett.W.C (1994). Obesity, fat distribution and weight gain as risk factors for clinical diabetes in men. Diabetes care 961-969.
- 61.Marshall J.A., Bessesen D.H. and Hamman RF (1997) High saturated fat and low starch and fibre are associated with hyperinsulinemia in a non diabetic population, the san luis valley diabetes study, Diabetologia 40: 430-438.
- 62.Vessb B, Unsitupa M, Hermansen K, Riccardi G, Rivellese A.A and Tapsell L.C et al., (2001) Substituting dietary saturated for monounsaturated fat impairs insulin sensitivity in healthy men and women; the KANWU study Diabetologia 44:B12.319.
- 63.Mc Keown N.M, Meigs JB, Lui S Wison P.W.F and Jacque P.F (2002) whole grain Intake is favourably associated with metabolic risk factors for type 2 diabetes and cardiovascular disease in the Framingham offspring study Am. J.Clin . Nutr 76: 390-398.
- 64.Wood P.O, Stefanick M.L, Williams P.T and Haskell W.L (1991).The effect on plasma lipoproteins of a prudent weight reducing diet, with or without exercise, in overweight men and women NEJM 325: 461-466.

- 65.Kroteivski M, Handroukas L and Sjostrom L et al (1970) Effect of long term physical training on body fat, metabolism and blood pressure in obesity Metabolism 28: 650-658.
- 66.Helmrick S.P, Ragland D.R., Leung R.W and Paffenbarger R.S (1991) Physical activity and reduced occurrence of non insulin dependent diabetes mellitus NEJM 325 147-152.
- 67.Hemibrecht R, Wolf A and Giefen S. et al (2000) Effect of exercise on coronary endothelial function in patients with coronary artery disease, NEJM 342 454-460.
- Randeva H.S., Lewandowski K.C., Drzowoski J, Brooke-wavelle K,
 O' Callaghan C, Hillhouse E.W and Prelevic G.H (2002) Exercise decreases plasma total homocysteine in overweight young women with polycystic ovary syndrome . J .Clin Endocrin Metab 87: 4496-4501.
- 69.Stratton J.R., Chandler W.L and Schwart z R.S et al (1991) Effects of physical conditioning on fibrinolytic variables and fibrinogen in young and old healthy adults Circulation 83:16
- 70.Rad Ziuk J. (2000) Insulin sensitivity and its Measurement: Structural commonalities among the methods J, Clin Endocrinol metab 85:4426-4433.

۰.

- 71.Wallace T.M and Mathews D.R (2002) The assessment of insulin resistance in man. Diabet Med 19, 527-534.
- 72.Bergman R.N, Finegood D.T and Andres R (1985) Assessment of insulin sensitivity in vivo Endocrine Rev 6:45 -86.
- 73. De Fronzo R.A., Tobin J.D and Ander M (1979) Glucose clamp technique ; a method for quantifying insulin secretion and resistance.
 Am J physiol 237 : E 214 E 223.
- 74.Kelley D.E and Mandarino L.J (2000) Fuel selection in human skeletal muscle in Inuslin resistance a re examination. Diabetes 49: 677-683.
- 75.Blaak E.E., Wagen Makers A.J., Glatz J.F Wolffenbuttfel B.H, Kemerink G.J., Langenberg C.J. Heidendal G.A and suris W.H (2000) plasma FFA uitilisation and fatty acid binding protein content are diminished in type 2 diabetic muscle .Am J physiol Endocrinol metab 279 E 146-E154.
- 76.Bergam R.N, Ider Y.Z, Bowden C.R and Cobelli (1979) Quantitative estimation of insulin sensitivity Am. J physiol 236; E667-E677.
- 77.Weber K.M, Martin I.K, Best J.D, Alford F.P and Boston R.C (1989) Alternative method for minimal model analysis of intravenous giucose tolerance data Am J. Physiol 256, E524-535.

- 78.Matthews D.R, Hosker J.P, Rudenski A.S, Naylor B.A, Teacher D.F and Turner R.C (1985) Homeostasis model assessment: Insulin resistance and beta cell function from fasting plasma glucose and Insulin concentration in man. Dialetologia 28; 412-419.
- 79.Katz A, Nambi S.S., Mather K, Baron A.D., Follman DA Sullivan G and Quoton M.J (2000) Quantitative insulin sensitivity check index: a simple, accurate method for assessing insulin sensitivity in humans . J.din endocrinol Metab 85: 2402-2410.
- 80.Expert committee on the diagnosis and classification of diabetes mellitus Report of the expert committee on the diagnosis and classification of diabetes mellitus. Diabetes care 1997; 20:1183-97.
- 81.Pyorala M,Miettinen H, Laaskso M.Pyorala K.Plama insulin and all cause, cardiovascular and non cardiovascular mortality; the 22 year follow up result of the Helsinki Policeman study. Diabetes care 2000; 23:1097-1020
- 82.Despres J.P, Lamarche B, Mauriege P, Cantin B Hyperinsulinemia as an independent risk factor for ischemic heart disease. NEJM 1996.
 334: 952 – 957
- 83.Rewers M, Zaccaro D, D' Agostino R, Haffner S, Selby J.V Insulin resistance Atherosclerosis study investigator. Insulin sensitivity,

insulinemia and coronary artery disease: the Insulin Resistance Atherosclerosis study. Diabetes care 2004, 27:781-787

- 84.Mansfield M.W., Heywood DM, Grant P.J. Circulating levels of factor VII fibrinogen, Von willebrand factor and features of insulin resistance in first degree relatives of patients with NIDDM Circulation 1996; 94: 2171-2176.
- 85.Inoue T, Matsunaga R, Sakai Y, Yaguchi I, Takaya nagi K, Marooka S. Insulin resistance affects endothelium dependent acetyl choline induced coronary artery response. Eur Heart J 2000; 21:895-900.
- 86.Yazici M, Demircan S, Aksakal E,Sahin H, Meric M, Dursun I et al. Plasma insulin, glucose and lipid count in patients with slow coronary flow, Turkish Andolu kardiyol Derg 2003;3:227-229.
- 87.Ugur Attun B, Altun A, Tatli E, Arikan E Relationship between Insulin Resistance assessed by HOMA- IR and exercise test varirables in asymptomatic middle aged patients with type diabetes J.Endocrinol invest 2004 ; 27:455-61.
- 88.Takezako T, Saku K, Zhang B, Shiraik , Arakawa K. Insulin resistance and angiographical characteristics of coronary atherosclerosis Jpn Circ. J. 1999, 63:666-676

۰.

- 89. Radke P.W., VoswinkelM, Reith M, Kaiser A, Haager PK, Hanrath P, Hoffman R.Relation of fasting Insulin plasma levels to restenosis after elective coronary stent implantation in patients without diabetes mellitus. Am J.Cardiol 2004; 93: 639-641.
- 90. Corpus R.A, O' Neill W.N, Dixon SR, Timmis G.C, Delvin W.H. Relation of HbA₁C to rate of major adverse cardiac events in non diabetic patients undergoing percutaneous coronary revascularization Am J cardiol 2003:92:1282-1286.
- 91.Sekiguchi M. Kurabayashi M, Adachi M, Oshima S Usefulness of insulin resistance measured by homeostais model assessment in predicting restenosis after coronary stent placement in non diabetic patients. Am. J cardiol 2004-93: 920-922.

ABBREVIATION

ABBREVIATION

| CAD | - | CORONARY ARTERY DISEASE |
|---------------------------------------|-----------------------|--|
| BMI | _ | BODY MASS INDEX |
| WHR | _ | WAIST HIP RATIO |
| SBP | _ | SYSTOLIC BLOOD PRESSURE |
| DBP | _ | DIASTOLIC BLOOD PRESSURE |
| HDL | _ | HIGH DENSITY LIPOPROTEIN |
| LDL | _ | LOW DENSITY LIPOPROTEIN |
| TGL | _ | TRIGLYCERIDES |
| ТС | _ | TOTAL CHOLESTEROL |
| FBS | _ | FASTING BLOOD SUGAR |
| | | |
| SI | _ | SERUM INSULIN |
| SI HOMA–IR | _ | |
| | _ | |
| | _ | HOMEOSTATIC MODEL ASSESSMENT – |
| HOMA–IR | | HOMEOSTATIC MODEL ASSESSMENT – INSULIN RESISTANCE |
| HOMA–IR IRS | _ _ _ _ | HOMEOSTATIC MODEL ASSESSMENT – INSULIN RESISTANCE INSULIN RECEPTOR SUBSTRATE |
| HOMA–IR IRS IR | | HOMEOSTATIC MODEL ASSESSMENT – INSULIN RESISTANCE INSULIN RECEPTOR SUBSTRATE INSULIN RESISTANCE |
| HOMA–IR IRS IR VLDL | | HOMEOSTATIC MODEL ASSESSMENT – INSULIN RESISTANCE INSULIN RECEPTOR SUBSTRATE INSULIN RESISTANCE VERY LOW DENSITY LIPOPROTEIN |
| HOMA–IR IRS IR VLDL APO B | - - - - - | HOMEOSTATIC MODEL ASSESSMENT – INSULIN RESISTANCE INSULIN RECEPTOR SUBSTRATE INSULIN RESISTANCE VERY LOW DENSITY LIPOPROTEIN APOLIPOPROTEIN B |

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TNF- ALPHA –TUMOUR NECROSIS FACTOR ALPHA

- LPL LIPOPROTEIN LIPASE
- PAI-1 PLASMINOGEN ACTIVATOR INHIBITOR -1
- HSL HORMONE SENSITIVE LIPASE
- GLUT -4 GLUCOSE TRANSPORTER -4
- PCOS POLYCYSTIC OVARIAN SYNDROME
- OGTT ORAL GLUCOSE TOLERANCE TEST
- QUICKI QUANTITATIVE INSULIN SENSITIVITY CHECK INDEX
- G/IRATIO GLUCOSE /INSULIN RATIO
- STEMI ST ELEVATION MYOCARDIAL INFARCTION
- NSTEMI NON ST ELEVATION MYOCARDIAL

INFARCTION

- ACS ACUTE CORONARY SYNDROME
- RVMI RIGHT VENTRICULAR MYOCARDIAL INFARCITON
- ECG ELECTROCARDIOGRAM

- IRAS INSULIN RESISTANCE ATHEROSCLEROSIS STUDY
- TIMI THROMBOLYSIS IN MYOCARDIAL INFARCTION
- MACE MAJOR ADVERSE CARDIAC EVENTS.

PROFOMA

PROFOMA

History

| Name | Age | Sex |
|--------------------|-------------|--------|
| Occupation | op.no | |
| Address | | |
| Diabetes : Y/N | Hypertensio | n: Y/N |
| H/o Stable angina: | | |
| Smoker : | | |
| alcoholic: | | |
| | | |

Family H/O DM/HT/CAD:

Examination :

| Height : | Weight: |
|----------|---------|
| BMI : | WHR: |
| Pulse : | BP : |
| CVS : | RS: |
| Abd : | CNS: |

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Investigations

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| Blood sugar | Fasting : | PP: | | | | | | | | |
|--------------|-----------|------------------|------------|--|--|--|--|--|--|--|
| Blood Urea | : | Serum creatinine | | | | | | | | |
| Urine Alb : | | Sugar : | deposits : | | | | | | | |
| Lipid Profil | e : | | | | | | | | | |
| TC : | TGL : | HDL: | LDL : | | | | | | | |
| ECG : | | Echocardiogram: | | | | | | | | |
| Fasting insu | ılin: | HOMA –IR : | | | | | | | | |

MASTER CHART

| SL.NO | NAME | AGE | SEX | GENDER | FAM HIS | SMOKING | ALCOHOL | BMI | WHR | SBP | DBP | HDL | LDL | TGL | тс | FBS | SI | HOMA IR | GROUP |
|-------|------------|-----|-----|--------|---------|---------|---------|-------|------|-----|-----|-----|-----|-----|-----|-----|------|---------|-------|
| 1 | Vincent | 51 | м | 1 | 1 | 0 | 1 | 28.4 | 0.91 | 100 | 70 | 37 | 122 | 209 | 244 | 105 | 25.4 | 6.58 | 1 |
| 2 | Susainat | 58 | м | 1 | 1 | 1 | 0 | 24.62 | 0.88 | 100 | 70 | 29 | 110 | 207 | 220 | 103 | 28.4 | 7.22 | 1 |
| 3 | Thangara | 64 | м | 0 | 1 | 1 | 1 | 29.4 | 0.92 | 110 | 70 | 31 | 88 | 163 | 147 | 102 | 32.4 | 8.16 | 1 |
| 4 | Balarama | 65 | м | 0 | 1 | 0 | 0 | 23.8 | 0.82 | 130 | 80 | 34 | 95 | 131 | 158 | 105 | 22.9 | 5.93 | 1 |
| 5 | Elumalai | 44 | м | 0 | 0 | 0 | 1 | 24.6 | 0.86 | 140 | 80 | 37 | 95 | 157 | 190 | 106 | 19.6 | 5.12 | 1 |
| 6 | Mathivan | 50 | м | 0 | 1 | 1 | 0 | 25.11 | 0.92 | 160 | 90 | 39 | 116 | 199 | 226 | 105 | 23.6 | 6.11 | 1 |
| 7 | Radhakri | 47 | м | 1 | 0 | 0 | 1 | 27.6 | 0.84 | 130 | 80 | 33 | 120 | 205 | 246 | 108 | 25 | 6.66 | 1 |
| 8 | Akila | 44 | F | 0 | 0 | 0 | 1 | 28.3 | 0.93 | 120 | 80 | 27 | 126 | 215 | 266 | 104 | 29.5 | 7.57 | 1 |
| 9 | Nasreen | 56 | F | 0 | 1 | 0 | 0 | 29.6 | 0.96 | 130 | 80 | 29 | 104 | 159 | 196 | 109 | 39.5 | 10.6 | 1 |
| 10 | Akila | 42 | F | 0 | 1 | 0 | 0 | 21.64 | 0.76 | 120 | 80 | 27 | 96 | 145 | 186 | 99 | 19.8 | 4.84 | 1 |
| 11 | baskar | 41 | м | 1 | 1 | 1 | 0 | 20.07 | 0.74 | 120 | 80 | 29 | 76 | 123 | 108 | 98 | 17.2 | 4.16 | 1 |
| 12 | Govindan | 45 | м | 1 | 1 | 1 | 1 | 22.8 | 0.72 | 140 | 70 | 32 | 90 | 152 | 155 | 106 | 22.4 | 5.86 | 1 |
| 13 | Pencilli | 65 | м | 0 | 0 | 1 | 1 | 26.37 | 0.9 | 130 | 80 | 35 | 132 | 209 | 264 | 108 | 32.9 | 8.77 | 1 |
| 14 | Govindar | 56 | м | 0 | 0 | 1 | 1 | 29.68 | 0.92 | 140 | 90 | 31 | 96 | 163 | 189 | 102 | 21 | 5.29 | 1 |
| 15 | Padmavathy | 55 | F | 0 | 0 | 0 | 0 | 21.6 | 0.76 | 130 | 80 | 37 | 102 | 165 | 176 | 91 | 29.5 | 6.62 | 1 |
| 16 | Lakshmi | 49 | F | 0 | 1 | 0 | 0 | 19.8 | 0.63 | 120 | 80 | 27 | 92 | 153 | 156 | 95 | 18 | 4.22 | 1 |
| 17 | Chinnath | 55 | м | 1 | 0 | 1 | 1 | 18.6 | 0.71 | 130 | 80 | 25 | 90 | 147 | 168 | 94 | 17.8 | 4.13 | 1 |
| 18 | Selvam | 48 | м | 1 | 0 | 0 | 0 | 22.5 | 0.82 | 120 | 80 | 27 | 92 | 167 | 184 | 98 | 19.5 | 4.71 | 1 |
| 19 | Vijayala | 63 | F | 0 | 1 | 0 | 0 | 23.43 | 0.76 | 130 | 70 | 29 | 96 | 165 | 182 | 96 | 14.2 | 3.36 | 1 |
| 20 | Rajendra | 52 | м | 1 | 1 | 0 | 1 | 28.62 | 0.92 | 130 | 80 | 33 | 124 | 213 | 258 | 108 | 15.8 | 4.21 | 1 |
| 21 | Saravana | 48 | м | 1 | 1 | 1 | 1 | 21.8 | 0.68 | 140 | 70 | 35 | 122 | 207 | 268 | 103 | 14.6 | 3.71 | 1 |
| 22 | Chinnasa | 45 | м | 1 | 0 | 1 | 1 | 27.8 | 0.94 | 130 | 80 | 39 | 126 | 209 | 276 | 105 | 14.5 | 3.75 | 1 |
| 23 | Munusamy | 46 | м | 1 | 0 | 1 | 1 | 22.5 | 0.68 | 120 | 70 | 35 | 88 | 157 | 161 | 93 | 13.8 | 3.16 | 1 |
| 24 | Kuppan | 55 | м | 1 | 1 | 0 | 1 | 27.5 | 0.92 | 150 | 90 | 37 | 86 | 163 | 194 | 102 | 14 | 3.52 | 1 |
| 25 | Balaji | 47 | м | 1 | 1 | 1 | 0 | 19.6 | 0.82 | 120 | 70 | 39 | 82 | 153 | 182 | 97 | 14 | 3.35 | 1 |
| 26 | Murali | 48 | м | 1 | 0 | 0 | 0 | 26.4 | 0.76 | 130 | 80 | 33 | 98 | 167 | 226 | 104 | 17.8 | 4.57 | 1 |
| 27 | Mahohar | 44 | м | 1 | 1 | 1 | 0 | 24.2 | 0.74 | 120 | 80 | 31 | 86 | 157 | 186 | 102 | 19.8 | 4.98 | 1 |
| 28 | Mani | 42 | м | 1 | 1 | 1 | 0 | 21.6 | 0.68 | 130 | 80 | 29 | 8 | 147 | 174 | 96 | 14.6 | 3.46 | 1 |
| 29 | Rugavan | 40 | м | 1 | 0 | 1 | 0 | 22.8 | 0.86 | 140 | 90 | 27 | 126 | 205 | 252 | 108 | 24.8 | 6.61 | 1 |
| 30 | Ganatham | 58 | м | 1 | 0 | 0 | 1 | 23.6 | 0.78 | 130 | 80 | 35 | 122 | 213 | 264 | 109 | 29.6 | 7.96 | 1 |
| 31 | Jilal | 54 | м | 1 | 0 | 0 | 0 | 28.6 | 0.94 | 140 | 80 | 37 | 120 | 227 | 284 | 107 | 28.8 | 7.6 | 1 |
| 32 | Daniel | 52 | м | 1 | 1 | 1 | 0 | 22.5 | 0.76 | 120 | 80 | 33 | 96 | 143 | 169 | 94 | 18.6 | 4.31 | 1 |
| 33 | Arunacha | 48 | М | 1 | 0 | 0 | 0 | 23.8 | 0.82 | 130 | 80 | 31 | 102 | 163 | 192 | 96 | 17.8 | 4.21 | 1 |

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| i - | i . | l | I | | 1 | | | 1 1 | | 1 | 1 | I | I. | I | 1 | | l | 1 | |
|-----|----------|----|---|---|---|---|---|-------|------|-----|-----|----|-----|-----|-----|-----|------|------|---|
| 34 | Shanthi | 50 | F | 0 | 0 | 0 | 1 | 26.2 | 0.8 | 140 | 80 | 33 | 104 | 189 | 220 | 98 | 29.6 | 7.16 | 1 |
| 35 | Kathires | 52 | м | 1 | 0 | 1 | 1 | 27.6 | 0.92 | 140 | 80 | 35 | 120 | 209 | 226 | 104 | 26.4 | 6.77 | 1 |
| 36 | Yusuf | 56 | м | 1 | 1 | 1 | 0 | 28.2 | 0.94 | 130 | 80 | 37 | 122 | 213 | 244 | 108 | 32.4 | 8.64 | 1 |
| 37 | Malini | 52 | F | 0 | 0 | 0 | 0 | 22.4 | 0.78 | 120 | 80 | 31 | 96 | 147 | 186 | 102 | 16.8 | 4.23 | 1 |
| 38 | Madhavan | 44 | м | 1 | 0 | 1 | 0 | 23.6 | 0.76 | 130 | 80 | 29 | 92 | 145 | 182 | 92 | 14.6 | 3.31 | 1 |
| 39 | Vinodhin | 50 | F | 0 | 1 | 0 | 0 | 21.6 | 0.68 | 120 | 80 | 31 | 86 | 141 | 176 | 96 | 13.8 | 3.27 | 1 |
| 40 | Srinivas | 55 | м | 1 | 0 | 0 | 0 | 26.4 | 0.88 | 140 | 80 | 35 | 104 | 183 | 192 | 98 | 21.6 | 5.22 | 1 |
| 41 | Usha | 52 | F | 0 | 0 | 0 | 0 | 20.6 | 0.66 | 110 | 70 | 33 | 86 | 165 | 168 | 88 | 14.4 | 3.12 | 1 |
| 42 | Chandra | 56 | F | 0 | 1 | 0 | 0 | 21.2 | 0.62 | 100 | 70 | 29 | 82 | 163 | 272 | 92 | 16.8 | 3.81 | 1 |
| 43 | Durai | 57 | м | 1 | 0 | 1 | 0 | 22.4 | 0.72 | 110 | 70 | 31 | 90 | 165 | 178 | 102 | 21.4 | 5.38 | 1 |
| 44 | Sakuntha | 68 | F | 0 | 0 | 0 | 0 | 28.4 | 0.94 | 110 | 80 | 37 | 122 | 209 | 246 | 104 | 26.8 | 6.88 | 1 |
| 45 | Venkates | 46 | м | 1 | 1 | 1 | 0 | 27.6 | 0.96 | 140 | 80 | 35 | 128 | 217 | 264 | 108 | 24.6 | 6 | 1 |
| 46 | Jayaraj | 44 | м | 1 | 0 | 1 | 0 | 24.6 | 0.82 | 130 | 80 | 33 | 102 | 205 | 202 | 104 | 24.8 | 6.36 | 1 |
| 47 | Thanagam | 52 | F | 0 | 0 | 0 | 0 | 22.4 | 0.78 | 120 | 80 | 29 | 98 | 149 | 186 | 98 | 21.4 | 5.17 | 1 |
| 48 | Subraman | 53 | м | 1 | 0 | 0 | 0 | 23.6 | 0.8 | 110 | 80 | 31 | 92 | 163 | 182 | 102 | 19.6 | 4.93 | 1 |
| 49 | Gajendra | 50 | м | 1 | 0 | 0 | 0 | 24.8 | 0.96 | 120 | 80 | 27 | 94 | 169 | 178 | 88 | 22.6 | 4.91 | 1 |
| 50 | Dhanalax | 66 | F | 0 | 0 | 0 | 0 | 20.6 | 0.68 | 100 | 80 | 29 | 86 | 153 | 168 | 86 | 15.5 | 3.29 | 1 |
| 51 | Devaraj | 55 | м | 1 | 0 | 0 | 0 | 22.6 | 0.84 | 130 | 90 | 46 | 92 | 132 | 184 | 109 | 13.3 | 3.3 | 0 |
| 52 | Ravi | 49 | м | 1 | 0 | 1 | 1 | 23.4 | 0.66 | 100 | 70 | 44 | 126 | 186 | 251 | 102 | 10.6 | 2.6 | 0 |
| 53 | Kumar | 54 | м | 1 | 0 | 1 | 1 | 20.8 | 0.77 | 130 | 100 | 47 | 70 | 133 | 144 | 103 | 11.1 | 2.8 | 0 |
| 54 | Yobu | 55 | м | 1 | 0 | 1 | 1 | 21.2 | 0.68 | 140 | 90 | 42 | 84 | 134 | 167 | 96 | 10.6 | 2.51 | 0 |
| 55 | Rajendra | 45 | м | 1 | 0 | 0 | 0 | 20.64 | 0.82 | 130 | 90 | 46 | 102 | 136 | 186 | 82 | 10.2 | 2.06 | 0 |
| 56 | Pushpa | 55 | F | 0 | 0 | 0 | 0 | 21.2 | 0.71 | 130 | 80 | 48 | 103 | 142 | 196 | 76 | 11.5 | 2.15 | 0 |
| 57 | Maniula | 56 | F | 0 | 1 | 0 | 0 | 22.6 | 0.58 | 120 | 80 | 42 | 104 | 138 | 168 | 85 | 14 | 2.93 | 0 |
| 58 | Latha | 54 | F | 0 | 0 | 0 | 0 | 21.8 | 0.66 | 130 | 80 | 38 | 96 | 134 | 188 | 78 | 10.8 | 2.02 | 0 |
| 59 | Lalitha | 57 | F | 0 | 0 | 0 | 0 | 27.6 | 0.86 | 120 | 80 | 42 | 115 | 194 | 230 | 95 | 12 | 2.8 | 0 |
| 60 | Manner | 45 | м | 1 | 0 | 1 | 0 | 19.2 | 0.63 | 130 | 80 | 44 | 84 | 106 | 164 | 102 | 11.6 | 2.92 | 0 |
| 61 | Viswanat | 42 | м | 1 | 0 | 1 | 1 | 243.2 | 0.76 | 120 | 80 | 46 | 122 | 146 | 247 | 84 | 11.2 | 2.32 | 0 |
| 62 | Subraman | 51 | м | 1 | 1 | 0 | 0 | 22.08 | 0.69 | 120 | 70 | 42 | 104 | 136 | 108 | 89 | 10.6 | 2.33 | 0 |
| 63 | Geetha | 58 | F | 0 | 1 | 0 | 0 | 21.35 | 0.7 | 110 | 70 | 44 | 94 | 108 | 116 | 83 | 10.6 | 2.61 | 0 |
| 64 | Padma | 42 | F | 0 | 1 | 0 | 0 | 31.62 | 0.88 | 120 | 70 | 42 | 126 | 184 | 266 | 99 | 12.9 | 2.81 | 0 |
| 65 | Muhamed | 47 | м | 1 | 1 | 0 | 0 | 22.8 | 0.68 | 120 | 80 | 44 | 98 | 125 | 189 | 85 | 11.5 | 1.99 | 0 |
| 66 | Prabu | 47 | M | 1 | 0 | 0 | 0 | 22.6 | 0.74 | 190 | 70 | 44 | 96 | 125 | 169 | 78 | 9.6 | 1.99 | 0 |
| 67 | Ettiappa | 43 | M | 1 | 0 | 0 | 0 | 23.6 | 0.74 | 100 | 80 | 42 | 94 | 128 | 169 | 104 | 9.6 | 3.21 | 0 |
| 67 | сшарра | 4/ | M | 1 | U | U | U | 22.4 | 0.82 | 100 | 80 | 44 | 92 | 134 | 108 | 104 | 10.2 | 3.21 | U |

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| 1 | 1 | | I | | | | I | 1 | | 1 | 1 | | I | I | | I | I | I | I I |
|-----|----------|----|---|---|---|---|---|-------|------|-----|----|----|-----|-----|-----|-----|------|------|-----|
| 68 | Mahima | 48 | F | 0 | 1 | 0 | 0 | 24.6 | 0.8 | 100 | 70 | 42 | 88 | 142 | 196 | 101 | 12.5 | 3.21 | 0 |
| 69 | Veerarag | 56 | М | 1 | 0 | 0 | 0 | 24.22 | 0.72 | 110 | 70 | 39 | 122 | 189 | 246 | 90 | 10.2 | 2.26 | 0 |
| 70 | Jayaraj | 56 | м | 1 | 0 | 1 | 1 | 22.8 | 0.68 | 120 | 70 | 42 | 106 | 196 | 184 | 96 | 11.8 | 2.79 | 0 |
| 71 | Santhosh | 55 | м | 1 | 0 | 0 | 0 | 20.8 | 0.72 | 110 | 70 | 40 | 98 | 188 | 208 | 92 | 8.2 | 1.86 | 0 |
| 72 | Jahir Hu | 40 | м | 1 | 0 | 1 | 1 | 22.6 | 0.67 | 100 | 70 | 36 | 88 | 142 | 196 | 86 | 11.5 | 2.44 | 0 |
| 73 | Vijaya | 48 | F | 0 | 0 | 0 | 0 | 28.6 | 0.84 | 108 | 70 | 44 | 122 | 186 | 232 | 91 | 9.6 | 2.16 | 0 |
| 74 | Chellamm | 51 | F | 0 | 0 | 0 | 0 | 24.6 | 0.72 | 130 | 80 | 44 | 98 | 136 | 208 | 86 | 10.8 | 2.25 | 0 |
| 75 | Bee Bee | 55 | F | 0 | 0 | 0 | 0 | 22.6 | 0.82 | 140 | 70 | 42 | 84 | 106 | 184 | 82 | 11.4 | 2.3 | 0 |
| 76 | Parvathy | 50 | F | 0 | 0 | 0 | 0 | 24.6 | 0.78 | 130 | 80 | 44 | 110 | 164 | 220 | 86 | 10.6 | 2.25 | 0 |
| 77 | Saraswat | 62 | F | 0 | 0 | 1 | 0 | 20.8 | 0.8 | 140 | 90 | 46 | 92 | 136 | 196 | 102 | 14 | 3.52 | 0 |
| 78 | Vasantha | 58 | F | 0 | 0 | 0 | 0 | 28.6 | 0.94 | 150 | 90 | 42 | 82 | 132 | 189 | 80 | 10.9 | 2.15 | 0 |
| 79 | Lakshmi | 55 | F | 0 | 1 | 0 | 0 | 31.1 | 0.92 | 130 | 80 | 36 | 78 | 128 | 157 | 88 | 7.5 | 1.62 | 0 |
| 80 | Thenmozh | 54 | F | 0 | 0 | 0 | 0 | 22.6 | 0.68 | 120 | 80 | 42 | 92 | 132 | 185 | 84 | 8.3 | 1.72 | 0 |
| 81 | Vijayala | 64 | F | 0 | 0 | 0 | 0 | 23.43 | 0.72 | 130 | 80 | 40 | 90 | 128 | 180 | 96 | 14.2 | 3.36 | 0 |
| 82 | Selvam | 58 | м | 1 | 1 | 1 | 0 | 21.8 | 0.66 | 120 | 80 | 38 | 88 | 126 | 190 | 109 | 12.3 | 3.31 | 0 |
| 83 | Frakash | 54 | м | 1 | 0 | 0 | 0 | 22.6 | 0.64 | 120 | 80 | 42 | 96 | 134 | 176 | 98 | 11.2 | 2.71 | 0 |
| 84 | Vinodh | 66 | м | 1 | 0 | 1 | 1 | 23.4 | 0.74 | 130 | 80 | 44 | 102 | 142 | 188 | 86 | 12.6 | 2.67 | 0 |
| 85 | Kala | 54 | F | 0 | 0 | 0 | 0 | 24.8 | 0.82 | 120 | 80 | 42 | 108 | 156 | 216 | 96 | 13.8 | 3.27 | 0 |
| 86 | Vikram | 42 | м | 1 | 0 | 0 | 0 | 21.6 | 0.72 | 130 | 80 | 44 | 104 | 166 | 204 | 104 | 14.6 | 3.75 | 0 |
| 87 | Sekar | 44 | м | 1 | 0 | 1 | 1 | 22.8 | 0.8 | 140 | 80 | 36 | 94 | 136 | 189 | 88 | 12.6 | 2.73 | 0 |
| 88 | Sankar | 46 | м | 1 | 1 | 0 | 0 | 23.6 | 0.66 | 130 | 80 | 38 | 96 | 134 | 194 | 92 | 3.2 | 2.99 | 0 |
| 89 | Gunasaga | 44 | м | 1 | 0 | 0 | 1 | 24.8 | 0.92 | 140 | 80 | 40 | 108 | 186 | 226 | 106 | 15.8 | 4.13 | 0 |
| 90 | Rajendra | 56 | м | 1 | 0 | 1 | 0 | 26.5 | 0.74 | 130 | 80 | 42 | 96 | 126 | 128 | 92 | 12.8 | 2.9 | 0 |
| 91 | Kumar | 52 | м | 1 | 1 | 0 | 0 | 18.5 | 0.68 | 120 | 80 | 38 | 94 | 124 | 189 | 88 | 11.6 | 2.52 | 0 |
| 92 | Ganesh | 44 | м | 1 | 0 | 0 | 1 | 20.5 | 0.66 | 130 | 80 | 40 | 92 | 134 | 192 | 86 | 10.8 | 2.29 | 0 |
| 93 | Vignesh | 52 | м | 1 | 0 | 0 | 1 | 21.5 | 0.71 | 120 | 80 | 37 | 90 | 132 | 184 | 102 | 12.5 | 3.14 | 0 |
| 94 | Seetha | 46 | F | 0 | 1 | 0 | 0 | 22.5 | 0.72 | 120 | 80 | 38 | 96 | 142 | 168 | 94 | 13.5 | 3.13 | 0 |
| 95 | Raju | 48 | м | 1 | 0 | 1 | 1 | 24.5 | 0.78 | 130 | 80 | 44 | 98 | 186 | 184 | 92 | 14.5 | 3.29 | 0 |
| 96 | Sundaram | 49 | м | 1 | 0 | 0 | 0 | 22.6 | 0.64 | 140 | 80 | 42 | 94 | 142 | 194 | 102 | 12.2 | 3.07 | 0 |
| 97 | Chandran | 51 | м | 1 | 1 | 0 | 0 | 20.5 | 0.71 | 120 | 80 | 36 | 86 | 126 | 178 | 90 | 11.8 | 2.62 | 0 |
| 98 | Saravana | 46 | м | 1 | 0 | 1 | 1 | 18.5 | 0.66 | 130 | 80 | 40 | 92 | 138 | 194 | 82 | 2.3 | 2.49 | 0 |
| 99 | Senthil | 52 | M | 1 | 0 | 0 | 0 | 20.75 | 0.68 | 120 | 80 | 40 | 88 | 128 | 174 | 84 | 14.2 | 2.45 | 0 |
| | | 48 | M | 1 | 0 | 0 | 0 | | | | 80 | | 98 | 128 | | 88 | | 3.71 | 0 |
| 100 | Murugesa | 48 | M | 1 | 0 | 0 | 0 | 22.75 | 0.72 | 130 | 80 | 38 | 98 | 138 | 184 | 88 | 14.6 | 3.71 | 0 |

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