

**“DESIGN, SYNTHESIS, CHARACTERIZATION AND BIOLOGICAL
EVALUATION OF SOME NOVEL INDOLE DERIVATIVES AS ANTI
TUBERCULAR AGENTS TARGETING GLUTAMINE SYNTHETASE 1”**

**A Dissertation Submitted to
THE TAMILNADU Dr.M.G.R. MEDICAL UNIVERSITY
CHENNAI – 600032**

In partial fulfilment of the requirements for the award of the Degree of

**MASTER OF PHARMACY
IN
PHARMACEUTICAL CHEMISTRY**

**Submitted by
KALAIYARASI.P
Reg.No : 261715702**

**Under the guidance of
Dr.A.JERAD SURESH M. Pharm., Ph.D., M.B.A
Professor & Head
Department of Pharmaceutical Chemistry
College of Pharmacy, Madras Medical College**



**DEPARTMENT OF PHARMACEUTICAL CHEMISTRY
COLLEGE OF PHARMACY, MADRAS MEDICAL COLLEGE
CHENNAI-600 003**

MAY 2019

**“DESIGN, SYNTHESIS, CHARACTERIZATION AND BIOLOGICAL
EVALUATION OF SOME NOVEL INDOLE DERIVATIVES AS ANTI
TUBERCULAR AGENTS TARGETING GLUTAMINE SYNTHETASE 1”**

**A Dissertation Submitted to
THE TAMILNADU Dr.M.G.R. MEDICAL UNIVERSITY
CHENNAI – 600032**

In partial fulfilment of the requirements for the award of the Degree of

**MASTER OF PHARMACY
IN
PHARMACEUTICAL CHEMISTRY**

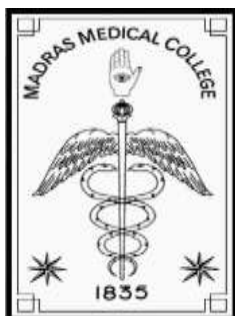
**Submitted by
KALAIYARASI.P
Reg.No : 261715702**

**Under the guidance of
Dr.A.JERAD SURESH M. Pharm., Ph.D., M.B.A
Professor & Head
Department of Pharmaceutical Chemistry
College of Pharmacy, Madras Medical College**



**DEPARTMENT OF PHARMACEUTICAL CHEMISTRY
COLLEGE OF PHARMACY, MADRAS MEDICAL COLLEGE
CHENNAI-600 003**

MAY 2019



**COLLEGE OF PHARMACY
MADRAS MEDICAL COLLEGE
CHENNAI-600 003
TAMILNADU**



CERTIFICATE

This is to certify that the dissertation entitled **“DESIGN, SYNTHESIS, CHARACTERIZATION AND BIOLOGICAL EVALUATION OF SOME NOVEL INDOLE DERIVATIVES AS ANTI TUBERCULAR AGENTS TARGETING GLUTAMINE SYNTHETASE 1”** submitted by the candidate bearing the Register No: **261715702** in partial fulfilment of the requirements for the award of degree of **MASTER OF PHARMACY in PHARMACEUTICAL CHEMISTRY** by the TamilNadu Dr.M.G.R Medical University is a bonafide work done by her during the academic year 2018-2019 in the Department of Pharmaceutical Chemistry, College of Pharmacy, Madras Medical College, Chennai- 600 003.

Dr.A.JERAD SURESH, M.Pharm.,Ph.D.,M.B.A.,

Principal & Head,

Department of Pharmaceutical Chemistry,

College of Pharmacy,

Madras Medical College,

Chennai- 600 003.



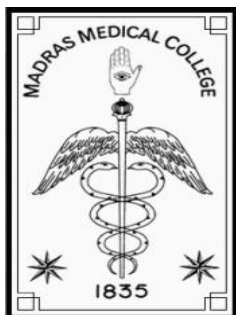
**COLLEGE OF PHARMACY
MADRAS MEDICAL COLLEGE
CHENNAI-600 003
TAMILNADU**



CERTIFICATE

This is to certify that the dissertation entitled **“DESIGN, SYNTHESIS, CHARACTERIZATION AND BIOLOGICAL EVALUATION OF SOME NOVEL INDOLE DERIVATIVES AS ANTI TUBERCULAR AGENTS TARGETING GLUTAMINE SYNTHETASE 1”** submitted by the candidate bearing the Register No: **261715702** in partial fulfilment of the requirements for the award of degree of **MASTER OF PHARMACY in PHARMACEUTICAL CHEMISTRY** by the TamilNadu Dr. M.G.R Medical University is a bonafide work done by her during the academic year 2018-2019 in the Department of Pharmaceutical Chemistry, College of Pharmacy, Madras Medical College, Chennai- 600003, under my direct supervision and guidance.

Dr. A.JERAD SURESH, M.Pharm., Ph.D., M.B.A.,
PROJECT ADVISOR,
Principal, Professor and Head,
Department of Pharmaceutical Chemistry,
College of Pharmacy,
Madras Medical College, Chennai- 600003.



COLLEGE OF PHARMACY
MADRAS MEDICAL COLLEGE
CHENNAI-600 003
TAMILNADU



CERTIFICATE

This is to certify that the dissertation entitled “**DESIGN, SYNTHESIS, CHARACTERIZATION AND BIOLOGICAL EVALUATION OF SOME NOVEL INDOLE DERIVATIVES AS ANTI TUBERCULAR AGENTS TARGETING *GLUTAMINE SYNTHETASE I***” submitted by the candidate bearing the Register No: **261715702** to the TamilNadu Dr. M.G.R Medical University, examination is evaluated.

EXAMINERS

1.

2.



ACKNOWLEDGEMENT

First and foremost I thank the Almighty for all the love, grace and blessings he has bestowed upon me, for strengthening me throughout, to successfully complete this dissertation.

I consider this as an opportunity to express my gratitude to all the people ,who have been involved directly or indirectly with the successful completion of this dissertation.

I sincerely express my honorable thanks to **Dr. R.JAYANTHI, M.D.,F.R.C.P (Glasg)** DEAN, Madras Medical College, for giving an opportunity to carry out my project work.

It is my great pleasure to express my gratitude to my esteemed Principal and Guide **DR.A.JERAD SURESH, M.Pharm, Ph.D,M.B.A,** Department of Pharmaceutical Chemistry, College of Pharmacy, Madras Medical College, Chennai-03, who took keen interest on my project work and guided me all along, till the completion of my work by providing all the necessary information for developing a good system.

I wish to thank all my teaching staff **Dr.R.Priyadarsini, M.Pharm, Ph.D., Mrs.T.Saraswathy, M.Pharm., Dr.M.Sathish, M.Pharm.,Ph.D., and Dr.P.G.Sunitha, M.Pharm,Ph.D.,** Assistant Professors in Pharmacy, Department of Pharmaceutical Chemistry and other Department staffs for their gracious support and encouragement in making this work successful.

I express my thanks to all non-teaching staff members **Mrs.Geetha, Mr. Umapathy Mrs.Vijaya, Mrs.Maheshwari and Mrs.Mala,** Department of Pharmaceutical Chemistry, College of Pharmacy, Madras Medical College, Chennai- 03, for their assistance during my thesis work.

I would also record my thanks to **Dr.K.M.Noorulla M.pharm.,Ph.D., Dr.P.R. Suriya M.Pharm.,Ph.D., Mr.S.Sureshkumar, Mrs.M.Fathima Jesiya, Mr.E.Ayyamperumal** for their timely help, valuable suggestions and comments during my project.

I convey my thanks to **CATERS-CLRI, IISRM CHENNAI** and **Dr.Kishore Bhatt**, Professor and Head, Department of Microbiology, Maratha Mandal's Institute Of Dental Science And Research Institute, Belgaum, Karnataka for support in carrying out the spectral studies and *in-vitro* evaluation of anti-tubercular activity in due time.

A special thanks to my dear friends **S.Nandhini, A.Varalakshmi, V.Meenakumari T.Kanimozhi, L.S.Dhivya, P.Lokeshkumar, K.Sangeetha, and B.Yuvaraj** and other friends for their constant motivation and support.

I thank my Juniors **Banupriya, Sumathi, Priyadharshini, Anitha, Nandhini, Saravanan, Aarthi, Soundarya and Suresh** for their help and support.

Last but not the least , I owe my heartfelt and deepest gratitude to my mother **P.Nageshwari**, my dear brother, sisters and my wellwisher **C.Radha Krishnan**.



CONTENTS

| S.NO | TITLE | PAGE NO |
|-------------|---|----------------|
| 1 | INTRODUCTION ✓ TUBERCULOSIS ✓ BIOLOGICAL TARGET ✓ DRUG DISCOVERY ✓ BASIC NUCLEUS | 1 |
| 2 | REVIEW OF LITERATURE | 13 |
| 3 | AIM AND OBJECTIVE | 19 |
| 4 | MATERIALS AND METHODS ✓ DRUG DESIGN ✓ SYNTHETIC METHODOLOGY ✓ CHARACTERIZATION ✓ BIOLOGICAL EVALUATION | 21 |
| 5 | RESULTS AND DISCUSSION | 33 |
| 6 | SUMMARY AND CONCLUSION | 66 |
| 7 | REFERENCES | 68 |

LIST OF FIGURES

| FIGURE NO | TITLE | PAGE NO |
|------------------|---|----------------|
| 1 | <i>Mycobacterium tuberculosis</i> | 1 |
| 2 | Global incidence of tuberculosis | 4 |
| 3 | Structure of <i>mycobacterium tuberculosis</i> | 5 |
| 4 | Cell wall of <i>mycobacterium tuberculosis</i> | 6 |
| 5 | Genomic structure of <i>Mycobacterium tuberculosis</i> H37RV strain | 7 |
| s6 | Structure of <i>Glutamine synthetase1</i> enzyme | 9 |
| 7 | Structure of indole nucleus | 11 |
| 8 | standard drug photograph (MABA) | 64 |
| 9 | MABA report photograph for synthesized compounds | 64 |

LIST OF TABLES

| TABLE NO | TITLE | PAGE NO |
|----------|---|---------|
| 1 | Table of the selected molecules with docking score | 33 |
| 2 | Molecules interaction with amino acids and docking view | 34 |
| 3 | Results of toxicity prediction for selected molecules | 37 |
| 4 | Results of drug likeness prediction by Molinspiration | 39 |
| 5 | Rf values of the synthesized compounds | 41 |
| 6 | IR interpretation of the compound PK1 | 43 |
| 7 | NMR interpretation of the compound PK1 | 45 |
| 8 | IR interpretation of the compound PK2 | 47 |
| 9 | NMR interpretation of the compound PK2 | 49 |
| 10 | IR interpretation of the compound PK3 | 51 |
| 11 | NMR interpretation of the compound PK3 | 53 |
| 12 | IR interpretation of the compound PK4 | 55 |
| 13 | NMR interpretation of the compound PK4 | 57 |
| 14 | IR interpretation of the compound PK5 | 59 |
| 15 | NMR interpretation of the compound PK5 | 61 |
| 16 | physical data of the synthesized compounds | 62 |
| 17 | Molecular weight determined by Mass Spectrometry | 63 |
| 18 | The MABA report of the synthesized compounds | 65 |
| 19 | Comparative study of docking score with MABA report | 65 |

LIST OF ABBREVIATIONS

| | |
|--------|--|
| TB | Tubercle Bacillus |
| HIV | Human Immuno Deficiency Virus |
| AIDS | Acquired Immuno Deficiency Syndrome |
| DOTS | Directly Observed Treatment Short-Course |
| MDR-TB | Multi Drug Resistant tuberculosis |
| XRD-TB | Extensively Drug Resistant-TB |
| LTBI | Latent Tuberculosis Infection |
| CADD | Computer Aided Drug Design |
| OSIRIS | Optical, Spectroscopic and Infrared Remote ImagingSystem |
| SBDD | Structure Based Drug Design |
| LBDD | Ligand Based Drug Design |
| Logp | Partition Co-Efficient |
| WHO | World Health Organization |
| MIC | Minimum Inhibitory Concentration |
| PDB | Protein Data Bank |
| TLC | Thin Layer Chromatography |
| IR | Infrared Spectroscopy |
| NMR | Nuclear Magnetic Resonance |
| GC-MS | Gas Chromatography-Mass Spectroscopy |
| REMA | Resazurin Micro Plate Assay |
| MABA | Micro Plate Alamar Blue Assay |
| µg/ml | Microgram per milliliter |
| QSAR | Quantitative Structural Activity Relationship |
| °C | Celsius |
| 3D | Three Dimensional |
| mins | Minutes |
| Hrs | Hours |
| % | Percentage |

INTRODUCTION

*DEDICATED TO
MY FAMILY,
RESPECTED
TEACHERS AND
MY DEAR FRIENDS*

INTRODUCTION

Tuberculosis is a chronic infectious disease, usually caused by the bacteria *Mycobacterium tuberculosis*. It is known to be the major reason for mortality of nearly two million people in each year.^[1] About one third of the world's population is infected with TB and 5 to 10% of those can develop active TB in their life time^[2]. TB generally affects the lungs called as pulmonary tuberculosis, but also affects other parts of the body like it can affect the bones, the nervous system or many other organ systems called as extra pulmonary tuberculosis. Basically it is characterized as pulmonary disease which occurs due to the accumulation of *Mycobacterium tuberculosis* (MTB) onto the lungs alveolar surfaces.^[3] Synonyms for tuberculosis are phthisis, phthisis pulmonalis, consumption^[4]. TB infection can occur by inhaling the droplet containing *M.tuberculosis* organism by susceptible persons through cough, sneeze, spits, laughing, talks.^[5]



Fig.1 *Mycobacterium tuberculosis*^[1]

NEED FOR AN ANTI-TUBERCULAR AGENT:

- To enhance the activity against MDR-TB strains.
- To reduce the toxicity.
- To shorten the duration of therapy
- To rapid the microbicidal mechanism of action

SIGNS OF TB: ^[5]

- Cough that lasts more than 3 weeks
- Chest pain
- Coughing up blood
- Tired
- Night sweat
- Chills, fever, loss of appetite and weight loss

TYPES OF TUBERCULOSIS: ^[6]

ACTIVE TB:

The TB bacteria rapidly multiply and invade different organs of the body. The person with active TB may spread TB to others through air.

LATENT TB:

In this condition the bacteria remains inactive in our body and causes no symptoms. Latent TB is non-contagious but has a chance of becoming active

MILIARY TB:

Miliary TB is characterized by wide dissemination into the human body and by the tiny size of the lesions (1-5mm)

EPIDEMIOLOGY:

- In 2011, there were 8.7 million new cases of active tuberculosis worldwide (13% of which involved co-infection with the Human Immuno Deficiency Virus [HIV]) and 1.4 million deaths, including 430,000 deaths among HIV-infected patients representing a slight decrease from peak numbers in the mid-2000s (Fig. 2).^[7]
- It has been estimated that there were 310,000 incident cases of multidrug-resistant tuberculosis, caused by organisms resistant to at least isoniazid and rifampicin, among patients who were reported to have tuberculosis in 2011.
- More than 60% of these patients were in China, India, the Russian Federation, Pakistan, and South Africa.
- The absolute number of cases is highest in Asia, with India and China having the greatest burden of the disease globally.^[7]
- A total of 84 countries have reported cases of extensively drug-resistant tuberculosis, a subset of multidrug-resistant tuberculosis with added resistance to all fluoroquinolones plus any of the three injectable antituberculosis drugs, kanamycin, amikacin, and capreomycin.^[8-9]

TB BURDEN IN INDIA

Each year 12 lakh (1,200,000) Indians are notified (that is reported to the RNTCP) as having newly diagnosed TB. In addition at least 2.7 lakh (270,000) Indians die. Some estimates calculate the deaths as being twice as high.^[7]

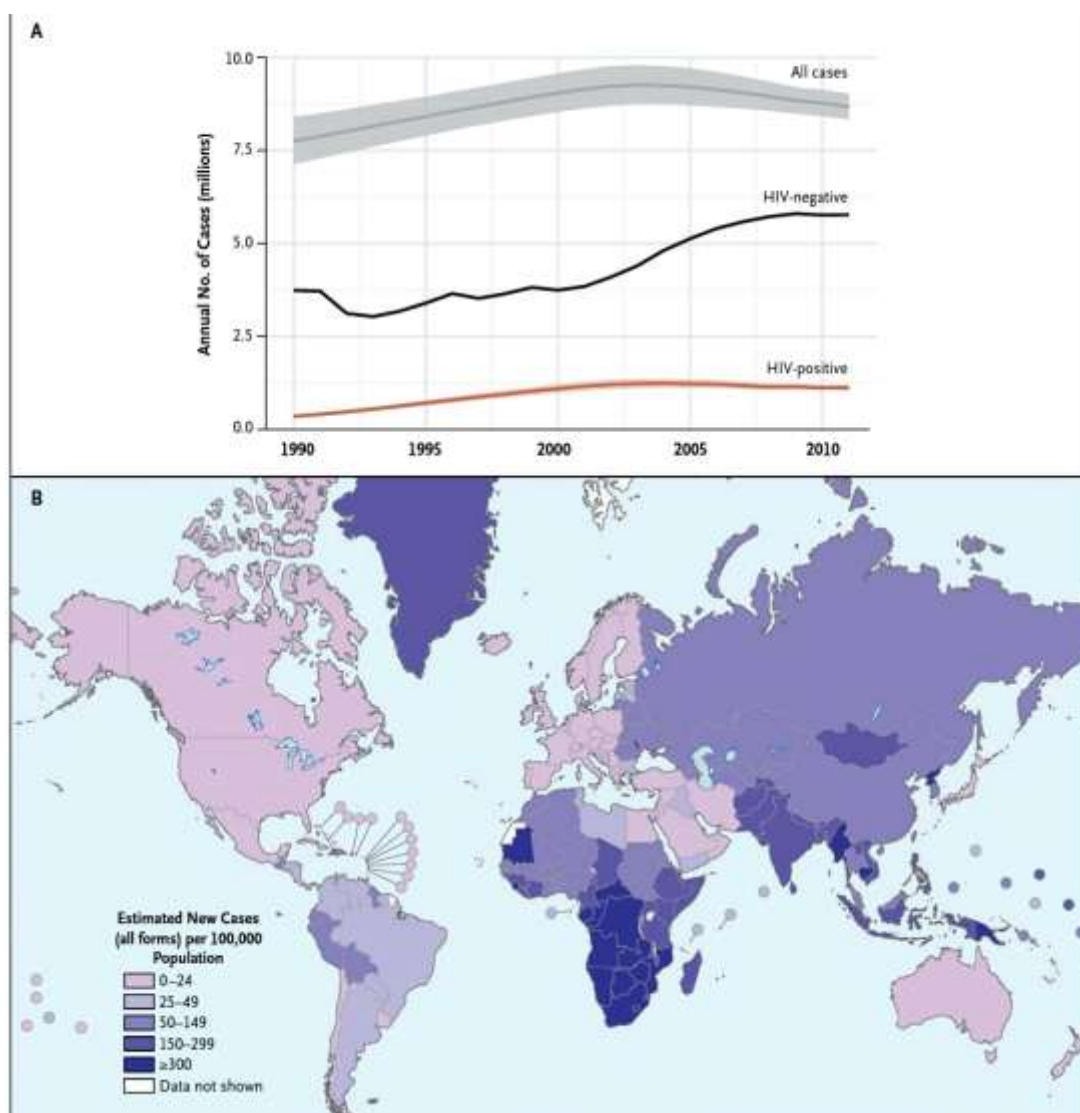


Fig.2 Global incidence of tuberculosis^[31]

Panel A shows global trends in the estimated incidence of tuberculosis from 1990 to 2011 among all patients, those with human immunodeficiency virus (HIV) co-infection, and without HIV co-infection. The shading around the data curves indicates uncertainty intervals on the basis of available data. Panel B shows the estimated global incidence of tuberculosis in 2011.

MYCOBACTERIUM TUBERCULOSIS:

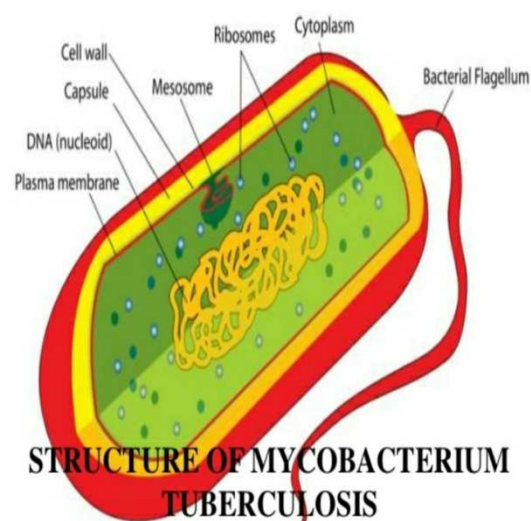


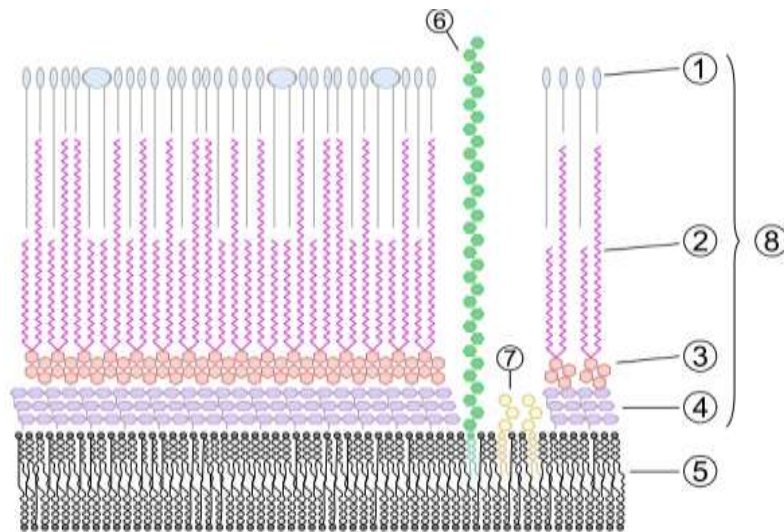
Fig.3 Structure of *Mycobacterium tuberculosis*^[10]

Mycobacterium tuberculosis is a species of pathogenic bacteria in the family of Mycobacteriaceae and the causative agent of tuberculosis. It was first discovered by Robert Koch in 1882. MTB are usually found on the well aerated upper lobes of the alveolar surfaces because they are aerobic in nature. MTB is not classified as a Gram negative or Gram positive bacteria because of a particular characteristic exhibited by its cell wall. It poses a waxy coating on its cell surface which is due to the presence of mycolic acid, which makes it insensitive to Gram staining. MTB is an intracellular parasite, and it has low generation time of 15-20 hours, which helps for its virulence factor. ^[10]

SCIENTIFIC CLASSIFICATION:^[4]

Domain : Bacteria
Phylum : Actinobacteria
Class : Actinobacteria
Order : Actinomycetales
Family : Mycobacteriaceae
Genus : *Mycobacterium*
Species : *M. tuberculosis*

CELL WALL OF MYCOBACTERIUM TUBERCULOSIS:



Mycobacterial cell wall: 1-outer lipids, 2-mycolic acid, 3-polysaccharides (arabinogalactan), 4-peptidoglycan, 5-plasma membrane, 6-lipoarabinomannan (LAM), 7-phosphatidylinositol mannoside, 8-cell wall skeleton

Fig 4. Cell wall of *mycobacterium tuberculosis*^[12]

The compositional and architectural complexity of mycobacterial cell envelope distinguishes species of the *Mycobacterium* genus from other prokaryotes. It is the basis for many of the physiological and pathogenic features of mycobacteria and the site of susceptibility and resistance to many other anti-tubercular drugs.^[11] The mycobacterial cell wall is made up of three segments, the plasma membrane, the cell wall core and outermost layer. The cell wall core is essential for viability, consists of peptidoglycan (PG) in covalent attachment via phosphoryl-N-acetylglucosaminosyl-ramnosyl linkage units with heteropolysaccharide arabinogalactan (AG). AG is in turn esterified at its non-reducing ends to long chain mycolic acids. The latter form the bulk of the inner leaflet of the outer membrane which consisting of non-covalently attached glycolipids, polysaccharides, lipoglycans and proteins.^[12]

GENOME OF *MYCOBACTERIUM TUBERCULOSIS*:

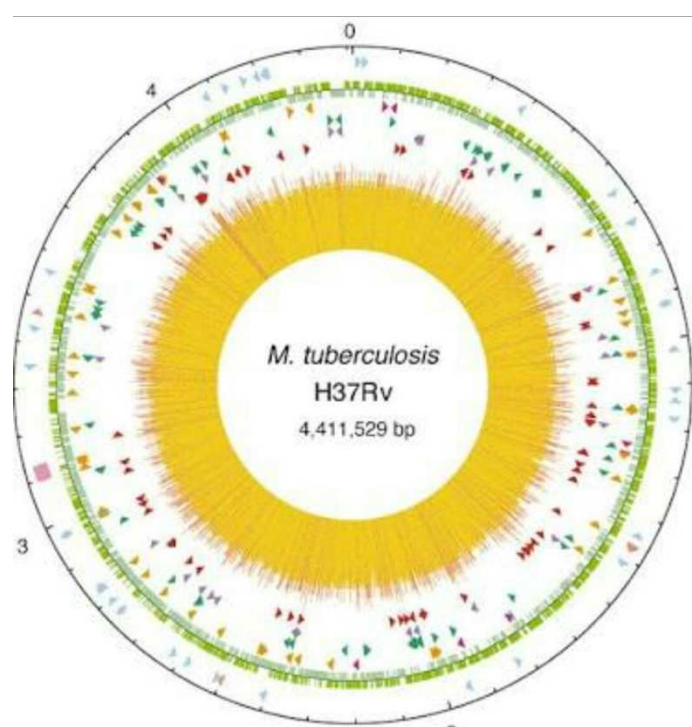


Fig.5 Genomic structure of *Mycobacterium tuberculosis* H37RV strain^[13]

The genome of the H37RV strain was published in 1998. *Mycobacterium tuberculosis* has circular chromosomes containing 4,200,000 nucleotides long and the Guanine+Cytosine content of 65%.^[13] The genome of *M. tuberculosis* was studied using the strain *M. tuberculosis* H37Rv^[14]. Its size is 4 million base pairs, with 3,959

genes. The genome contains 250 genes involved in fatty acid metabolism, with 39 of these involved in the polyketide metabolism generating the waxy coat. About 10% of the coding capacity is taken up by the PE/PPE gene families that encode acidic, glycine-rich proteins. Nine noncoding sRNAs have been characterized in *M.tuberculosis*.^[15-16]

DRUG DISCOVERY PROCESS:^[17]

CADD Strategy depends upon the extent of structural information available regarding the target (enzyme/receptor) and the ligands. Direct and indirect design are the two major modeling strategies used in the drug design process. Indirect approach involves designing of drug based on comparative analysis of the structural features of known active and inactive compounds which is otherwise known as Ligand based drug design. Direct design involves the three dimensional features of the target (enzyme/receptor), otherwise known as Structure based drug design.

PREPARATION OF A TARGET STRUCTURE:

Virtual screening depends upon the amount and quality of structural information about both the target and the small molecules being docked. The first step is to evaluate the target for the presence of an appropriate binding pocket. This is done through the analysis of known target-ligand co-crystal structures or using in-silico methods to identify novel binding sites.^[18-19]

X-ray crystallography or NMR techniques are used to determine the target structure experimentally. It is deposited in the PDB is the ideal starting point for docking. Based on comparative models of target proteins several successful virtual screening campaigns have been reported in the absence of experimentally determined structures.^[20-21]

STRUCTURE-BASED VIRTUAL HIGH-THROUGHPUT SCREENING:

SB-vHTS selects for ligand predicted to bind a particular binding site, inhibit, or allosterically alter the protein's function. The key steps in SB-vHTS are: (1) preparation of the target protein and compound library for docking, (2) determining a favorable binding pose for each compound and (3) ranking the docked structures.^[22]

BIOLOGICAL TARGET:

There are various biosynthetic enzymes that are essential for the survival of the Mycobacterium and are considered as potential drug targets. A comprehensive *insilico* target identification pipeline for *Mycobacterium tuberculosis* was identified and reported. It comprises a total of 451 high-confidence targets.^[23]

GLUTAMINE SYNTHETASE 1 ENZYME:

Glutamine Synthetase is an enzyme that plays an important role in nitrogen metabolism by catalyzing the condensation of glutamate and ammonia to form glutamine. *Glutamine synthetase* uses ammonia produced by nitrate reduction, amino acid degradation, and photorespiration. The amide group of glutamate is a nitrogen source for the synthesis of glutamine pathway metabolites. Competition between ammonium ion, influences glutamine synthesis and glutamine hydrolysis.^[24]

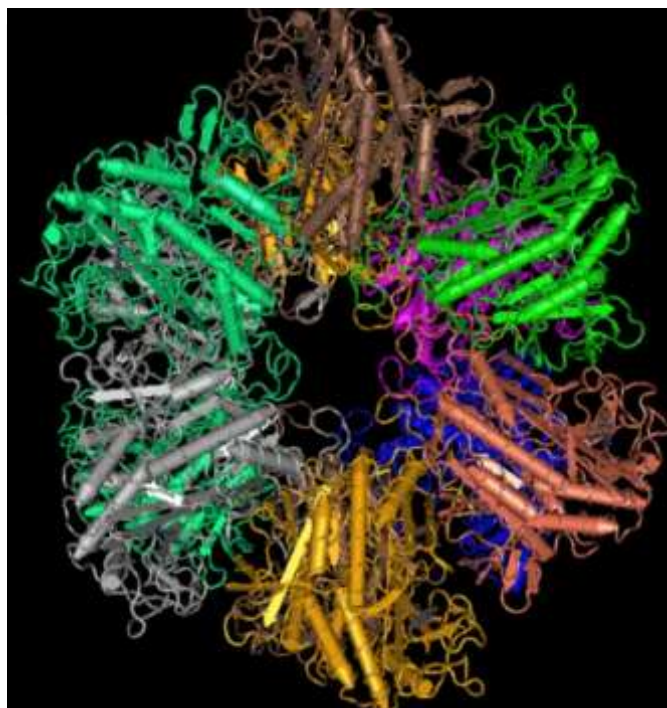


Fig 6. Structure of *Glutamine synthetase1* enzyme

| | | |
|------------------------|---|---|
| Protein name | : | <i>Glutamine synthetase1</i> |
| Classification | : | Ligase |
| Chains | : | A, B, C, D, E,F |
| Total structure weight | : | 332264.16 |
| Length | : | 486 |
| Gene name | : | <i>glnA1 glnA Rv2220 MTCY190.31 MTCY427.01</i> . [25] |

MECHANISM:

GS catalyzes the ATP dependent condensation of glutamate with ammonia to yield glutamine. This mechanism takes place in two step. [26-27]

The first step is the formation of the activated intermediate glutamyl phosphate. The Mg²⁺ ion coordinates the γ -phosphate oxygen of ATP to allow phosphoryl transfer to the carboxylate group of glutamate, yielding the intermediate (acyl phosphate). ADP and Pi do not dissociate until ammonia binds and glutamine is released. The presence of ADP causes a conformational shift in GS that stabilizes the γ -glutamyl phosphate moiety.

The second step is deprotonation of ammonium, which allows ammonia to attack the intermediate from its nearby site to form glutamine. The inhibition of GS secreted by *M.tuberculosis* is sufficient to halt the growth of the bacterium, suggesting that TB-GS might be a valid target for anti-tuberculosis drug-design. The structure of TB-GS is currently being solved to aid in the design of novel inhibitors for this enzyme .

BASIC NUCLEUS :

Indole nucleus continuously drawing interest for development of newer drug moiety due to its wide range of pharmacological activities such as antibacterial , antiinflammatory , analgesic, anti-viral , antifungal, anti-tubercular, anti-depressant. [28]

Indole is a bicyclic, heterocyclic ring system in which the benzene ring is fused with pyrrole ring through the α , β -position. The word Indole is derived from the words Indigo and Oleum since indole is first isolated by treatment of indigo dye with oleum. Indole nucleus is also found in many natural products such as indole alkaloids, fungal metabolites and marine natural products. Indole occurs in coal tar and in the oils of jasmine and orange blossoms. [30]

INDOLE:

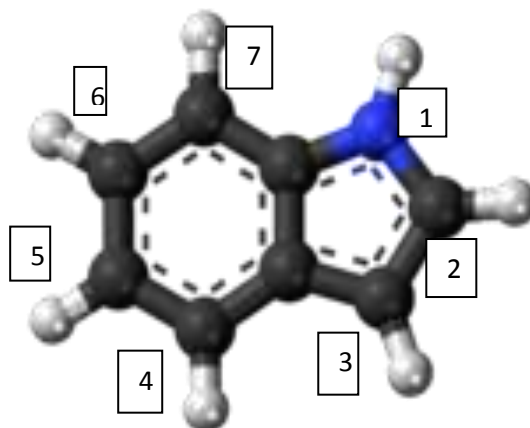


Fig.7 Structure of indole nucleus

Indole nucleus posses various medicinal activities. Insecticidal, Anti-viral activities of isatin and indole oximes and anti-inflammatory activity of indole-3-acetic acids has been reported. Oximes derivatives of 2-substituted indoles and 3-substituted indoles are reported to have Fungicidal activity. Antibacterial activity of some substituted 3-(aryl) and 3-(Heteroaryl) indoles have been reported. Indole derivatives were reported to have antioxidant activity . A new series of 1H-indole-2, 3-dione derivatives were reported for in vitro antituberculosis activity against Mycobacterium tuberculosis H37Rv [29]

REACTIVITY OF INDOLE:

- Indole can readily undergo aromatic electrophilic substitution. The C-3 position is the most nucleophilic, followed by the N and C-2 positions.
- The C-2 and C-3 bonds can often react like alkenes.
- Indole can be deprotonated at nitrogen. The resulting salts can be good nucleophiles.
- When N is substituted, C-2 can be deprotonated.
- Highly ionic salts favour N substitution.
- Softer counter ions favour C-3 substitution. ^[30]

*REVIEW OF
LITERATURE*

REVIEW OF LITERATURE

Review of literature regarding tuberculosis:

1. Alimuddin Zumla et al.,(2013) ^[31] reviewed the global incidence of tuberculosis and global number of cases with MDR-TB and current recommendations for tuberculosis treatment
2. Williams B.G et al.,(2010)^[32] studied about Population Dynamics and Control of Tuberculosis.
3. Robert Koch (2008) ^[33] detailed about the history of Tuberculosis.
4. Pierpalo de colombai *et al.*,(2007) ^[34] has described The Global Plan to Stop TB.
5. Balabanova Y *et al.*, (2006) ^[35] made a summary about “The Directly Observed Treatment Short-Course (DOTS) therapy.
6. Ruohonen RP *et al.*, (2002) ^[36] Implemented the Directly Observed Treatment Short-course strategy.
7. Keane J *et al.*, (1997) ^[37] reported that *Mycobacterium Tuberculosis* promotes Human alveolar macrophage apoptosis.

Review of literature regarding genomic aspects of *Mycobacterium Tuberculosis*:

8. Thomas R. Iorger et al.,(2010)^[38] studied about the variation among the Genomic sequences of H37Rv strains of *M.tuberculosis*. They carried whole genomic sequencing on six strains of H37Rv from different laboratories.
9. Zheng et al.,(2008) ^[39] determined the whole genomic sequence of attenuated *M.Tuberculosis* H37Ra, and performed comparative genomic analysis of H37Ra against its virulent counterpart H37Rv , and studied their genetic variations which is useful for understanding the pathogenesis of *M.Tuberculosis*.

Review of literature regarding the enzyme *glutamine synthetase I*:

10. Wojciech W. Krajewski et al.(2008) ^[40] summarized that *glutamine synthetase* catalyzes the ligation of glutamate and ammonia to form glutamine, with the hydrolysis of ATP.

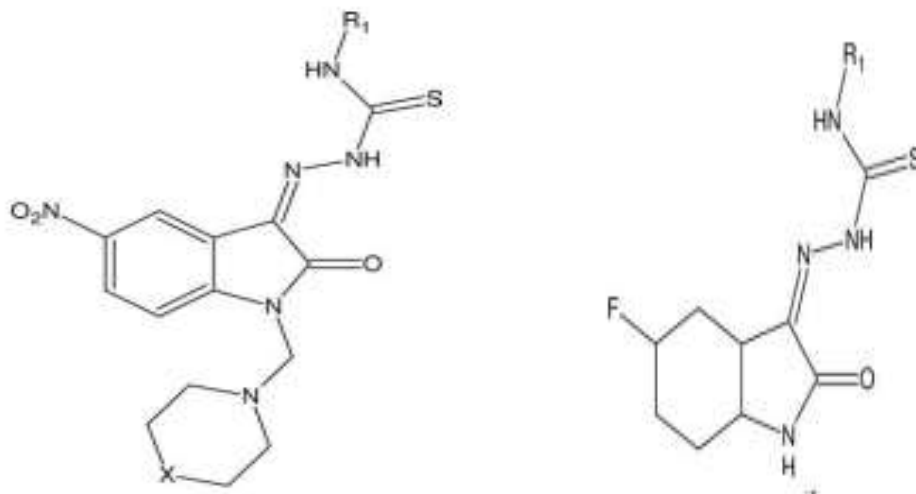
11. Andreas Burkovski (2003) ^[41] studied about the Ammonium assimilation and nitrogen control in *M. tuberculosis* with emphasis on the GSI enzyme, which has been identified as a potentially important determinant of pathogenicity.
12. David Eisenberg et al (1999) ^[42] studied about the highly regulated glutamine synthetase enzyme at the core of nitrogen metabolism. They studied about both bacterial and eukaryotic glutamine synthetases, with emphasis on enzymatic inhibitors.
13. Woolfolk and Stadtman (1967) ^[43] done the kinetic studies showed that the biosynthetic reaction catalyzed by *E.coli* GS is inhibited by nine end products of glutamine metabolism: serine, alanine, glycine, AMP,CTP, tryptophan, histidine, carbamoyl phosphate,and glucosamine-6-phosphate.

Review of literature regarding drug design:

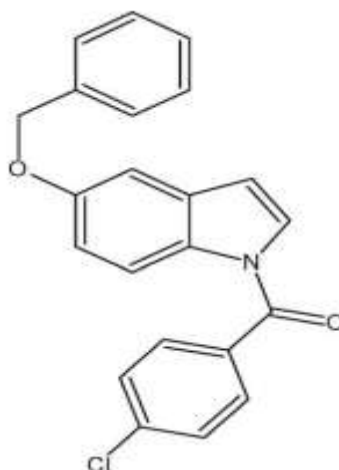
14. Pratik Swarup Das and Puja Saha (2017) ^[44] studied about the tools and techniques to assist in drug discovery process.
15. Wermuth C G., (2006) ^[45] reviewed the similarity in drugs with the importance and reflections on analogue design.
16. Laurie AT, Jackson RM (2006) ^[46] studied about the methods for the prediction of protein-ligand binding sites for the structure based drug design in virtual screening process.
17. Lipinski CA. et al. (2001) ^[47] reported the approaches to estimate the solubility and permeability in drug discovery experimentally and computationally.

Review of literature regarding indole nucleus:

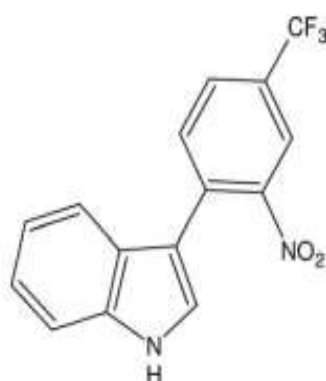
18. Amit K Singh et al(2013) ^[48] synthesized novel indole derivatives and evaluated for invitro anti-bacterial, anti fungal and anti-inflammatory activity.
19. Karali et al(2007) ^[49] synthesized a series of 1H-indole-2,3-dione derivatives and evaluated for in vitro antituberculosis activity against *Mycobacterium tuberculosis* H37Rv strains. Among the synthesized compounds 5-nitro-1H-indole-2,3-dione-3-thiosemicarbazones and its 1-morpholinomethyl derivatives exhibited significant inhibitory activity with MIC values 75%.



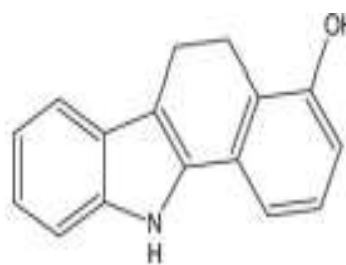
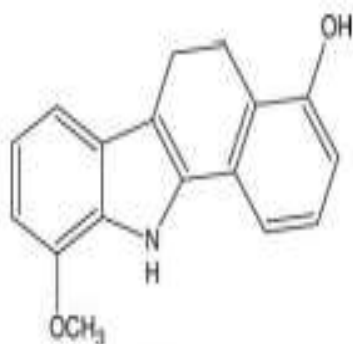
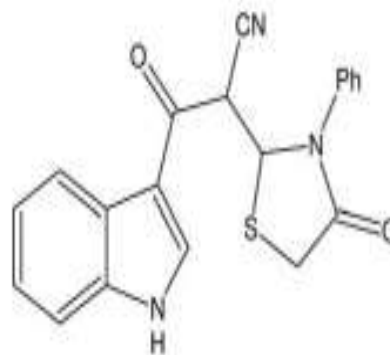
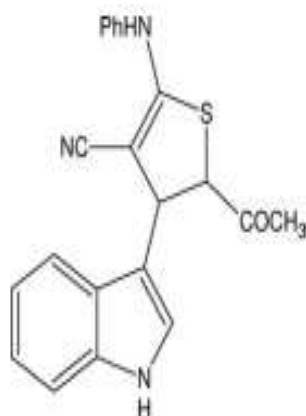
20. Li et al (2007) ^[50] synthesized some indole derivatives and the compounds were evaluated for their insulin sensitizing and glucose lowering effects. The indole derivatives showed good anti-diabetic activity.



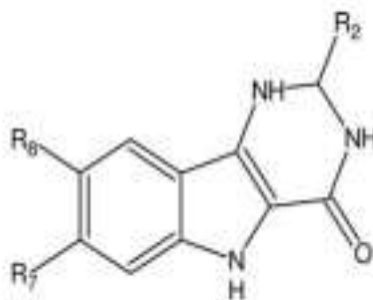
21. Hiari et al (2006) ^[51] synthesized some substituted 3-(aryl) and 3-(heteroaryl) indoles. The most active compound was reported to be 3-(4-trifluoromethyl-2-nitrophenyl) indole.



22. Hong et al (2006) ^[52] synthesized a series of tricyclic and tetracyclic indoles and evaluated for anticancer activity. the compounds found to exhibit highest in vitro activity against human nasopharyngeal carcinoma (HONE-1) and gastric adenocarcinoma (NUGC-3)
23. Enien et al (2004) ^[53] found that Indole-2 and 3-carboxamides having antioxidant properties.
24. Abele et al (2003) ^[54] synthesized indole oximes were found to be exhibiting high fungicidal activity. where the oxime derivates of 2-substituted indoles demonstrated significant antifungal activity.
25. Radwan et al (1997) ^[55] carried out the synthesis and biological evaluation of 3-substituted indole derivatives as potential anti-inflammatory and analgesic agents. They reported 3-(3-indolyl) thiophene derivative as apotent anti-inflammatory compound whereas thiazolidine-4-one derivative exhibit analgesic activity.

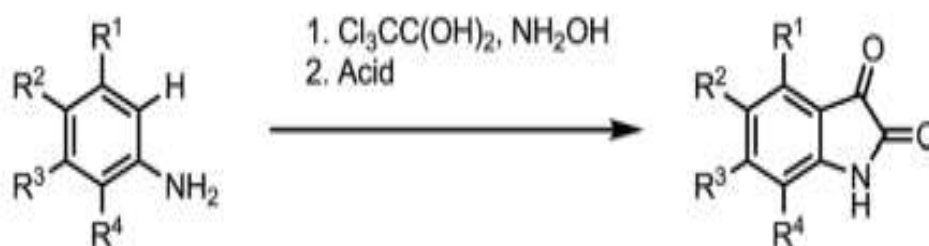


26. Merino et al(1999) ^[56] synthesized analogs of pyrimido [5,4-b] indoles and biologically evaluated for their possible HIV inhibitory activity.

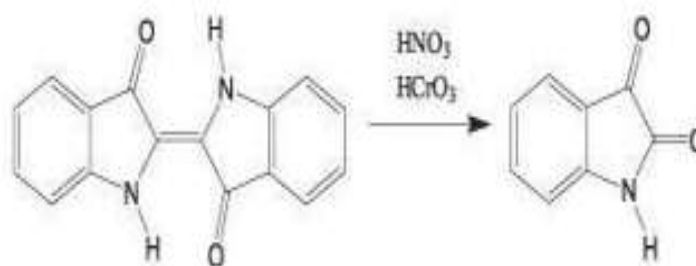


Review of literature regarding isatin derivatives:

27. Deng et al.,(2018) ^[57] synthesized a series of novel heteronuclear 5-fluoroisatin dimers tethered through ethylene and examined for their in vitro anti-mycobacterial activities against Mycobacterium tuberculosis H37Rv strains and multi-drug resistant tuberculosis (MDR-TB).
28. Parvanesh Pakravan et al.,(2013) ^[58] studied that Isatin derivatives has the ability to intercalate with DNA. Among them, Isatin-3-isonicotinyl hydrazone was found to be potentially capable of intercalation with DNA.
29. Verma et al.,(2003) ^[59] reported that Schiff bases of N-methyl and N-acetyl isatin derivatives with different aryl amines possess potent anticonvulsant activities.
30. Garden et al.,(1998) ^[60] described the synthesis of N-alkylated isatins from the respective isatins using calcium hydride and alkyl halide in DMF.
31. Pinto et al., (1994) ^[61] detailed that the synthesis of N-alkyl indoles under mild reaction conditions using N-acetyl isatins as substrates.
32. Sandmeyer et al., (1919) ^[62] developed the method for the synthesis of isatin. It consists of the reaction of aniline with chloral hydrate and hydroxylamine hydrochloride in aqueous sodium sulfate to form an isonitrosoacetanilide, which after isolation, when treated with concentrated sulfuric acid, gives isatin.



33. Erdmann et al.,(1840)^[63] synthesized 1H-indole-2,3-dione (Isatin) by reaction of indigo with chromic and nitric acids.



Review of literature regarding to the evaluation of anti TB activity:

34. Anitha G. et al (2009)^[64] described a rapid invitro method using Microplate Alamar Blue Assay to determine the Minimum Inhibitory Concentration for novel antimycobacterial agents.
35. Neetu kumar Taneja and Jaya Siwaswami Tyagi (2007)^[65] determined the MIC for M.tuberculosis using aerobic resazurin microplate assay(REMA) and correlated with those obtained by the Colony Forming Unit assay.
36. Juan-Carlos P et al,(2002)^[66] summarized the method for detecting multidrug-resistant Mycobacterium tuberculosis by using Resazurin Microtiter Assay Plate method (REMA).

AIM
AND
OBJECTIVE

AIM AND OBJECTIVES

AIM

The aim is to design and synthesize some novel heterocyclic analogues such as indole derivatives which will prove to be effective against *Mycobacterium tuberculosis*.

OBJECTIVES:

- Design of *Glutamine synthetase* inhibitors by docking studies using Autodock® software
- Prediction of *Insilico* Drug likeness by Molinspiration® software.
- *Insilico* Toxicity Assessment done by OSIRIS® software.
- Laboratory synthesis of chosen compounds with top Docking Scores.
- Characterization of the synthesized compounds by
 - TLC
 - Melting point
 - IR Spectroscopy
 - H1 NMR Spectroscopy
 - LC-Mass Spectrometry.
- Evaluation of *In-vitro* anti-tubercular activity of the synthesized compounds by (MABA).

PLAN OF WORK

Design of *Glutamine synthetase 1* inhibitors by using Autodock software.



In silico Drug likeness Prediction by Molinspiration[®]



In silico Toxicity prediction by OSIRIS[®]



Laboratory synthesis of the selected compounds



Justification of purity by TLC, Melting point and Characterization by IR, NMR, LC-MS



Evaluation of *invitro* anti -tubercular activity of synthesized compounds by MABA

MATERIALS
AND
METHODS

MATERIALS AND METHODS

The Project is carried out in the following phases.

- Drug design by using Autodock[®] software.
- Synthesis of the designed molecules.
- Characterization of the synthesized molecules.
- Biological evaluation of the synthesized molecules.

DRUG DESIGN

Drug design is referred to as rational drug design, which is an inventive process of finding newer drug molecules based on the knowledge of a biological target. [67] The drug is an organic small molecule that activates or inhibits the function of a biomolecule such as protein, which in turn results in a therapeutic benefit to the patient. Drug design involves the design of molecules that are complementary in shape and charge to the biomolecular target with which they interact and will bind to it. [68]

TYPES OF DRUG DESIGN

There are two major types of drug design.

- Ligand-based drug design
- Structure-based drug design

LIGAND-BASED [69]

Ligand-based drug design (or indirect drug design) depends upon the knowledge of other molecules that bind to the biological target of interest (protein). The other molecules may be used to derive a pharmacophore model that offers the minimum necessary structural characteristics a molecule must possess in order to bind to the target.

STRUCTURE-BASED ^[69]

Structure-based drug design (or direct drug design) depends upon the knowledge of the three dimensional structure of the biological target obtained through methods such as x-ray crystallography or NMR spectroscopy. If an experimental structure of a target is not available, it may be possible to create a homology model of the target based on the experimental structure of a related protein.

TARGET ENZYME: GLUTAMINE SYNTHETASE I ^[70-71]

The crystal structure of the enzyme was downloaded from the Protein Data Bank (An Information Portal to Biological Macromolecular Structures) (PDB id – 3zxr). The target enzyme *glutamine synthetase I* from *Mycobacterium tuberculosis*, is one of the key enzymes involved in GLUTAMINE SYNTHESIS, which is critical for the survival and growth of *Mycobacterium tuberculosis*.

BINDING SITE IDENTIFICATION ^[72-73]

Binding site identification is an important step in structure based drug design. Location of the binding site is trivial, if the structure of the target or a sufficiently similar homolog is determined in the presence of a bound ligand. However, there may be unoccupied allosteric binding sites that may be of interest. Furthermore, it may be only apoprotein (protein without ligand) structures are available and the reliable identification of unoccupied sites that have the potential to bind ligands with high affinity is non-trivial.

Molecular Docking by AUTODOCK [®]

AutoDock [®] 4.2.5.1 is a software for predicting the interaction of ligands with biomacromolecular targets. In any docking scheme, two conflicting requirements must be balanced: the desire for a robust. The current version of AutoDock, using the Lamarckian Genetic Algorithm and empirical free energy scoring function, typically will provide reproducible docking results for ligands with approximately 10 flexible bonds. The quality of any docking results depends on the starting structure of both the protein and the potential ligand. The protein and ligand structure need to be prepared to achieve the best docking results. The following steps are employed ^[74]

1. Protein preparation.
2. Ligand preparation .
3. Receptor grid generation.
4. Ligand docking (screening)

Preparation of Protein

- Read molecule from the file (allows reading of PDB coordinate files.)
- Edit -Charges – Compute Gasteiger (for arbitrary molecules)
- Edit – Hydrogen – Merge non polar.
- Save as .pdb in AutoDock folder.

Preparation of Ligand

- Ligand – Input from file
- Ligand – Torsion – choose torsion: Rotatable bonds are shown in green, and non-rotatable bonds are shown in red. Bonds that are potentially rotatable but treated as rigid, such as amide bonds and bonds that are made rigid by the user, are shown in magenta.
- Ligand – Torsion – set number of torsion: sets the number of rotatable bonds in the ligand by leaving the specified number of bonds as rotatable.
- Ligand – Output – save as pdbqt in AutoDock folder

Grid preparation

- Grid – Macromolecule -open (open the pdb file that has been saved and then save it in pdbqt extension in AutoDock folder)
- Grid – Set map types – open ligand : tools to define the atom types for the grids that will be calculated
- Grid – Grid box – launches interactive commands for setting the grid dimensions and center (Set dimension of 60: 60:60 – Center :center on macromolecule)
- File – Close saving current
- Grid – Output – save as .gpf (grid parameter file)
- Open command prompt [cd AutoDock

cd 4.2.5.1

autogrid4.exe -p a.gpf -l a.glg]

Preparation of Docking Parameters

- Docking –macromolecules – set rigid filename
- Docking – ligand –open
- Docking –search parameters – genetic algorithm parameters : this command open a panel for setting the parameters used by each of the search algorithms, such as temperature schedules in simulated annealing and mutation/crossover rates in genetic algorithms.
- Docking – docking parameters: opens a panel for setting the parameters used during the docking calculation, including options for the random number generator, options for the force field, step sizes taken when generating new conformations, and out put options.
- Docking- output –Lamarckian GA –save as .dpf (docking parameterfile)
- Open command prompt [autodock4.exe –p a.dpf –la.dlg]

Visualization / Interpretation of Docking

- Analysis –Docking – open .dlg (docking log file)file
- Analysis – macromolecule open
- Analysis – Confirmation –Play and Play ranked by energy : Play- will use the order of conformations as they were found in the docking calculations, and Play Ranked By Energy will order the conformations from lowest energy to highest energy.
- Analysis – Load : Information on the predicted interaction energy is shown at the top, and individual conformations
- Analysis – Docking – show interaction: specialized visualization to highlight interactions between the docked conformation of the ligand and the receptor.

INSILICO TOXICITY ASSESSMENT^[75]

OSIRIS[®]

- *Insilico* toxicity Assessment for the molecules were predicted by using OSIRIS[®], a JAVA based online tool.
- The tool predicts toxicity related parameters such as Mutagenicity, Tumorigenicity, Skin Irritancy and the effects on reproduction.

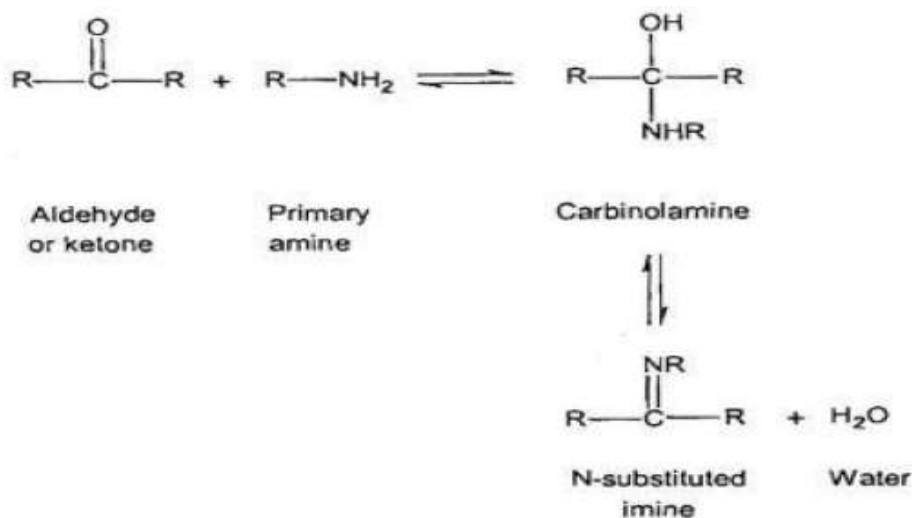
- The prediction is based the fragment contribution group present in the structure of the molecule.
- Properties with high risks of undesired effects are shown in red color. Whereas a green color indicates drug-conform behavior.

PREDICTION OF DRUG LIKENESS (MOLINSPIRATION®)

- The designed and docked molecules were screened *insilico* using MOLINSPIRATION® software to evaluate drug likeness.
- It is the online software available for the calculation of important molecular properties such as log P, polar surface area, number of hydrogen bond donors and acceptors., etc

SYNTHETIC SCHEME

The selected compounds with top docking score were selected for synthesis according to the scheme given below.



PROCEDURE

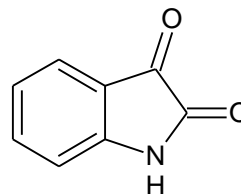
Equimolar quantities of ketone (0.01mol) and Para-substituted amine (0.01mol) are added into 20mL of absolute ethanol and 5mL of glacial acetic acid is added to it. Reaction mixture is refluxed for 24hrs at 60°C. Completion of reaction is confirmed by TLC. The product obtained was filtered and dried. Recrystallisation is done by using ethanol.

REACTANT PROFILE

The following ketone and primary amines were used for the synthesis.

KETONES

ISATIN:



Chemical formula : $C_8H_5NO_2$

Molecular weight : 148.13g/mol

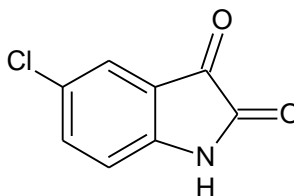
Appearance : Orange red solid

Melting point : 200°C

Solubility : dichloro methane, acetone

Boiling point : 360.3±52.0°C

5-CHLORO ISATIN:



Chemical formula : $C_8H_4NO_2Cl$

Molecular weight : 181.58 g/mol

Appearance : Yellow to Orange crystal

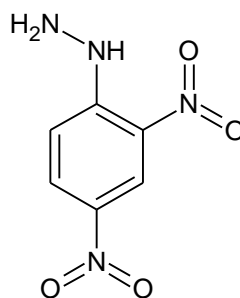
Melting point : 254-258°C

Solubility : hot water

Boiling point : 254-258°C

PRIMARY AMINES

2,4-DINITRO PHENYL HYDRAZINE



Chemical formula : C₆H₃(NO₂)₂NHNH₂

Molecular weight : 198.14 g/mol

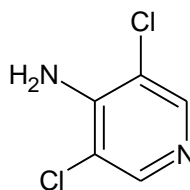
Appearance : Red to Orange crystal

Melting point : 198-202°C

Solubility : slightly soluble in water

Boiling point : 378.6±32.0°C

4-AMINO 3,5-DICHLORO PYRIDINE



Chemical formula : C₅H₄Cl₂N₂

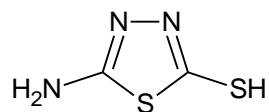
Molecular weight : 163 g/mol

Appearance : off white colour

Melting point : 159°C

Solubility : slightly soluble in methanol

5-AMINO 1,3,4-THIADIAZOLE-2-THIOL



Chemical formula : C₂H₃S₂N₃

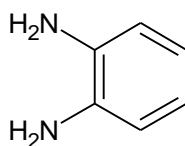
Molecular weight : 133.18 g/mol

Appearance : Crystalline white powder

Melting point : 235°C

Solubility : insoluble in water, soluble in DMSO, slightly soluble in methanol.

O-PHENYLENE DIAMINE



Chemical formula : C₆H₄(NH₂)₂

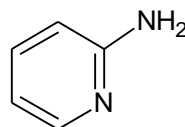
Molecular weight : 108.14 g/mol

Melting point : 102-104°C

Boiling point : 252°C

Solubility : soluble in hot water

2-AMINO PYRIDINE



Chemical formula : C₅H₆N₂

Molecular weight : 94.12 g/mol

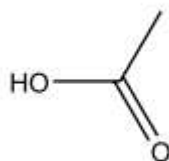
Appearance : colourless solid

Melting point : 59-60°C

Boiling point : 210°C

Solubility : slightly soluble in water.

GLACIAL ACETIC ACID:



Molecular Formula : C₂H₄ N₂

Molecular Weight : 60.05

Boiling Point : 390.87[K]

ETHANOL:



Molecular Formula : C₂H₆O

Molecular Weight : 46.07

Boiling Point : 337.54[K]

CHARACTERIZATION

PHYSICAL EVALUATION

1. Physical properties of the synthesized compounds are evaluated, such as
 - Color
 - Nature
 - Solubility
 - Molecular weight
 - Molecular formula
 - Melting point

2. Further the synthesized compounds are characterized by the following Spectroscopic and Spectrometric methods ^[76] such as
 - IR Spectroscopy by ABB MB 3000-PH FTIR spectrometer using KBr pellets.
 - ¹H-NMR Spectrometry by 500 MHZ BrukerTopSpin using DMSO
 - LC-MS by AGILANT technologies 6230B Time Of Flight(TOF)

IR SPECTROSCOPY ^[77]

The absorption of infrared radiations can be expressed in terms of wavelength or in wavenumber. The infrared region ranges from wavenumber 4000cm⁻¹ to 667cm⁻¹. The bonds in a molecule which are accompanied by a change in dipole movement will absorb IR radiation. Ex. C=O, N-H, O-H. The identity of an organic compound can be established from its finger print region(1400-900 cm⁻¹). Alkali metal halides such as KBr or NaCl which is transparent to IR region is used for sampling of solids.

The possible characteristic bands expected for the compounds to be synthesized are

- 3540-3300 cm-1 N-H StretchingVibration
- 3670-3230 cm-1 O-H StretchingVibration
- 0-1630 cm-1 C=N StretchingVibration
- 2975-2840 cm-1 C-H Aliphatic StretchingVibration
- 3100-3000 cm-1 C-H Aromatic StretchingVibration

NMR SPECTROMETRY ^[78]

NMR spectrometry involves the change in spin state of a nuclear magnetic moment when the nucleus absorbs electromagnetic radiation in a strong magnetic field. By using ¹H-NMR Spectroscopy we can get the following information. They are,

1. The relationship between the number of signals or peaks in the spectrum and the number of different kinds of hydrogen atoms in the molecule. Thus we can know the different kinds of environment of the hydrogen atom in the molecule.
2. The areas underneath each signal are in the same ratio as the number of hydrogen atoms causing each signal.

3. The principal signal may get split into smaller peaks, such as spin-spin splitting may be observed. The type of splitting depends upon the number of neighbouring non-equivalent protons.
4. The spacing between the peaks gives information on molecular structure and stereochemical features.

The basic peaks expected for the compounds to be synthesized are,

- Aromatic and hetero aromatic compounds 6-8.5 δ
- Alcoholic hydroxyl protons 1-5.5 δ
- Aldehyde protons 9-10 δ

LC-MS

LC-MS is a hyphenated technique, coupling the separation power of HPLC, with the detection power of Mass spectrometry. When the sample cannot be vapourised, GC-MS cannot be performed. So we can prefer LC-MS.

EVALUATION OF ANTI TUBERCULAR ACTIVITY

MICROPLATE ALAMAR BLUE ASSAY (MABA) ^[79] is performed to evaluate the *invitro* anti tubercular activity.

PROCEDURE ^[80-81]

- *The anti mycobacterial activity of compounds were assessed against M. tuberculosis using microplate Alamar Blue assay (MABA).*
- This methodology is non-toxic, uses a thermally stable reagent and shows good correlation with proportional and BACTEC radiometric method.
- Briefly, 200 μ l of sterile de-ionized water was added to all outer perimeter wells of sterile 96 wells plate to minimized evaporation of medium in the test wells during incubation.
- The 96 wells plate received 100 μ l of the Middle brook 7H9 broth and serial dilution of compounds were made directly on plate.
- The final drug concentrations tested were 100 to 0.2 μ g/ml. Plates were covered and sealed with parafilm and incubated at 37°C for five days.

- After this time, 25 μ l of freshly prepared 1:1 mixture of Almar Blue reagent and 10% tween 80 was added to the plate and incubated for 24hrs.
- A blue color in the well was interpreted as no bacterial growth, and pink color was scored as growth. The MIC was defined as lowest drug concentration which prevented the color change from blue to pink.

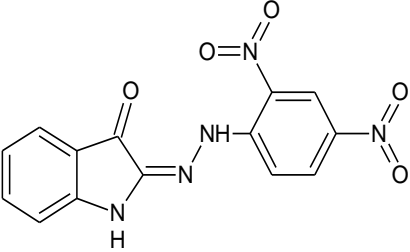
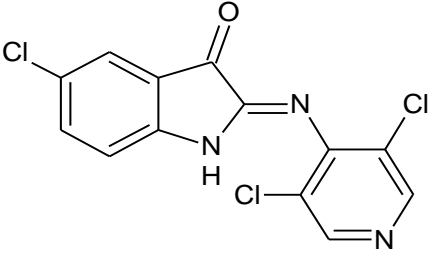
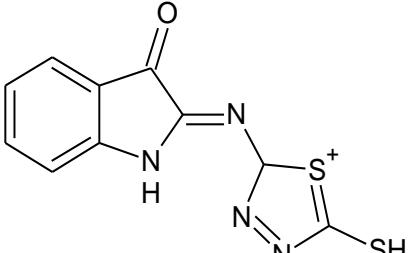
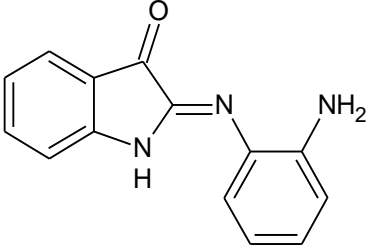
RESULTS
AND
DISCUSSION

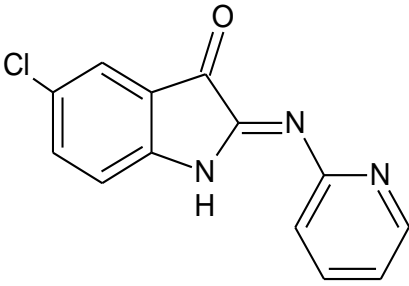
RESULTS AND DISCUSSION

ACTIVITY PREDICTION :

More than 150 compounds were docked against the enzyme Glutamine synthetase I using AUTODOCK[®] tools 4.2.5.1 software. The molecules with good docking score and good interactions were synthesized and characterized.

Table 1 : the selected molecules with docking score are mentioned below:

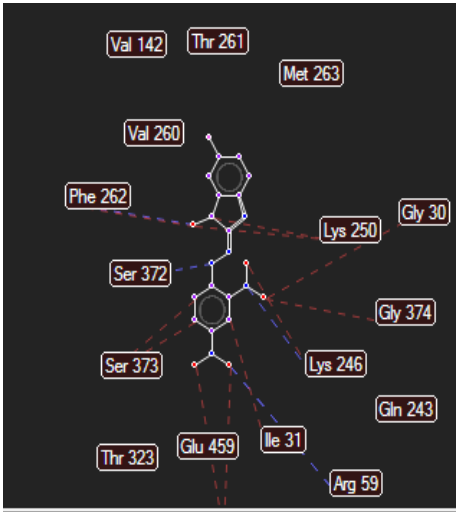
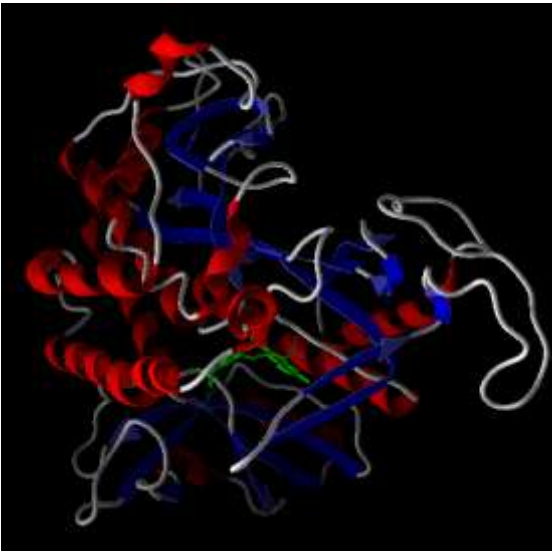
| SAMPLE CODE | STRUCTURE | DOCKING SCORE |
|-------------|--|---------------|
| PK1 |  | -7.56 |
| PK2 |  | -7.1 |
| PK3 |  | -6.57 |
| PK4 |  | -5.83 |

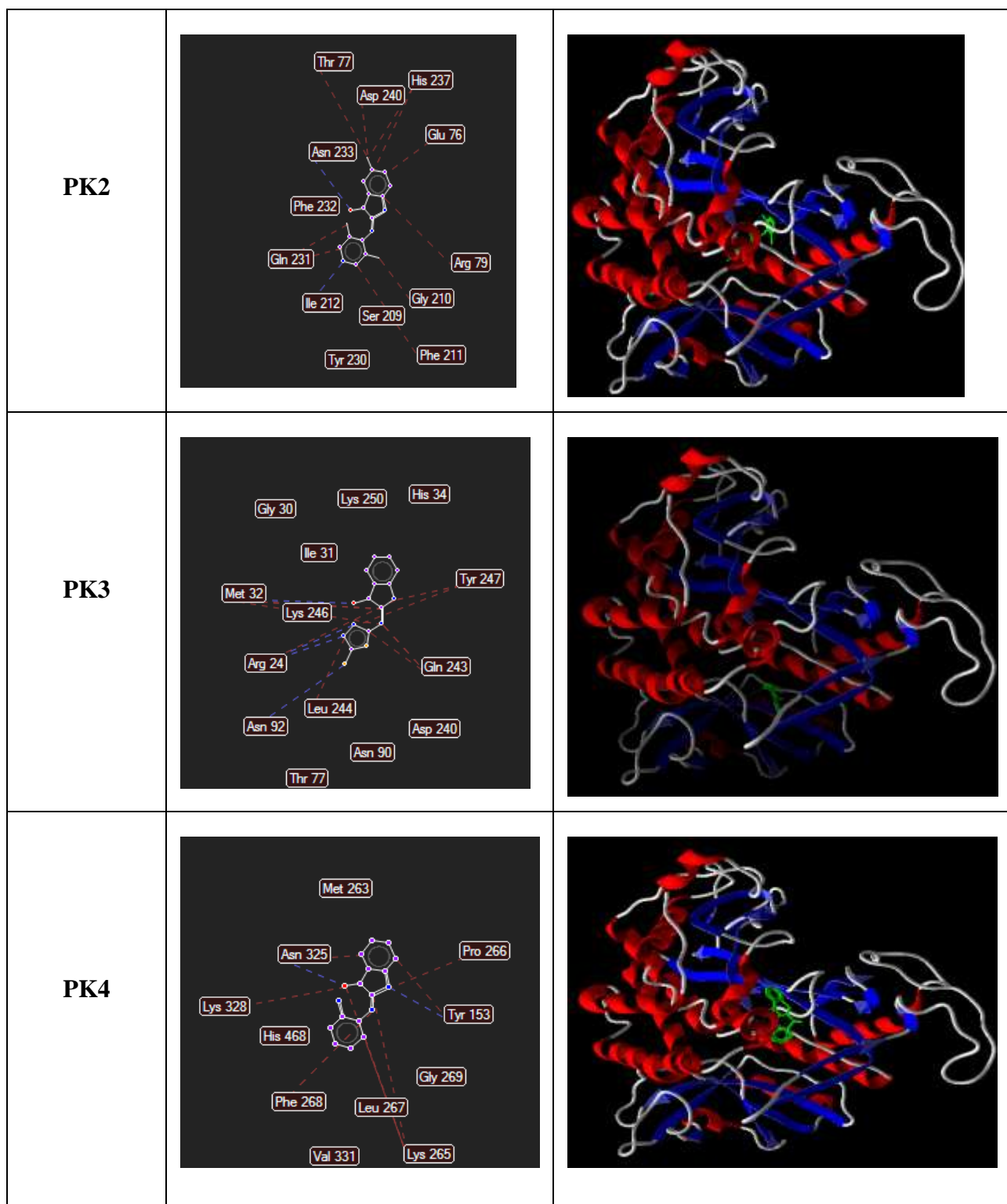
| | | |
|-------------------|---|-------------|
| <p>PK5</p> |  | <p>-5.7</p> |
|-------------------|---|-------------|

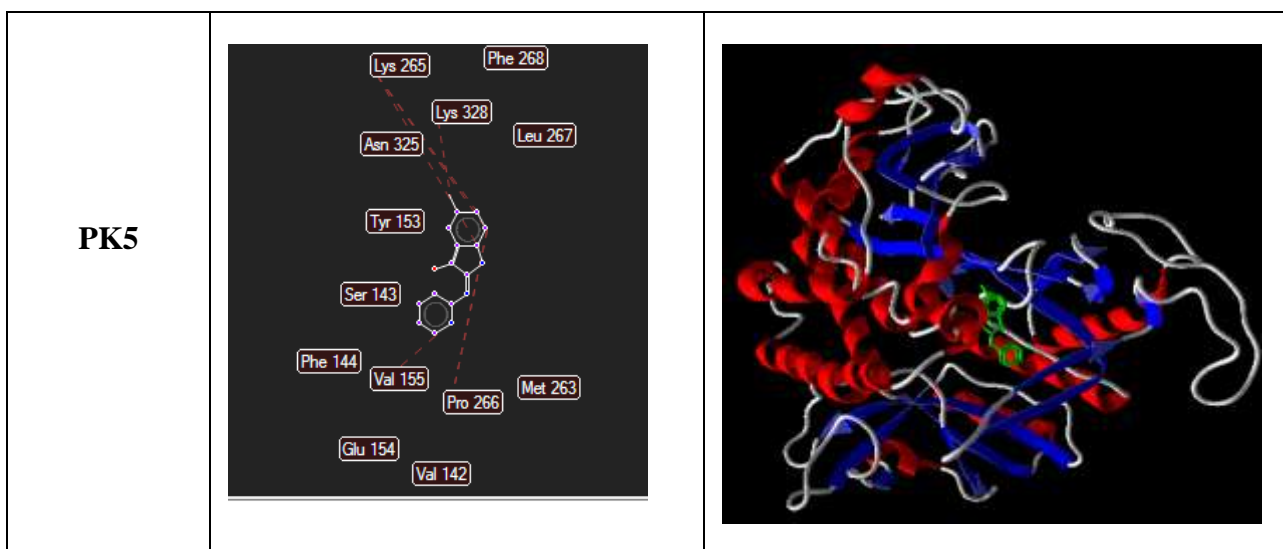
DOCKING VIEW AND INTERACTION OF THE DOCKED MOLECULES WITH THE AMINOACIDS:

AUTODOCK[®] 4.2.5.1 tools performs a complete systemic search of the conformations, orientations and position of a compound in the defined binding site and eliminates unwanted poses using scoring and energy optimization. The best poses were selected on the basis of the scoring function and the quality of pose orientation within the active site of the aminoacids

Table 2 : Interaction of the molecules with amino acids and docking view

| <p>SAMPLE CODE</p> | <p>INTERACTION WITH THE AMINO ACID</p> | <p>DOCKING VIEW</p> |
|---------------------------|---|--|
| <p>PK1</p> |  |  |








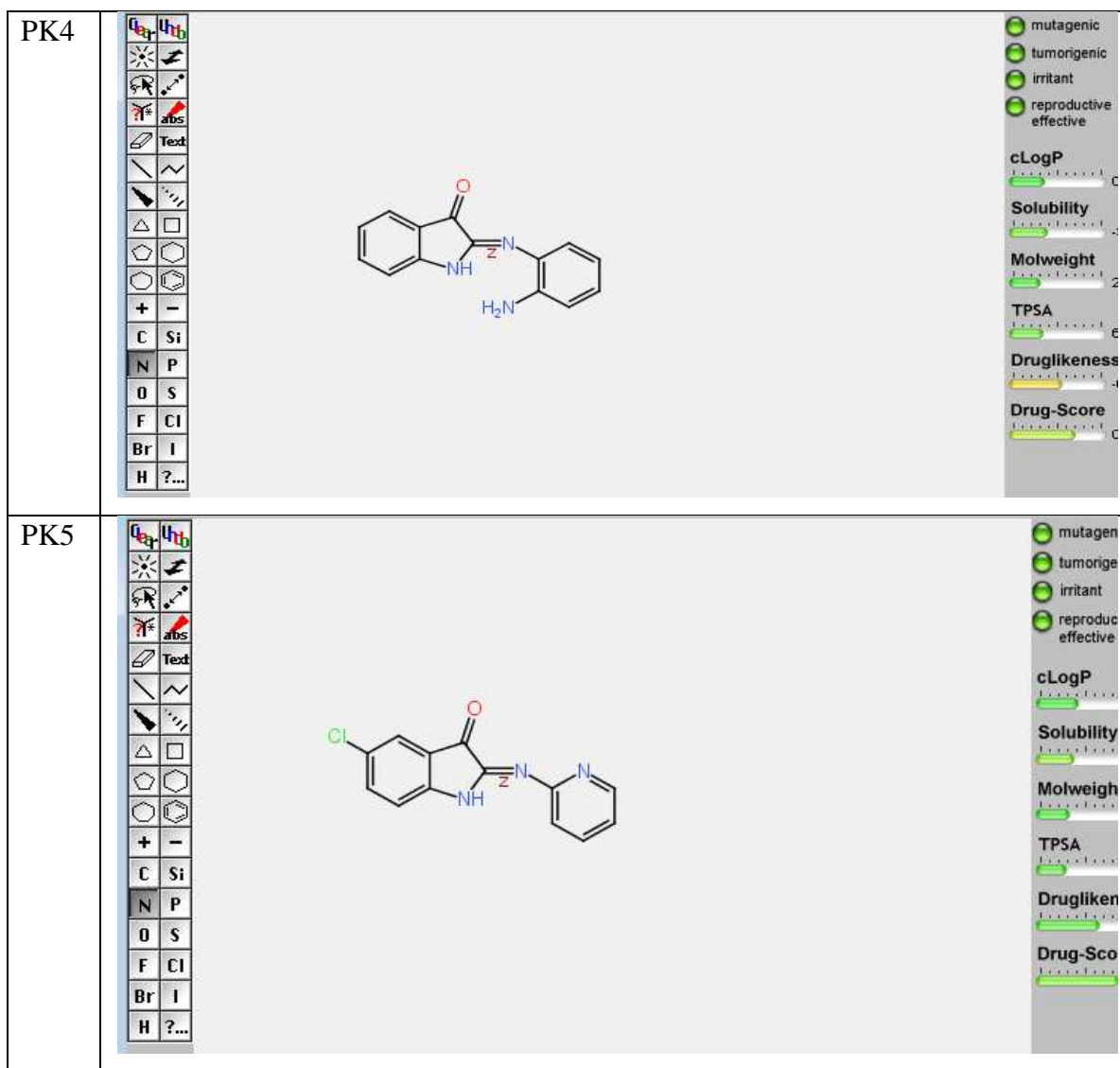
VARIANTS:

PREDICTION OF DRUG TOXICITY (INSILICO):

- *In-silico* toxicity assessment for the chosen molecules were predicted by using OSIRIS[®], a JAVA based online tool.
- The tool predicts toxicity related parameters such as Mutagenicity, Tumorigenicity, Skin Irritancy and Teratogenicity apart from other toxicities.
- The prediction is based on the fragment contribution group present in the structure of the molecule.
- Properties with high risks or undesired effects are shown in red color. Green colour shows drug conform behavior.

Table 3 : The following are the results of the toxicity prediction for five selected molecules based on docking score:

| | | |
|------------|---|--|
| <p>PK1</p> |  | <ul style="list-style-type: none"> <input type="checkbox"/> mutagenic <input type="checkbox"/> tumorigenic <input type="checkbox"/> irritant <input type="checkbox"/> reproductive effective <p>cLogP: 1.62</p> <p>Solubility: -4.32</p> <p>Molweight: 327.1</p> <p>TPSA: 146.1</p> <p>Druglikeness: -7.39</p> <p>Drug-Score: 0.39</p> |
| <p>PK2</p> |  | <ul style="list-style-type: none"> <input type="checkbox"/> mutagenic <input type="checkbox"/> tumorigenic <input type="checkbox"/> irritant <input type="checkbox"/> reproductive effective <p>cLogP: 2.48</p> <p>Solubility: -4.82</p> <p>Molweight: 325.0</p> <p>TPSA: 54.35</p> <p>Druglikeness: 1.89</p> <p>Drug-Score: 0.66</p> |
| <p>PK3</p> |  | <ul style="list-style-type: none"> <input type="checkbox"/> mutagenic <input type="checkbox"/> tumorigenic <input type="checkbox"/> irritant <input type="checkbox"/> reproductive effective <p>cLogP: [unreadable]</p> <p>Solubility: [unreadable]</p> <p>Molweight: [unreadable]</p> <p>TPSA: [unreadable]</p> <p>Druglikeness: [unreadable]</p> <p>Drug-Score: [unreadable]</p> |



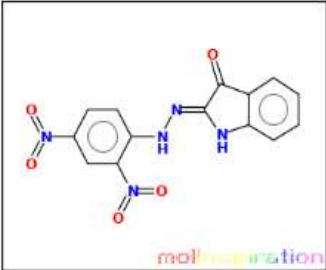
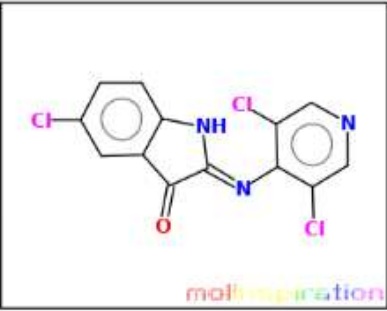
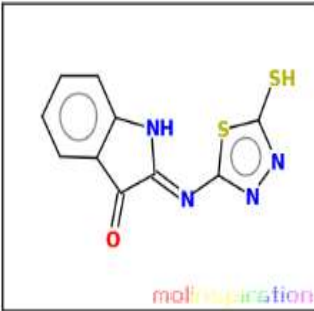
PREDICTION OF DRUGLIKENESS:

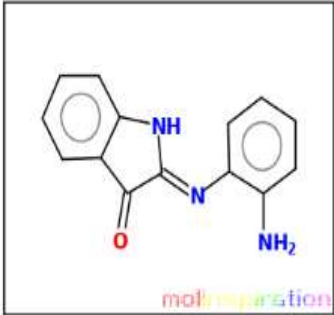
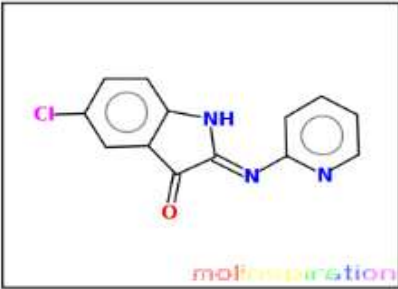
The designed molecules were screened *insilico* using MOLINSPIRATION[®] software to evaluate drug likeness. It is an online software available for the calculation of important molecular properties such as log P, polar surface area, number of hydrogen bond donors acceptors and number of rotatable bonds.

In an attempt to improve the predictions of drug likeness, the rules have spawned many extensions. They are given below:

- Partition coefficient log P, range from -0.4 to +5.6
- Molar refractivity from 40 to 130
- Molecular weight from 180 to 500 daltons.
- Number of atoms from 20 to 70
- Not more than 5 hydrogen bond donors and 10 hydrogen bond acceptors

Table 4 : results of drug likeness prediction by molinspiration

| | | | | | | | | | | | | | | | | | | | |
|-----------------------------|---|------------------------|------|----------------------|--------|------------------------|----|--------------------|--------|---------------------|----|-----------------------|---|-----------------------------|---|-----------------------|---|------------------------|--------|
| PK1 | <p>molinspiration</p> <p>miSMILES: <chem>O=c2c(=NNc1ccc(N(=O)=O)cc1N(=O)=O)[nH]c3ccccc23</chem></p>  <p>Molinspiration property engine v2018.10</p> <table border="0"> <tr><td>miLogP</td><td>2.84</td></tr> <tr><td>TPSA</td><td>148.90</td></tr> <tr><td>natoms</td><td>24</td></tr> <tr><td>Mw</td><td>327.26</td></tr> <tr><td>nON</td><td>10</td></tr> <tr><td>nOHNH</td><td>2</td></tr> <tr><td>nviolations</td><td>0</td></tr> <tr><td>nrotb</td><td>4</td></tr> <tr><td>volume</td><td>258.33</td></tr> </table> | miLogP | 2.84 | TPSA | 148.90 | natoms | 24 | Mw | 327.26 | nON | 10 | nOHNH | 2 | nviolations | 0 | nrotb | 4 | volume | 258.33 |
| miLogP | 2.84 | | | | | | | | | | | | | | | | | | |
| TPSA | 148.90 | | | | | | | | | | | | | | | | | | |
| natoms | 24 | | | | | | | | | | | | | | | | | | |
| Mw | 327.26 | | | | | | | | | | | | | | | | | | |
| nON | 10 | | | | | | | | | | | | | | | | | | |
| nOHNH | 2 | | | | | | | | | | | | | | | | | | |
| nviolations | 0 | | | | | | | | | | | | | | | | | | |
| nrotb | 4 | | | | | | | | | | | | | | | | | | |
| volume | 258.33 | | | | | | | | | | | | | | | | | | |
| PK2 | <p>molinspiration</p> <p>miSMILES: <chem>O=c2c(=Nc1c(Cl)cncc1Cl)[nH]c3ccc(Cl)cc23</chem></p>  <p>Molinspiration property engine v2018.10</p> <table border="0"> <tr><td>miLogP</td><td>4.25</td></tr> <tr><td>TPSA</td><td>58.12</td></tr> <tr><td>natoms</td><td>20</td></tr> <tr><td>Mw</td><td>326.57</td></tr> <tr><td>nON</td><td>4</td></tr> <tr><td>nOHNH</td><td>1</td></tr> <tr><td>nviolations</td><td>0</td></tr> <tr><td>nrotb</td><td>1</td></tr> <tr><td>volume</td><td>235.71</td></tr> </table> | miLogP | 4.25 | TPSA | 58.12 | natoms | 20 | Mw | 326.57 | nON | 4 | nOHNH | 1 | nviolations | 0 | nrotb | 1 | volume | 235.71 |
| miLogP | 4.25 | | | | | | | | | | | | | | | | | | |
| TPSA | 58.12 | | | | | | | | | | | | | | | | | | |
| natoms | 20 | | | | | | | | | | | | | | | | | | |
| Mw | 326.57 | | | | | | | | | | | | | | | | | | |
| nON | 4 | | | | | | | | | | | | | | | | | | |
| nOHNH | 1 | | | | | | | | | | | | | | | | | | |
| nviolations | 0 | | | | | | | | | | | | | | | | | | |
| nrotb | 1 | | | | | | | | | | | | | | | | | | |
| volume | 235.71 | | | | | | | | | | | | | | | | | | |
| PK3 | <p>molinspiration</p> <p>miSMILES: <chem>O=c2c(=Nc1nnc(S)s1)[nH]c3ccccc23</chem></p>  <p>Molinspiration property engine v2018.10</p> <table border="0"> <tr><td>miLogP</td><td>2.47</td></tr> <tr><td>TPSA</td><td>71.01</td></tr> <tr><td>natoms</td><td>17</td></tr> <tr><td>Mw</td><td>262.32</td></tr> <tr><td>nON</td><td>5</td></tr> <tr><td>nOHNH</td><td>1</td></tr> <tr><td>nviolations</td><td>0</td></tr> <tr><td>nrotb</td><td>1</td></tr> <tr><td>volume</td><td>199.32</td></tr> </table> | miLogP | 2.47 | TPSA | 71.01 | natoms | 17 | Mw | 262.32 | nON | 5 | nOHNH | 1 | nviolations | 0 | nrotb | 1 | volume | 199.32 |
| miLogP | 2.47 | | | | | | | | | | | | | | | | | | |
| TPSA | 71.01 | | | | | | | | | | | | | | | | | | |
| natoms | 17 | | | | | | | | | | | | | | | | | | |
| Mw | 262.32 | | | | | | | | | | | | | | | | | | |
| nON | 5 | | | | | | | | | | | | | | | | | | |
| nOHNH | 1 | | | | | | | | | | | | | | | | | | |
| nviolations | 0 | | | | | | | | | | | | | | | | | | |
| nrotb | 1 | | | | | | | | | | | | | | | | | | |
| volume | 199.32 | | | | | | | | | | | | | | | | | | |

| | | | | | | | | | | | | | | | | | | | |
|-----------------------------|--|------------------------|------|----------------------|-------|------------------------|----|--------------------|--------|---------------------|---|-----------------------|---|-----------------------------|---|-----------------------|---|------------------------|--------|
| PK4 | <p>molinspiration</p> <p>miSMILES: <chem>Nc1ccccc1N=c3[nH]c2ccccc2c3=O</chem></p> <div style="display: flex; justify-content: space-between;"> <div style="text-align: center;">  <p>molinspiration</p> </div> <div> <p>Molinspiration_property_engine v2018.10</p> <table border="0"> <tr><td>miLogP</td><td>2.67</td></tr> <tr><td>TPSA</td><td>71.25</td></tr> <tr><td>natoms</td><td>18</td></tr> <tr><td>Mw</td><td>237.26</td></tr> <tr><td>nON</td><td>4</td></tr> <tr><td>nOHNH</td><td>3</td></tr> <tr><td>nviolations</td><td>0</td></tr> <tr><td>nrotb</td><td>1</td></tr> <tr><td>volume</td><td>210.54</td></tr> </table> </div> </div> | miLogP | 2.67 | TPSA | 71.25 | natoms | 18 | Mw | 237.26 | nON | 4 | nOHNH | 3 | nviolations | 0 | nrotb | 1 | volume | 210.54 |
| miLogP | 2.67 | | | | | | | | | | | | | | | | | | |
| TPSA | 71.25 | | | | | | | | | | | | | | | | | | |
| natoms | 18 | | | | | | | | | | | | | | | | | | |
| Mw | 237.26 | | | | | | | | | | | | | | | | | | |
| nON | 4 | | | | | | | | | | | | | | | | | | |
| nOHNH | 3 | | | | | | | | | | | | | | | | | | |
| nviolations | 0 | | | | | | | | | | | | | | | | | | |
| nrotb | 1 | | | | | | | | | | | | | | | | | | |
| volume | 210.54 | | | | | | | | | | | | | | | | | | |
| PK5 | <p>molinspiration</p> <p>miSMILES: <chem>O=c2c(=Nc1cccn1)[nH]c3ccc(Cl)cc23</chem></p> <div style="display: flex; justify-content: space-between;"> <div style="text-align: center;">  <p>molinspiration</p> </div> <div> <p>Molinspiration_property_engine v2018.10</p> <table border="0"> <tr><td>miLogP</td><td>2.99</td></tr> <tr><td>TPSA</td><td>58.12</td></tr> <tr><td>natoms</td><td>18</td></tr> <tr><td>Mw</td><td>257.68</td></tr> <tr><td>nON</td><td>4</td></tr> <tr><td>nOHNH</td><td>1</td></tr> <tr><td>nviolations</td><td>0</td></tr> <tr><td>nrotb</td><td>1</td></tr> <tr><td>volume</td><td>208.64</td></tr> </table> </div> </div> | miLogP | 2.99 | TPSA | 58.12 | natoms | 18 | Mw | 257.68 | nON | 4 | nOHNH | 1 | nviolations | 0 | nrotb | 1 | volume | 208.64 |
| miLogP | 2.99 | | | | | | | | | | | | | | | | | | |
| TPSA | 58.12 | | | | | | | | | | | | | | | | | | |
| natoms | 18 | | | | | | | | | | | | | | | | | | |
| Mw | 257.68 | | | | | | | | | | | | | | | | | | |
| nON | 4 | | | | | | | | | | | | | | | | | | |
| nOHNH | 1 | | | | | | | | | | | | | | | | | | |
| nviolations | 0 | | | | | | | | | | | | | | | | | | |
| nrotb | 1 | | | | | | | | | | | | | | | | | | |
| volume | 208.64 | | | | | | | | | | | | | | | | | | |

CHARACTERIZATION:

The selected compounds were synthesized, recrystallised and purified by using ethanol .

JUSTIFICATION OF PURITY OF SYNTHESIZED COMPOUNDS:**Melting point:**

The melting points of the synthesized compound were determined by one end open capillary method. Sharp melting point indicated that the synthesized compounds were pure.

TLC:

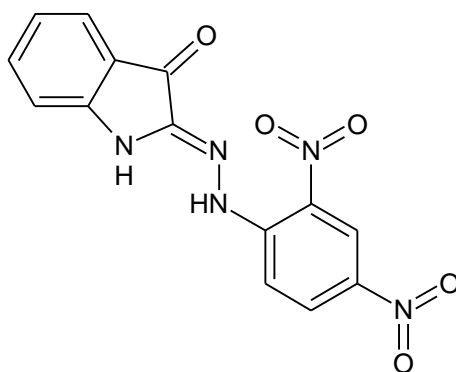
- Precoated aluminum TLC plates were used. Solutions of the reactants and products were prepared by dissolving them in methanol.
- Appearance of a single spot not corresponding to the parent compounds confirms the purity of the synthesized Compounds.
- Rf values of the synthesized compounds varies from the parent compounds indicates that the reaction was completed.

Table 5 : Rf values of the synthesized compounds:

| S.NO | COMPOUND CODE | MOBILE PHASE | Rf VALUE |
|------|------------------|--------------------------|----------|
| 1 | PK1 | HEXANE:ETHYLACETATE(7:3) | 0.63 |
| 2 | PK2 | HEXANE:ETHYLACETATE(7:3) | 0.71 |
| 3 | PK3 | HEXANE:ETHYLACETATE(7:3) | 0.78 |
| 4 | PK4 | HEXANE:ETHYLACETATE(7:3) | 0.69 |
| 5 | PK5 | HEXANE:ETHYLACETATE(7:3) | 0.75 |

PRODUCT PROFILE:

PK1



(2Z)-2-[2-(2,4-dinitrophenyl)hydrazinylidene]-1,2-dihydro-3H-indol-3-one

| | |
|---------------------|--|
| Molecular Formula | = C ₁₄ H ₉ N ₅ O ₅ |
| Formula Weight | = 327.25176 |
| Composition | = C(51.38%) H(2.77%) N(21.40%) O(24.45%) |
| Molar Refractivity | = 80.75 ± 0.5 cm ³ |
| Molar Volume | = 194.5 ± 7.0 cm ³ |
| Parachor | = 586.3 ± 8.0 cm ³ |
| Index of Refraction | = 1.768 ± 0.05 |
| Surface Tension | = 82.4 ± 7.0 dyne/cm |
| Density | = 1.68 ± 0.1 g/cm ³ |
| Dielectric Constant | = Not available |
| Polarizability | = 32.01 ± 0.5 10 ⁻²⁴ cm ³ |

IR SPECTRUM OF THE COMPOUND PK1

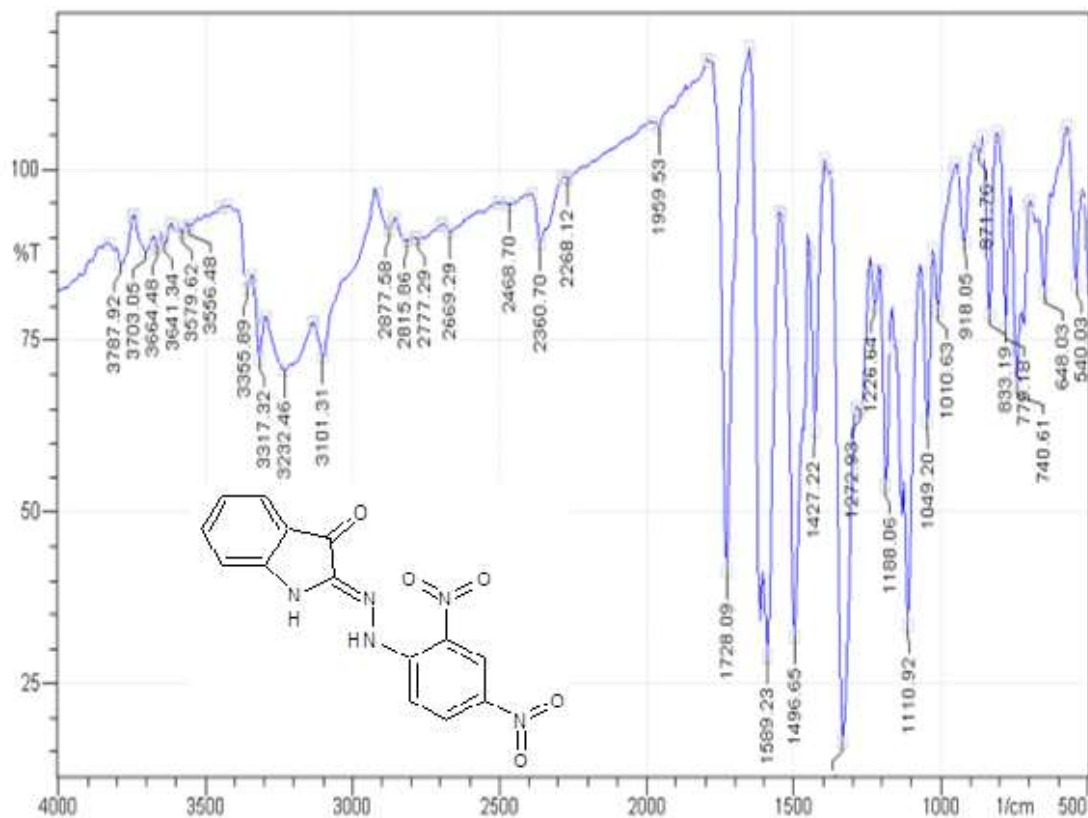
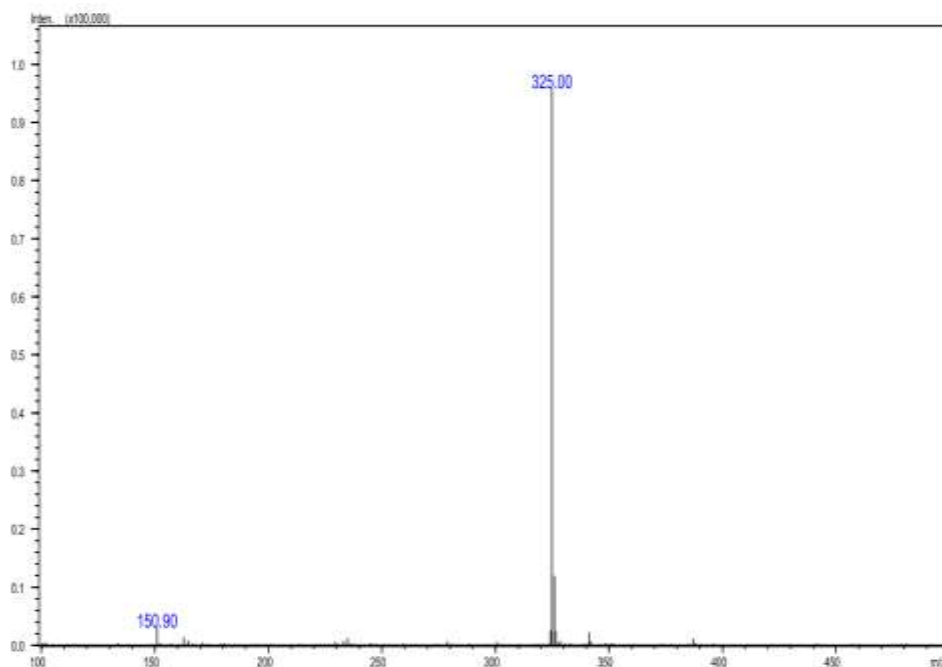
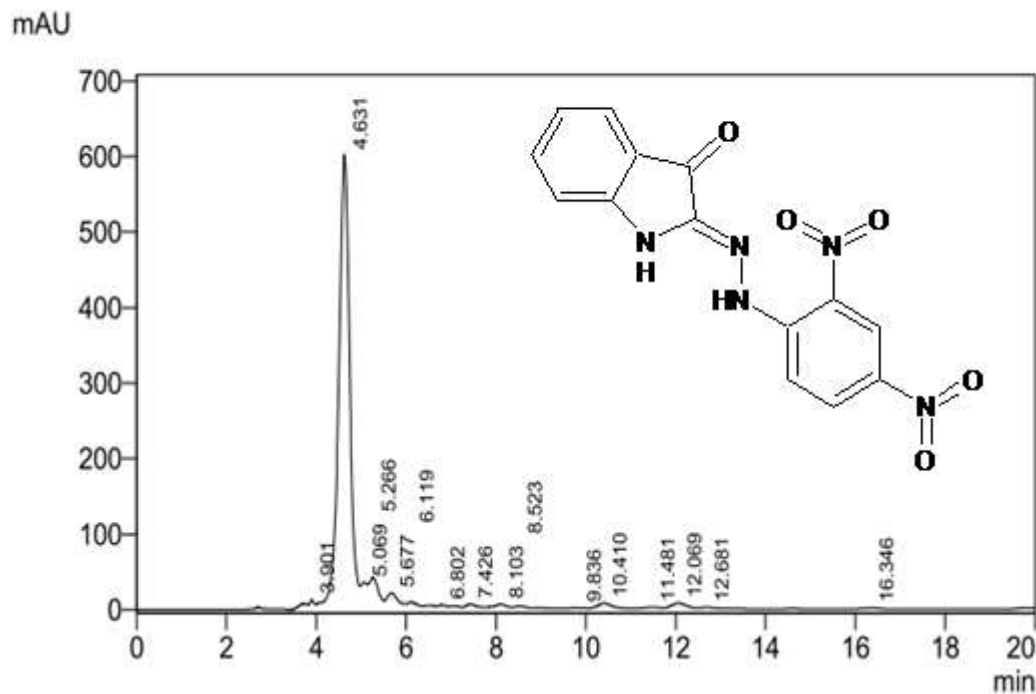


Table 6: IR interpretation of the compound PK1

| S.NO | WAVE NUMBER (cm ⁻¹) | TYPES OF VIBRATIONS | FUNCTIONAL GROUP |
|------|------------------------------------|----------------------------|-------------------------------------|
| 1 | 2885 | C-H Stretching | Presence of alkyl group |
| 2 | 1496.65 | NO ₂ Stretching | Presence of aromatic nitro group |
| 3 | 1728.09 | C=O Stretching | Presence of keto group |

LC-MS SPECTRUM OF THE COMPOUND PK1 (molecular weight:327.26)



NMR SPECTRUM OF COMPOUND PK1

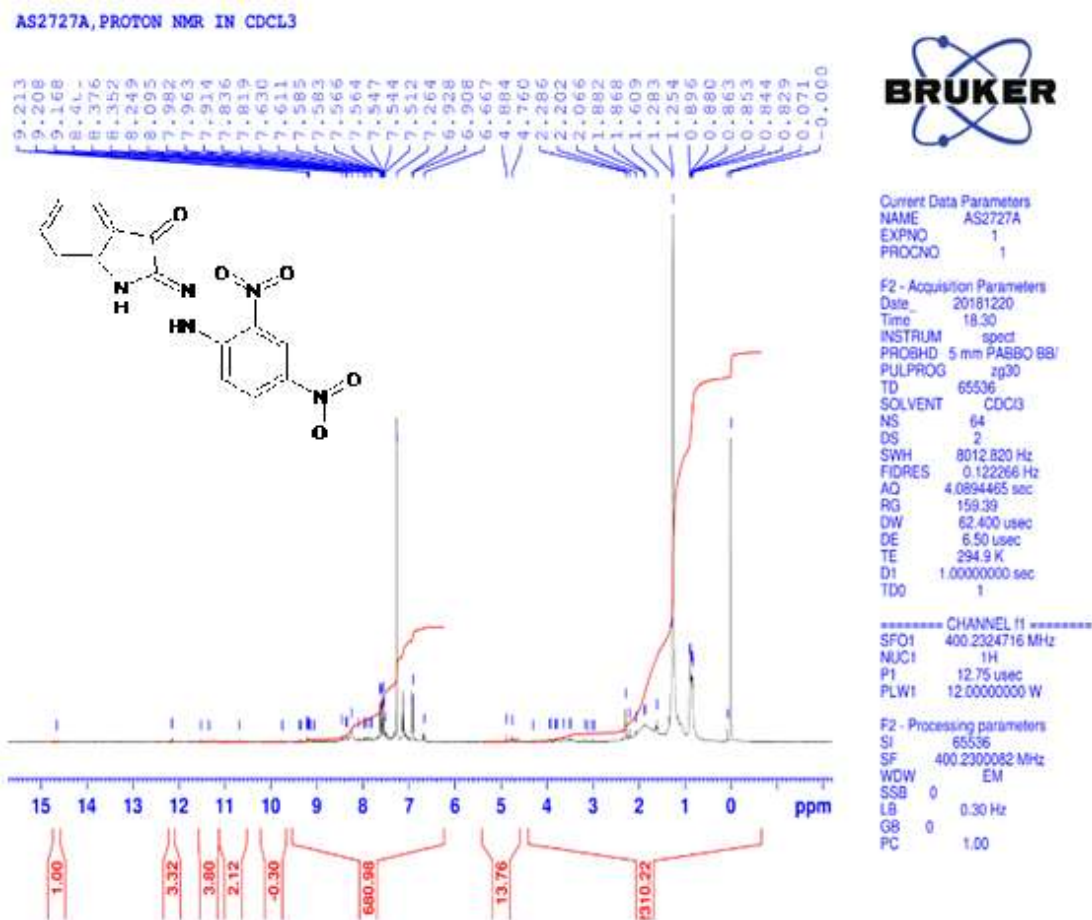
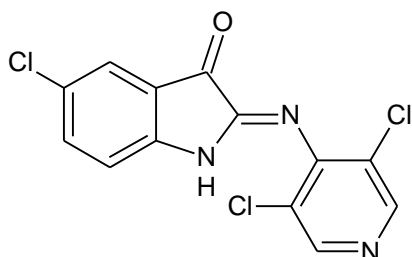


Table 7: NMR interpretation of compound PK1

| S.NO | δ VALUES | TYPES OF PEAK | NUMBER OF PROTONS |
|------|-----------------|---------------|-------------------|
| 1 | 7.6-7.8 | Multiplet | Five protons |
| 2 | 7.9-8.4 | Multiplet | Four protons |

PK2



| | |
|---------------------|---|
| Molecular Formula | = C ₁₃ H ₆ Cl ₃ N ₃ O |
| Formula Weight | = 326.56524 |
| Composition | = C(47.81%) H(1.85%) Cl(32.57%) N(12.87%) O(4.90%) |
| Molar Refractivity | = 78.62 ± 0.5 cm ³ |
| Molar Volume | = 195.7 ± 7.0 cm ³ |
| Parachor | = 543.3 ± 8.0 cm ³ |
| Index of Refraction | = 1.736 ± 0.05 |
| Surface Tension | = 59.4 ± 7.0 dyne/cm |
| Density | = 1.66 ± 0.1 g/cm ³ |
| Dielectric Constant | = Not available |
| Polarizability | = 31.16 ± 0.5 10 ⁻²⁴ cm ³ |

IR SPECTRUM OF THE COMPOUND PK2

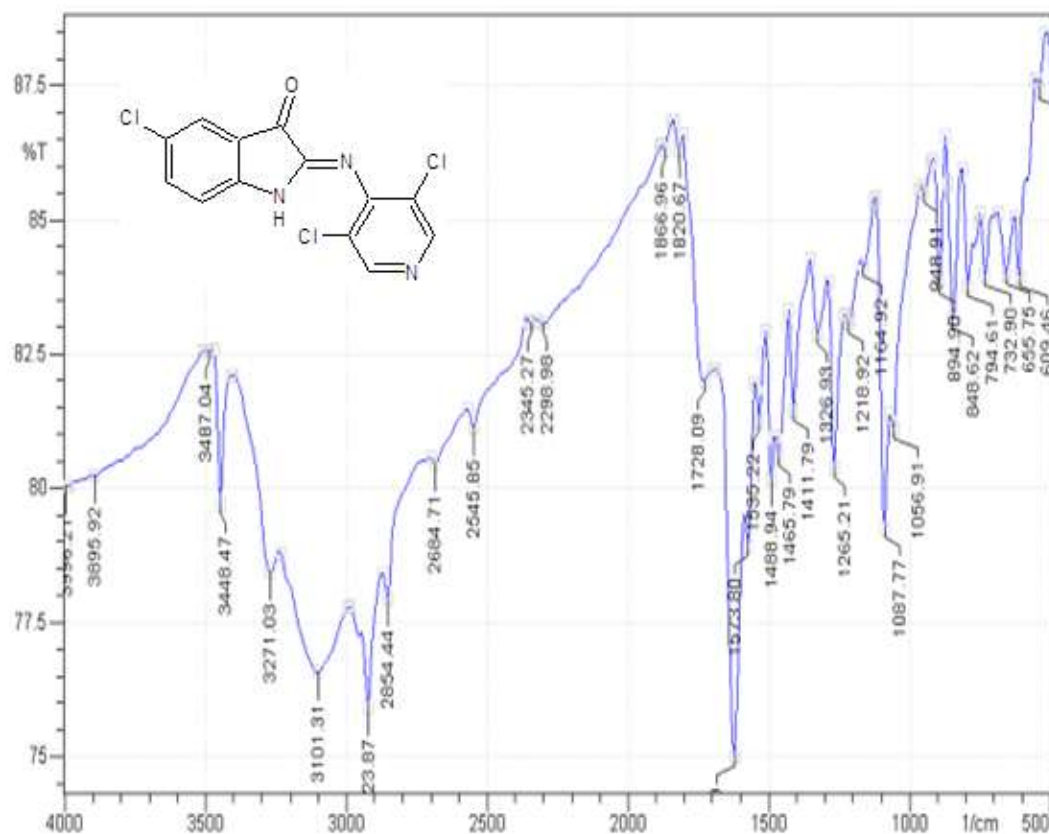
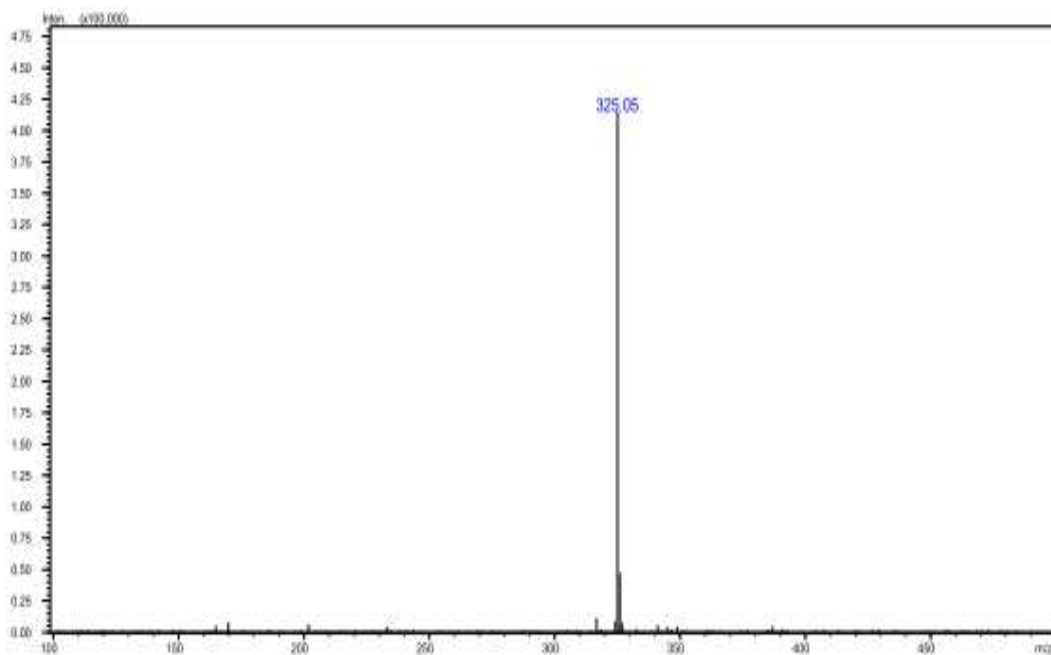
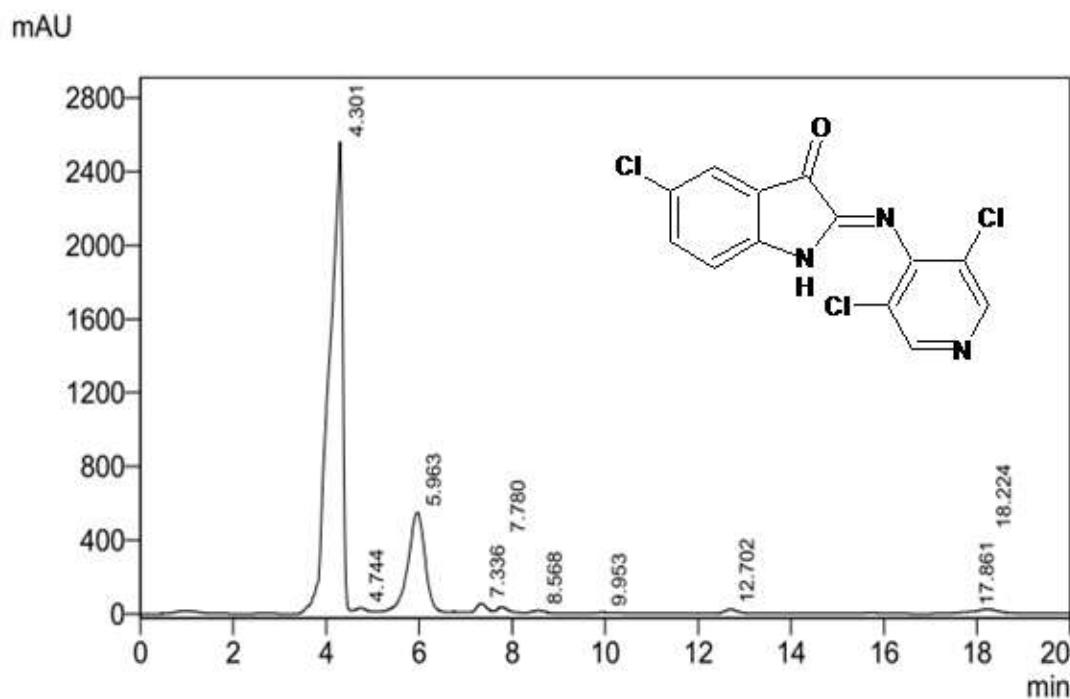


Table 8: IR interpretation of the compound PK2

| S.NO | WAVE NUMBER | TYPES OF VIBRATIONS | FUNCTIONAL GROUP |
|------|-------------|---------------------|------------------------|
| 1 | 2854.44 | C-H stretching | Alkyl group |
| 2 | 794.61 | C-Cl stretching | Presence of C-Cl group |
| 3 | 1728.09 | C=O Stretching | Presence of keto group |

LC-MS SPECTRUM OF THE COMPOUND PK2 (molecular weight-326.56)



NMR SPECTRUM OF THE COMPOUND PK2

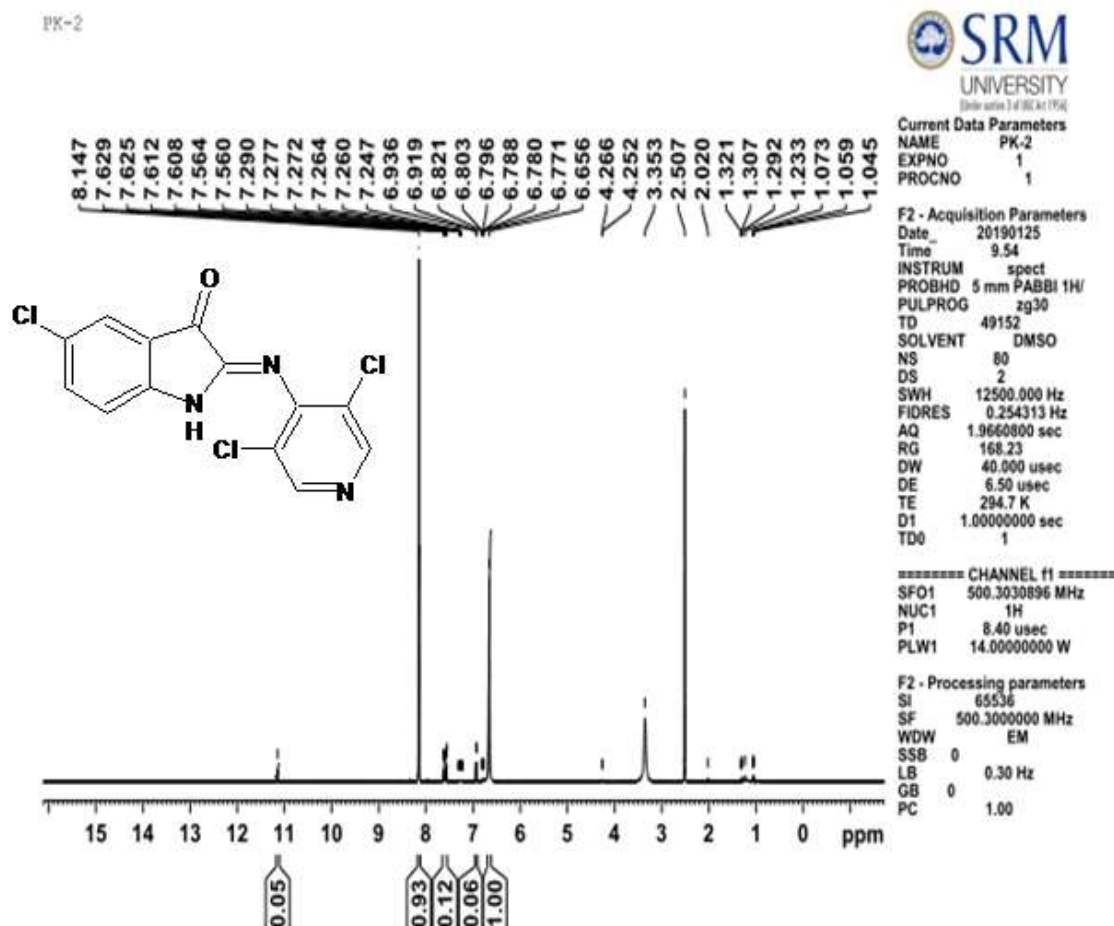
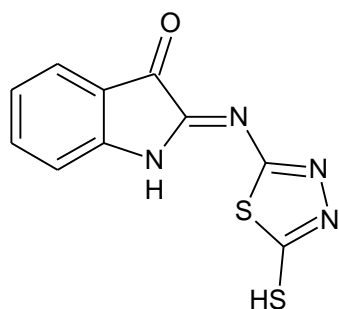


Table 9: NMR interpretation of the compound PK2

| S.NO | δ VALUES | TYPES OF PEAK | NUMBER OF PROTONS |
|------|-----------------|---------------|-------------------|
| 1 | 6.8-7.5 | Multiplet | Two protons |
| 2 | 3.3-4.2 | Triplet | Four protons |

PK3



| | |
|---------------------|---|
| Molecular Formula | = C ₁₀ H ₆ N ₄ OS ₂ |
| Formula Weight | = 262.31084 |
| Composition | = C(45.79%) H(2.31%) N(21.36%) O(6.10%) S(24.45%) |
| Molar Refractivity | = 69.18 ± 0.5 cm ³ |
| Molar Volume | = 147.4 ± 7.0 cm ³ |
| Parachor | = 439.0 ± 8.0 cm ³ |
| Index of Refraction | = 1.911 ± 0.05 |
| Surface Tension | = 78.7 ± 7.0 dyne/cm |
| Density | = 1.77 ± 0.1 g/cm ³ |
| Dielectric Constant | = Not available |
| Polarizability | = 27.42 ± 0.5 10 ⁻²⁴ cm ³ |

IR SPECTRUM OF COMPOUND PK3

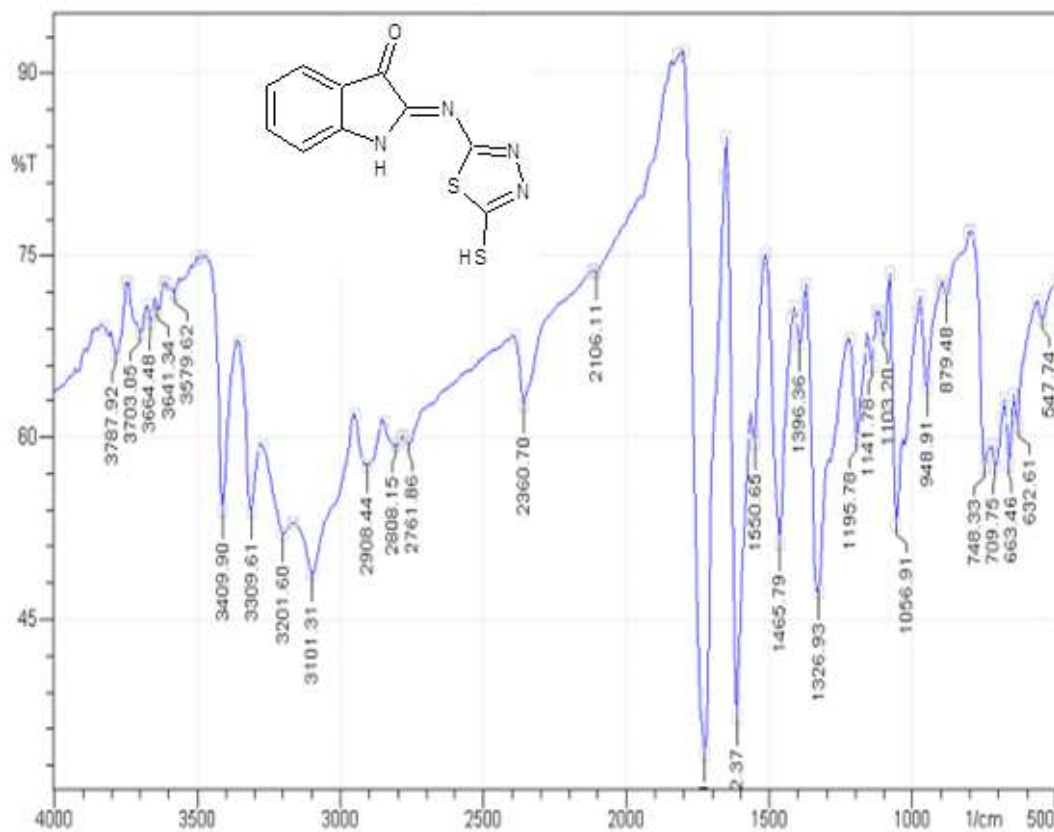
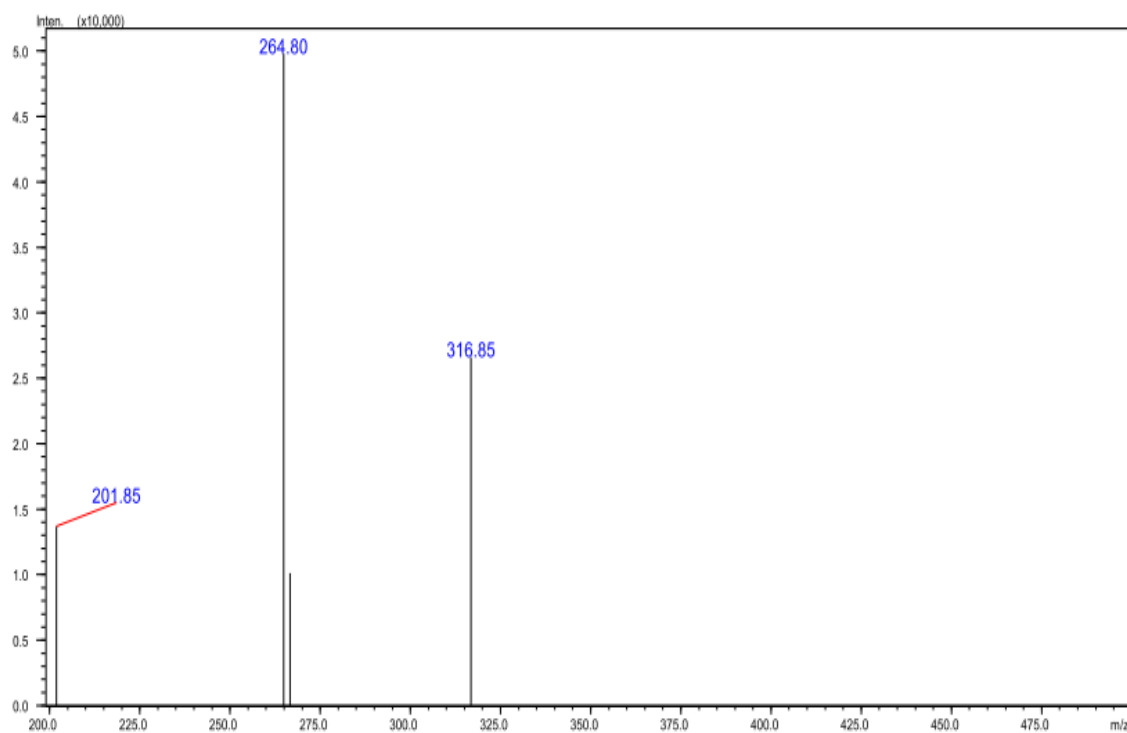
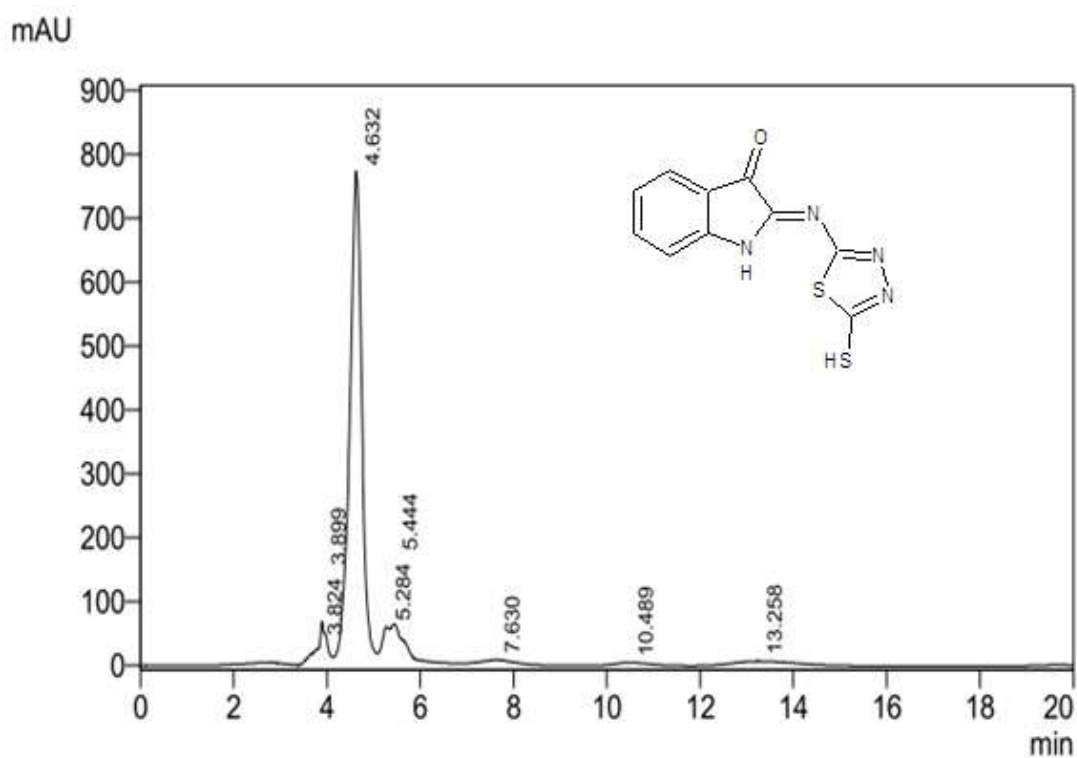


Table 10 : IR interpretation of the compound PK3

| S.NO | WAVE NUMBER | TYPES OF VIBRATIONS | FUNCTIONAL GROUP |
|------|-------------|---------------------|------------------|
| 1 | 2908.44 | C-H stretching | Alkyl group |
| 2 | 1612.37 | C=N stretching | Imine group |
| 3 | 748.33 | C-S stretching | Thiol group |

LC-MS SPECTRUM OF COMPOUND PK3 (molecular weight-262.31)



NMR SPECTRUM OF THE COMPOUND PK3

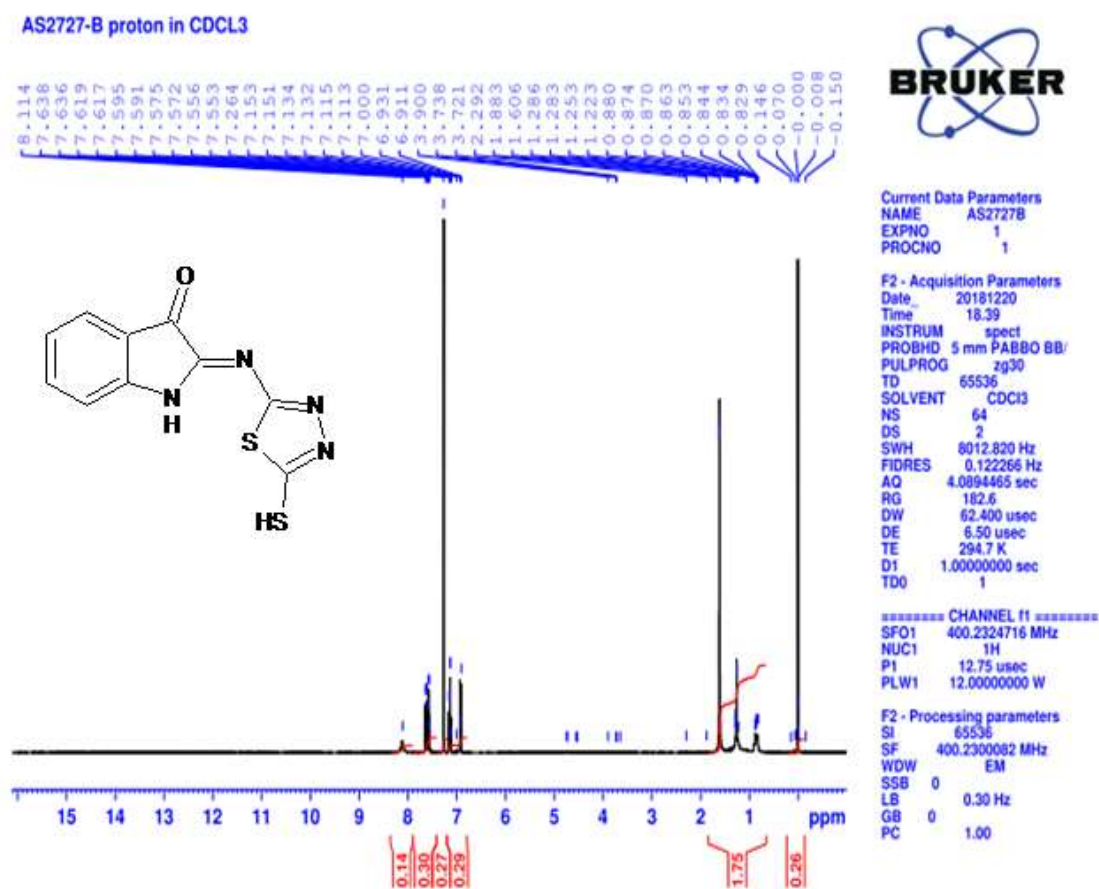
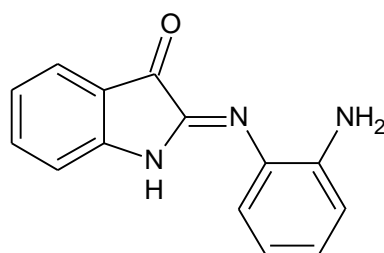


Table 11 : NMR interpretation of the compound PK3

| S.NO | δ VALUES | TYPES OF PEAK | NUMBER OF PROTONS |
|------|-----------------|---------------|-------------------|
| 1 | 6.9-7.5 | Doublet | Five protons |
| 2 | 7.6-8.1 | Multiplet | One proton |

PK4



(2Z)-2-[(2-aminophenyl)imino]-1,2-dihydro-3H-indol-3-one

| | |
|---------------------|--|
| Molecular Formula | = C ₁₄ H ₁₁ N ₃ O |
| Formula Weight | = 237.25664 |
| Composition | = C(70.87%) H(4.67%) N(17.71%) O(6.74%) |
| Molar Refractivity | = 68.29 ± 0.5 cm ³ |
| Molar Volume | = 175.9 ± 7.0 cm ³ |
| Parachor | = 482.8 ± 8.0 cm ³ |
| Index of Refraction | = 1.703 ± 0.05 |
| Surface Tension | = 56.7 ± 7.0 dyne/cm |
| Density | = 1.34 ± 0.1 g/cm ³ |
| Dielectric Constant | = Not available |
| Polarizability | = 27.07 ± 0.5 10 ⁻²⁴ cm ³ |

IR SPECTRUM OF COMPOUND PK4

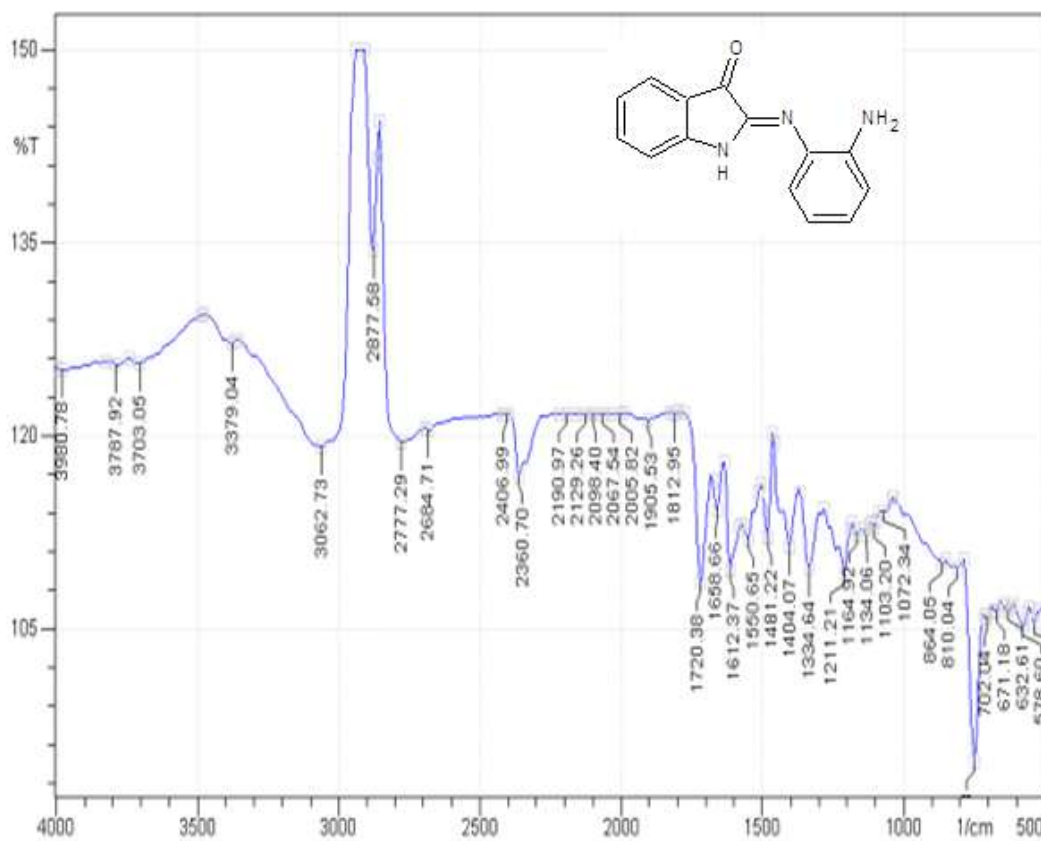
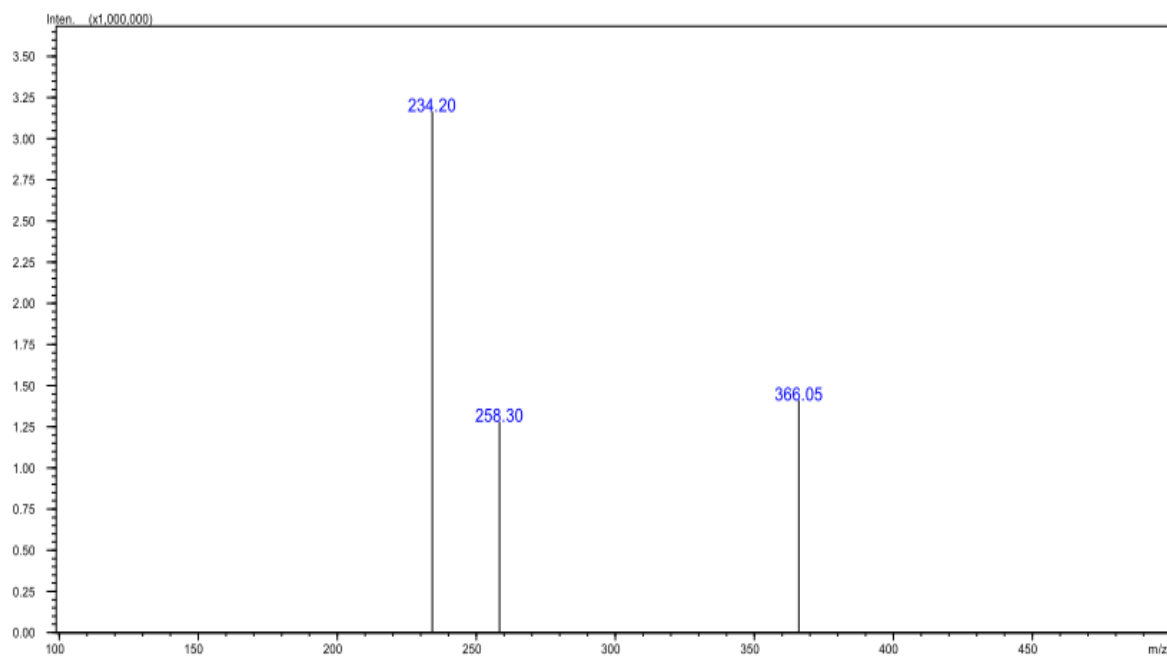
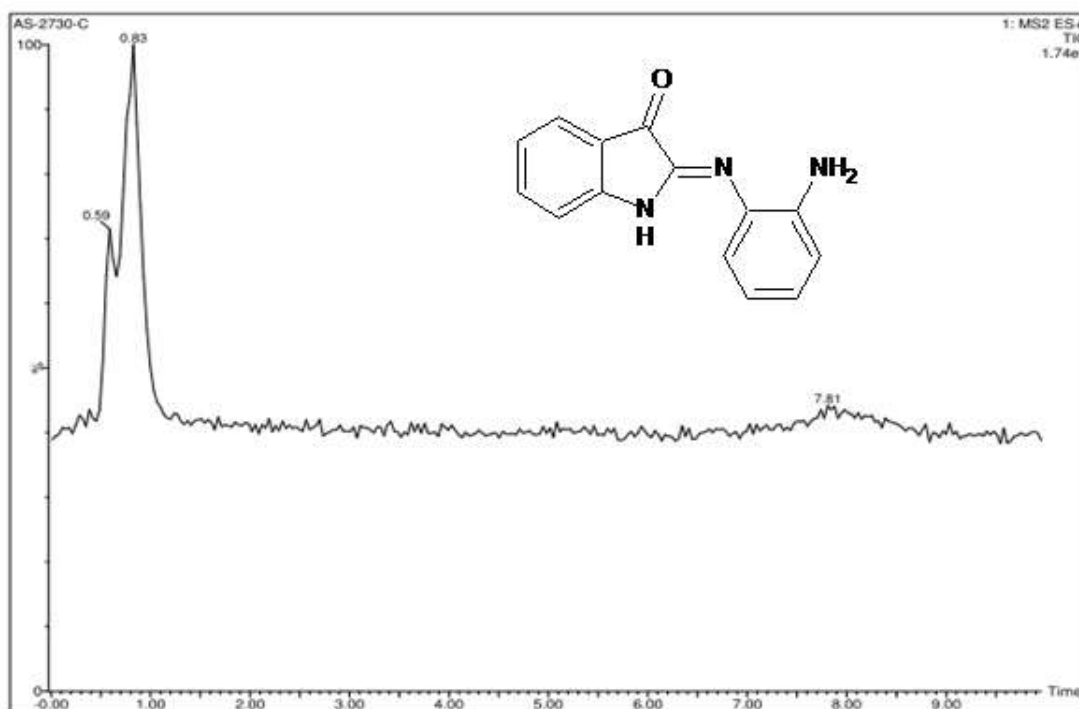


Table 12: IR interpretation of the compound PK4

| S.NO | WAVE NUMBER | TYPES OF VIBRATIONS | FUNCTIONAL GROUP |
|------|-------------|---------------------|------------------------|
| 1 | 3062.73 | C-H stretching | Alkene group |
| 2 | 1612.37 | C=N stretching | Imine group |
| 3 | 1720.38 | C=O Stretching | Presence of keto group |

LC-MS SPECTRUM OF COMPOUND PK4(molecular weight - 237.25)



NMR SPECTRUM OF COMPOUND PK4

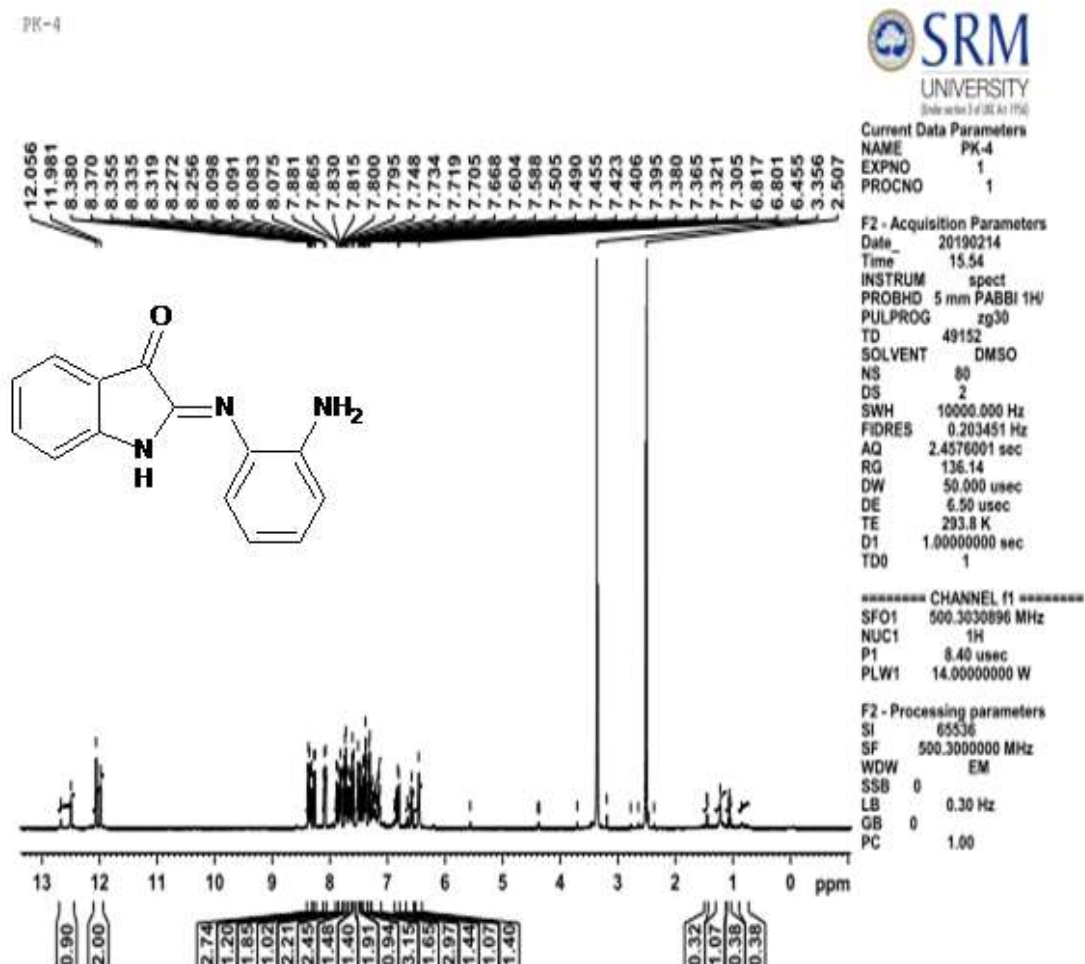
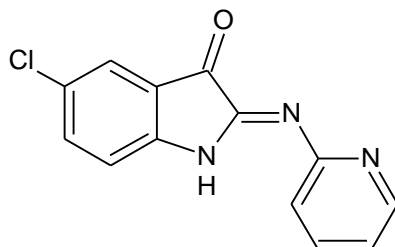


Table 13: NMR interpretation of the compound PK4

| S.NO | δ VALUES | TYPES OF PEAK | NUMBER OF PROTONS |
|------|-----------------|---------------|-------------------|
| 1 | 6.8-8.3 | Triplet | Nine protons |
| 2 | 2.5-3.3 | Multiplet | Two protons |

PK5



(2Z)-5-chloro-2-(pyridin-2-ylimino)-1,2-dihydro-3H-indol-3-one

| | |
|---------------------|---|
| Molecular Formula | = C ₁₃ H ₈ ClN ₃ O |
| Formula Weight | = 257.67512 |
| Composition | =C(60.60%)H(3.13%)Cl(13.76%)N(16.31%) O(6.21%) |
| Molar Refractivity | = 69.42 ± 0.5 cm ³ |
| Molar Volume | = 177.0 ± 7.0 cm ³ |
| Parachor | = 485.6 ± 8.0 cm ³ |
| Index of Refraction | = 1.712 ± 0.05 |
| Surface Tension | = 56.5 ± 7.0 dyne/cm |
| Density | = 1.45 ± 0.1 g/cm ³ |
| Dielectric Constant | = Not available |
| Polarizability | =27.52 ± 0.5 10 ⁻²⁴ cm ³ |

IR SPECTRUM OF COMPOUND PK5

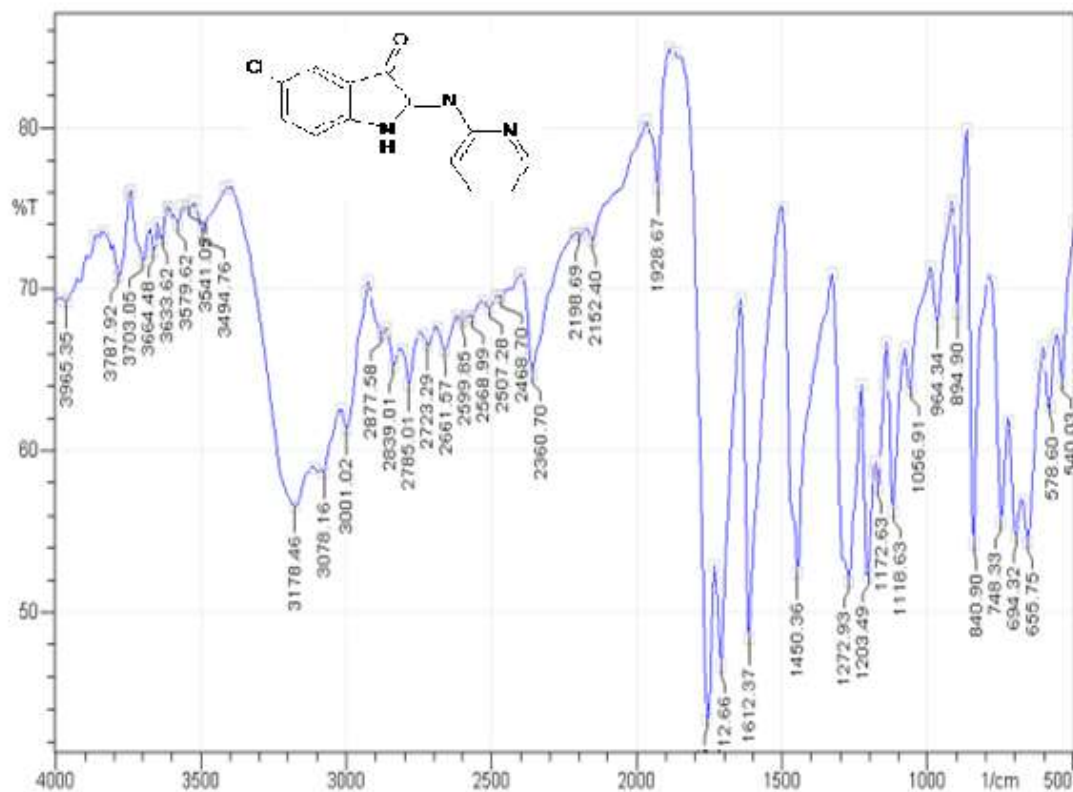
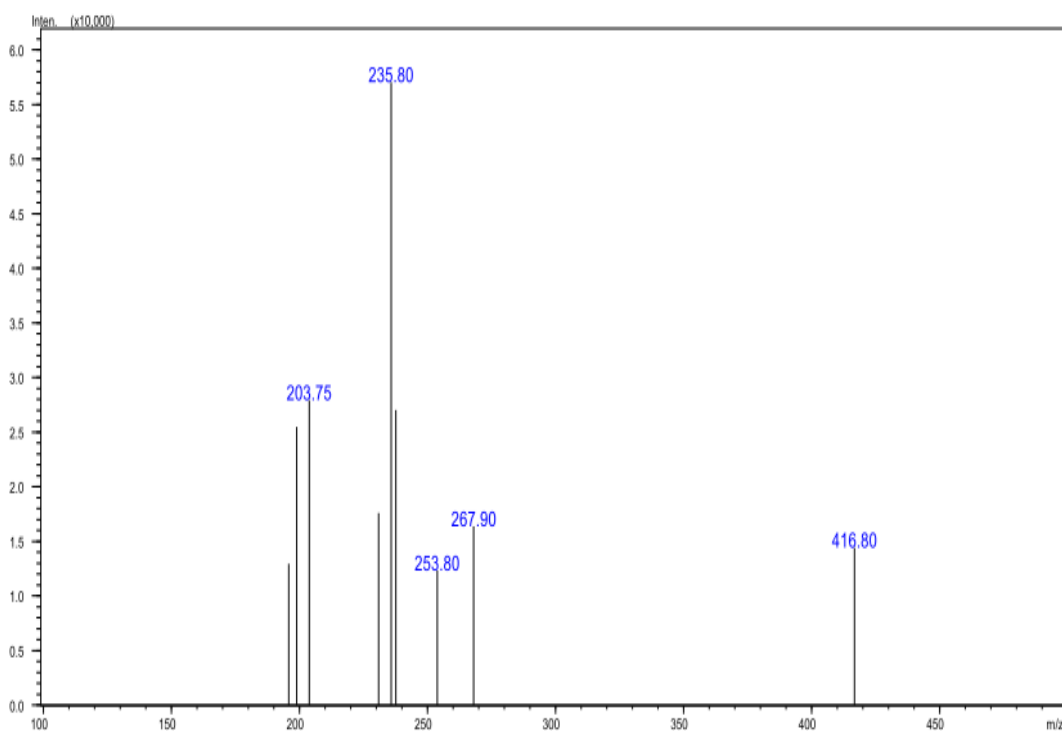
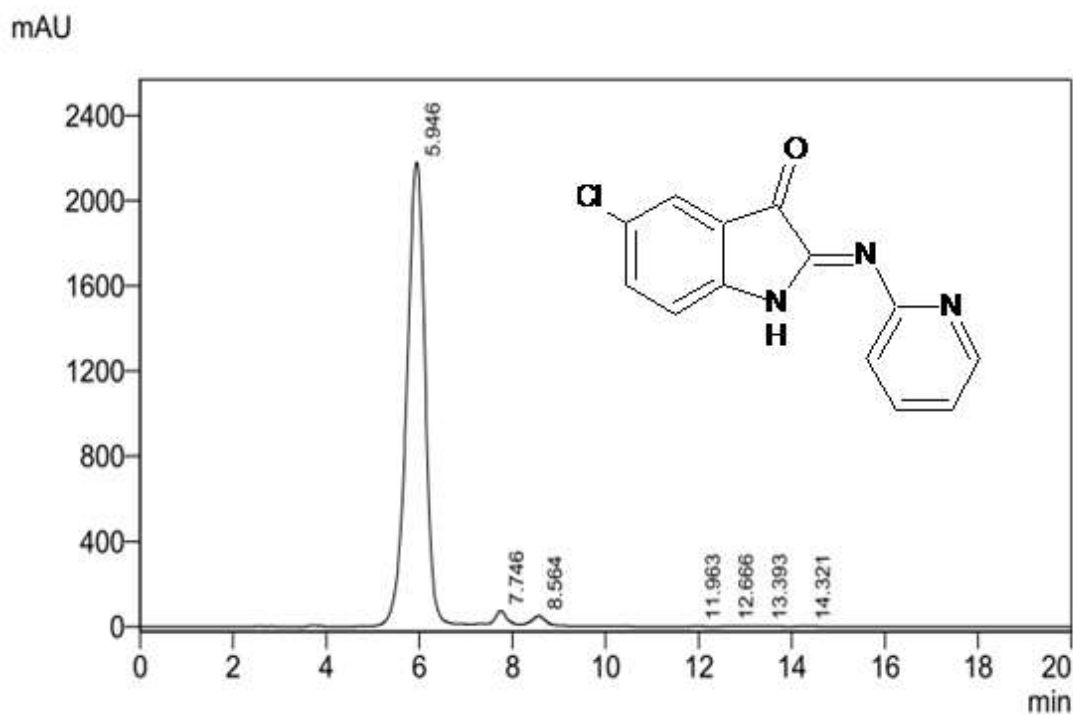


Table 14: IR interpretation of the compound PK5

| S.NO | WAVE NUMBER | TYPES OF VIBRATIONS | FUNCTIONAL GROUP |
|------|-------------|---------------------|------------------------|
| 1 | 2877.58 | C-H stretching | Alkyl group |
| 2 | 1612.37 | C=N stretching | Imine group |
| 3 | 748.39 | C-Cl stretching | Presence of C-Cl group |

LC-MS SPECTRUM OF COMPOUND PK5



NMR SPECTRUM OF COMPOUND PK5

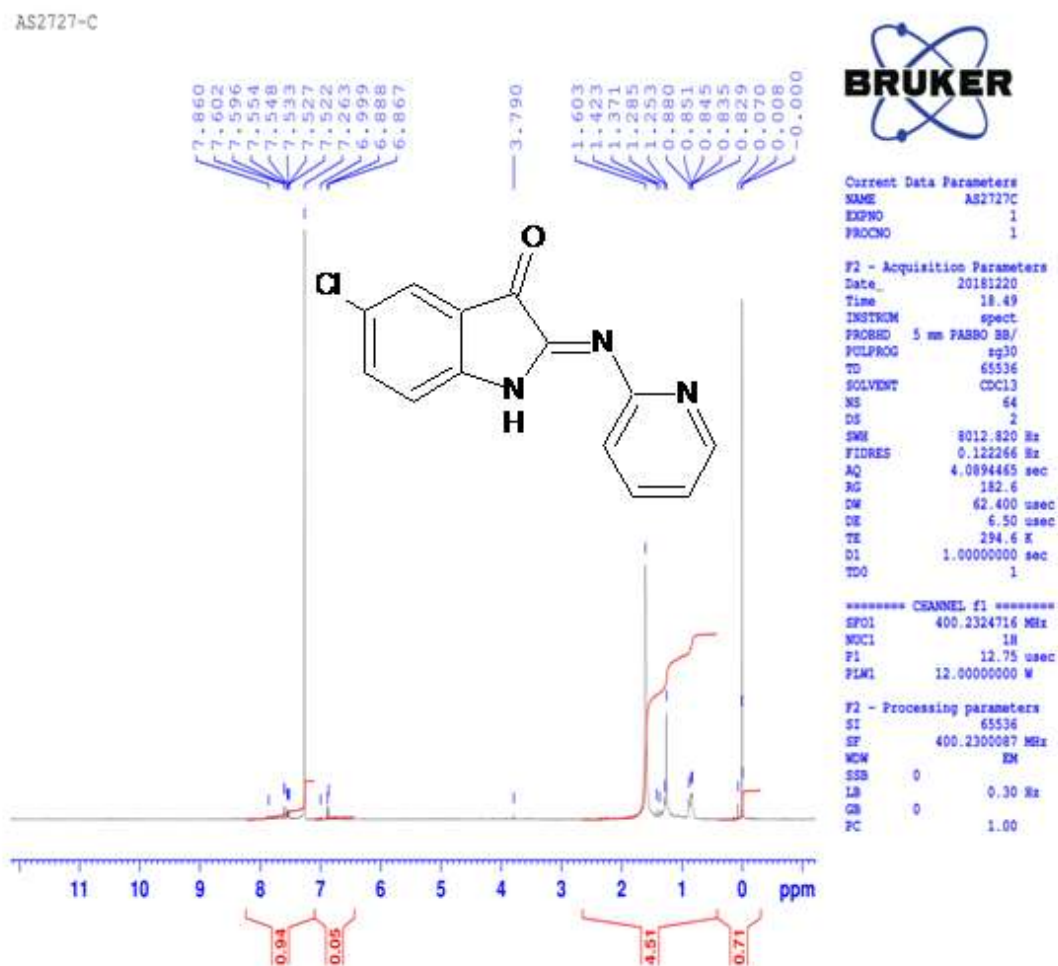


Table 15 : NMR interpretation of the compound PK5

| S.NO | δ VALUES | TYPES OF PEAK | NUMBER OF PROTONS |
|------|-----------------|---------------|-------------------|
| 1 | 6.8-7.8 | Triplet | Six protons |
| 2 | 3.7 | Multiplet | Two protons |

Table 16 : physical data of the synthesized compounds:

| S.NO | COMPOUND NAME | MOLECULAR FOUMLA | MOLECULAR WEIGHT | % YIELD | MELTING POINT |
|------|---------------|---|------------------|---------|---------------|
| 1 | PK1 | C ₁₄ H ₉ N ₅ O ₅ | 327.25 | 85 | 164-166°C |
| 2 | PK2 | C ₁₃ H ₆ Cl ₃ N ₃ O | 326.56 | 80 | 154-156°C |
| 3 | PK3 | C ₁₀ H ₆ N ₄ OS ₂ | 262.31 | 72 | 158-160°C |
| 4 | PK4 | C ₁₄ H ₁₁ N ₃ O | 237.25 | 80 | 180-182°C |
| 5 | PK5 | C ₁₃ H ₈ ClN ₃ O | 257.67 | 70 | 210-212°C |

IR SPECTROSCOPY:

- The absence of old functional group of the parent compound and the presence of new functional group of the synthesized compounds indicates the reaction is completed.
- All the synthesized compounds showed presence of the Imine formation. All these compounds show the absence of the stretching/bending for the reactant functional groups.

H¹ NMR SPECTROMETRY:

- The number of signals present in the NMR spectrum indicates the set of equivalent protons present in the molecule.
- Shielded protons shows upfield effect, and they are directed towards the signal of TMS. Shielded protons have lower δ values.
- Deshielded protons shows downfield effect, and they are directed away from the signal of TMS. Deshielded protons have higher δ values. Ex: OH, NO₂, F, Cl, Br containing protons have deshielding effect.

LC-MS:

Liquid Chromatography – Mass Spectrometry is used to determine the purity and mass of the synthesized compounds.

Table 17 : Molecular weight determined by mass spectrometry:

| S.NO | SAMPLE CODE | ACTUAL MASS | CALCULATED MASS |
|------|-------------|-------------|-----------------|
| 1 | PK1 | 327.25 | 325 |
| 2 | PK2 | 326.56 | 325.05 |
| 3 | PK3 | 262.31 | 264.80 |
| 4 | PK4 | 237.25 | 234.20 |
| 5 | PK5 | 257.67 | 253.80 |

BIOLOGICAL EVALUATION:

MICROPLATE ALAMAR BLUE ASSAY [MABA]:

1. The compounds were evaluated for their invitro antitubercular activity by the MABA method.
2. Strains used: Mycobacterium tuberculosis H37RV strains.
3. The standard drugs used are:
 - Pyrazinamide-3.125µg/ml.
 - Streptomycin-6.25µg/ml.
 - Ciprofloxacin-3.125µg/ml

The results for the standard drugs is presented below

Standard Drug Photograph

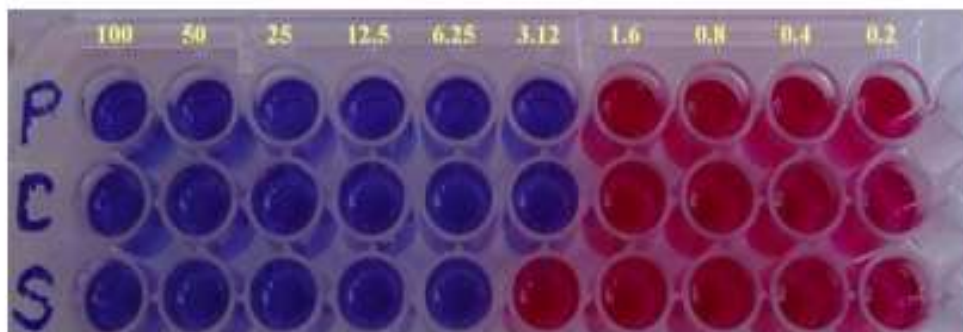


Fig 8. standard drug photograph (MABA)

| S. No. | Sample | 100 µg/ml | 50 µg/ml | 25 µg/ml | 12.5 µg/ml | 6.25 µg/ml | 3.12 µg/ml | 1.6 µg/ml | 0.8 µg/ml | |
|--------|--------|--------------|-------------|-------------|---------------|---------------|---------------|--------------|--------------|--|
| 1 | PK1 | | | | | | | | | |
| 2 | PK2 | | | | | | | | | |
| 3 | PK3 | | | | | | | | | |
| 4 | PK4 | | | | | | | | | |
| 5 | PK5 | | | | | | | | | |

Fig 9: MABA photograph for synthesized compounds

Table 18 : The MABA report of the synthesized compounds were tabulated below:

| S. No. | Sample | 100 $\mu\text{g/ml}$ | 50 $\mu\text{g/ml}$ | 25 $\mu\text{g/ml}$ | 12.5 $\mu\text{g/ml}$ | 6.25 $\mu\text{g/ml}$ | 3.12 $\mu\text{g/ml}$ | 1.6 $\mu\text{g/ml}$ | 0.8 $\mu\text{g/ml}$ |
|--------|--------|----------------------|---------------------|---------------------|-----------------------|-----------------------|-----------------------|----------------------|----------------------|
| 1 | PK1 | S | S | R | R | R | R | R | R |
| 2 | PK2 | S | S | S | S | S | S | S | R |
| 3 | PK3 | S | S | R | R | R | R | R | R |
| 4 | PK4 | S | S | S | S | S | S | S | R |
| ss5 | PK5 | S | S | R | R | R | R | R | R |

Table 19 : Comparative study of docking score with MABA report:

| Compound name | Docking score Kcal/mol | MABA (biological activity) ($\mu\text{g/ml}$) |
|---------------|---------------------------|---|
| PK1 | -7.56 | 50 |
| PK2 | -7.1 | 1.6 |
| PK3 | -6.57 | 50 |
| PK4 | -5.83 | 1.6 |
| PK5 | -6.39 | 50 |
| PYRAZINAMIDE | -4.69 | 3.125 |
| CIPROFLOXACIN | -6.45 | 3.125 |

*SUMMARY
AND
CONCLUSION*

SUMMARY

- Glutamine synthetase I enzyme is the critical enzyme for the survival and growth for *Mycobacterium tuberculosis* is chosen as the potential drug target.
- Molecules were designed and docked against Glutamine synthetase I enzyme 3ZXR protein using Autodock® 4.2.1 software.
- Molecules with good docking score were screened for insilico toxicity by using OSIRIS® software and evaluation of drug likeness by MOLINSPIRATION software.
- The selected compounds were synthesized and labeled as PK1, PK2, PK3, PK4, PK5.
- Purity of the synthesized compounds were justified by its sharp melting point and TLC.
- Further the synthesized compounds were characterized by IR, LC-MS, ¹H NMR.
- Biological evaluation is done by MICROPLATE ALAMAR BLUE ASSAY
- Compounds PK2 and PK4 showed activity at 1.6 µg/ml concentrations which are comparable to the activity of the standard drugs Pyrazinamide, Streptomycin, Ciprofloxacin which are active at 3.125µg/ml, 6.25µg/ml, 3.125µg/ml concentrations.

CONCLUSION

- A series of Schiff bases of isatin derivatives were designed, docked, synthesized and evaluated against Glutamine synthetase 1 enzyme which is critical for the survival and growth of MTB.
- The Minimum Inhibitory Concentration of the synthesized compounds ranges from 50-1.6 μ g/ml.
- The work concludes that the Compounds Compounds PK2 and PK4 showed activity at 1.6 μ g/ml concentrations which are comparable to the activity of the standard drugs Pyrazinamide, Streptomycin, Ciprofloxacin which are active at 3.125 μ g/ml,6.25 μ g/ml,3.125 μ g/ml concentrations.
- Further, structural refinement of the synthesized compounds is expected to yield promising molecules against the pathogen MTB.

REFERENCES

REFERENCES

1. <http://en.wikipedia.org/wiki/Tuberculosis>.
2. Dye C, Williams B. G. The population dynamics and control of tuberculosis. *Science* 2010;328:856-861.
3. Issar Smith. Mycobacterium tuberculosis Pathogenesis and Molecular Determinants of Virulence. *Clin Microbiol Rev.* 2003; 16: 463-496.
4. <https://en.m.wikipedia.org>
5. World health organization. Definitions and reporting framework for tuberculosis.2013
6. <https://www.healthxchange.sg>heart-lungs>.
7. J Global tuberculosis report 2012. Geneva: World Health Organization (http://www.who.int/tb/publications/global_report/en/).
8. Zignol M, van Gemert W, Falzon D. Surveillance of anti-tuberculosis drug resistance in the world: an updated analysis 2007-2010. *Bull World Health Organ.* 2012;90:111D-119D.
9. Tuberculosis MDR-TB and XDR-TB: 2011 progress report. Geneva: World Health Organization, 2011 (http://www.who.int/tb/challenges/mdr/factsheet_mdr_progress_march2011.pdf).
10. Mercy Eleanor G , Aditya T , Amarendra Kumar E, Meria M Dan. Review on Mycobacterium Tuberculosis. *Research and Reviews: Journal of Microbiology and Biotechnology*: aug 2016
11. Barry C E, Crick D C, Mc Neil M R. Targeting the formation of the cell wall core of mycobacterium tuberculosis. *Infect. Disord. Drug targets* 2007 (7);182-202
12. Jacson M, McNeil M R, Brennan P J. Progress in targeting cell envelope biogenesis in Mycobacterium tuberculosis. *Fut. Microbiol.* 2013(8); 855-875
13. <http://www.Genome structure of M.Tuberculosis.org>.
14. Hertzberg RP, Pope AJ. High-throughput screening: New technology for the 21st century. *Curropinchem boil* 2000;4:445-451.
15. Cole S T, Brosch R, Parkhill J , Garnier T, Churcher C, Harris D. Deciphering the biology of Mycobacterium tuberculosis from the complete genome sequence. *Nature.* June1998 (6685):537-44.

16. Mycobacterium tuberculosis, sanger institute. 2007-03-29. Retrived 2008-11-16
17. Pratik Swarup Das and Puja Saha. A review on computer aided drug design in drug discovery. World journal of pharmacy and pharmaceutical sciences. 2017;6(7):279-291.
18. Hajduk PJ, Huth JR, and Tse C. Predicting protein druggability. Drug Discov Today. 2005; 10: 1675-1682.
19. Fauman EB, Rai BK, and Huang ES. Structure-based druggability assessment-identifying suitable targets for small molecule therapeutics. Curr. Opin. Chem. Biol.2011; 15:463-468.
20. Becker OM, Dhanoa DS, Marantz Y, Chen D, Shacham S, Cheruku S, Heifetz A, Mohanty P, Fichman M, Sharadendu A. An integrated in silico 3D model-driven discovery of a novel, potent, and selective amidosulfonamide 5-HT1A agonist (PRX-00023) for the treatment of anxiety and depression. J. Med. Chem.2006; 49: 3116-3135.
21. Warner SL, Bashyam S, Vankayalapati H, Bearss DJ, Han H, Mahadevan D, Von Hoff DD, Hurley LH. Identification of a lead small-molecule inhibitor of the Aurora kinases using a structure-assisted, fragment-based approach. Mol. Cancer.Ther.2006;5:1764-1773.
22. Mitchell JBO, Laskowski RA, Alex A, Forster MJ, Thornton JM. BLEEP-Potential of mean force describing protein-ligand interactions: II. Calculation of binding energies and comparison with experimental data. J. Comput. Chem.1999; 20: 1177-1185.
23. Robert A, Ronzio W, Bruce Rowe and Alton Meister. Studies on the Mechanism of Inhibition of Glutamine Synthetase by Methionine Sulfoximine. Biochemistry. 1969;8(3):1066-1075.
24. Einsberg D, Almasy R J, janson C A, Chapman M S, Suh S W, Cascio D, Smith W W. Some evolutionary relationships of the primary biological catalysts glutamine synthetase and Rubisco. Cold spring Harb. Symp. Quant. Boil. 1987;52:483-90.
25. <http://www.rcsb.org>

26. Berlicki L, Kafarski. Inhibitors of glutamine synthetase and their potential application in medicine. *Mini reviews in medicinal chemistry*. 2008 Aug; 8(9):869-878.
27. Mats Larhed, Olof Lagerlund, Luke R Odell, Daniel Muthas Anneli Nordqvist. Functionalized 3-amino-imidazo[1,2-a]pyridines: A novel class of drug-like Mycobacterium tuberculosis glutamine synthetase inhibitors. *Bioorganic & Medicinal Chemistry Letters*. 2009; 19(16):4790-4793.
28. Hardik Patel, Nilesh Darji, Jagath Pillai and Bhagirath Patel. Recent advance in anticancer activity of indole derivatives. *International journal of drug research and technology*. 2012; 2(3):225-230
29. Amit k. singh, raj k. Prasad, Chandra s.singh. Synthesis, characterization and pharmacological evaluation of some novel 3-indole derivatives. *Der pharma chemica*. 2013; 5(2):311-319
30. Hardik Patel, Nilesh Darji, Jagath Pillai and Bhagirath Patel. Recent advance in anti-cancer activity of indole derivatives. *Int. J.drug Res. Tech.* 2012; 2(3):225-230
31. Alimuddin Zumla, Mario Raviglione, Richard Hafner, and C. Fordham von Reyn. current concepts Tuberculosis. *N Engl J Med* 2013; 368:745-55.
32. Dye, C. Williams, B.G, the Population Dynamics and Control of Tuberculosis. *Science* 2010; 328:856-861.
33. "Robert Koch and Tuberculosis: Koch's Famous Lecture" . Nobel Foundation. 2008.
34. Pierpalo de colombai. The Global Plan to Stop TB. 2006–2015 [web site]. Geneva, Stop TB Partnership, (2007); www.stoptb.org/globalplan/, accessed 6.
35. Balabanova Y, Drobniewski F, Fedorin I, Zakharova S, Nikolayevskyy V, Atun R, Coker R. "The Directly Observed Therapy Short-Course (DOTS) strategy in Samara Oblast, Russian Federation." *Respir Res*. Mar (2006); 23:7:44.
36. Ruohonen RP, Goloubeva TM, Trnka L, Fomin MM, Zhemkova GA, Sinitsyn AV, Lichachev AA, Koskela KG. "Implementation of the DOTS strategy for

- tuberculosis in the Leningrad Region, Russian Federation. *Int J Tuberc Lung Dis.* Mar 2002; 6(3): 192-7.
37. Keane, J., Balcewicz Sablinska, M. K., Remold, H. G., Chupp, G. L., Meek, B. B., Fenton, M.J., & Kornfeld, H. Infection by *Mycobacterium tuberculosis* promotes human alveolar macrophage apoptosis. *Infection and immunity* 1997; 65(1), 298-304.
38. Thomas R. Ioerger, Yicheng Feng, Krishna Ganesula, Xiaohua Chen, Karen M. Dobos, Sarah Fortune, William R. Jacobs, Valerie Mizrahi, Tanya Parish, Eric Rubin, Chris Sasseti and James C. Sacchetti. Variation among Genome Sequences of *M. Tuberculosis* from Multiple Laboratories. *Journal of bacteriology.* July 2010; 192(14): 3645-3653.
39. Huajun Zheng, Liangdong Lu, Bofei Wang, Shiyong Pu, Xianglin Zhang, Genfeng Zhu, Wanliang Shi, Lu Zhang, Honghai Wang, Shengyue Wang, Guoping Zhao, Ying Zhang. Genetic basis of Virulence Attenuation Revealed by Comparative Genomic Analysis of *M. Tuberculosis* Strain H37Ra versus H37Rv. *PLoS ONE*. Feb 2008; 3(6): e2375
40. Krajewski WW Jones TA and Mowbray SL. Structure of *Mycobacterium tuberculosis* glutamine synthetase in complex with a transition-state mimic provides functional insights. *Proceedings of the National Academy of Sciences USA.* 2005; 102: 10499-10504.
41. Andreas Burkovski. Ammonium assimilation and nitrogen control in *Corynebacterium glutamicum* and its relatives: an example for new regulatory in actinomycetes. *FEMS Microbiology Reviews* 27. 2003: 617-628.
42. David Eisenberg, Harindarpal S. Gill, Gaston M.U. Puegl, Sergio H. Rotstein. Structure function relationships of glutamine synthetases. *Biochimica et Biophysica Acta.* 2000; 1477: 122- 145.
43. C.A. Woolfolk, E.R. Stadtman, *Arch. Biochem. Biophys.* 1967; 118: 736-755.
44. Pratik Swarup Das and Puja Saha. a review on computer aided drug design in drug discovery. *World journal of pharmacy and pharmaceutical sciences.* 2017; 6(7): 279-291.
45. Wermuth CG. Similarity in drugs: reflections on analogue design. *Drug Discov Today.* 2006Apr; 11(7-8): 348-54.

46. Laurie AT, Jackson RM. Methods for the prediction of protein-ligand binding sites for structure-based drug design and virtual ligand screening. *Curr. Protein. Pept. Sci.*2006; 7: 395-406.
47. Lipinski CA (December 2004). "Lead- And Drug-Like Compounds: The Rule-Of-Five Revolution". *Drug Discovery Today: Technologies.* 2004;1(4): 337– 341.Doi:10.1016/J.Ddtec.2004.11.00
48. Amit k. singh, raj k. Prasad, Chandra s.singh. Synthesis, characterization and pharmacological evaluation of some novel 3-indole derivatives. *Der pharma chemica.*2013;5(2):311-319
49. Karali N, Gursoy A, Kandemirli F, Shvets N, Kaynak F.B, Ozbey S, Kovalishyn, V, Dimoglo A. Synthesis and structure- antituberculosis activity relationship of 1H-indole-2,3-dione derivatives. *Bioorg Med Chem.* June 2007;15(17):5888-5904.
50. Li Y Y, Wu H. S, Tang L, Feng C. R., Yu J. H, Li Y, Yang Y. S, Yang B, He Q. The potential insulin sensitizing and glucose lowering effects of a novel indole derivative in vitro and in vivo. *J Pharmacol Res* 2007;56:335-43
51. Hiari Y.M.A, Qaisi A. M, Abadelah M.M, Voelter W. *Monatshefte Fur Chemie* 2006;137:243.
52. Hong B. C, Jiang Y, Chang Y, Lee S. *J Chin ChemSoc* 2006, 53, 647.
53. Enein H.Y.A, Kruk I, Lichszeld K, Michalska T, Kiadna A. Marczynski S, Olgen, S. *Luminescence* 2004;19(1)
54. Abele E, Abele R, Dzenitis O, Lukevics E. indole and isatin oximes synthesis, reactions and biological activity. *Chem Heterocycl Compd* 2003;39(3)
55. Radwan, M. A. A, Ragab E.A, Sabry N.M, Shenawy S.M. E. *Bioorg Med Chem* 1997;15:3832.
56. Merino I, Monge A, Font M, Irujo J. J. M, Alberdi E, Santiago E, Prieto I, Lasarte J. J, Sarobe P, Borra´s F. *Il Farmaco* 1999;54:255.
57. Jia-Lun Deng, Xiao-Cheng Liu, Guang-Wei Cai, Gang Zhang, Li Hu, Li Qiu, Zi-Yong Li, and Zhi Xu, Heteronuclear 5-Fluoroisatin Dimers: Design, Synthesis, and Evaluation of Their In Vitro Anti-mycobacterial Activities. *J. Heterocyclic Chem.*2018;00:00
58. Parvanesh Pakravan, Soheila Kashanian, Mohammad M .khodaei.2013; [http://dx.doi.org/10.1016/S1734-1140\(13\)71007-7](http://dx.doi.org/10.1016/S1734-1140(13)71007-7).

59. manjusha verma ,surendra nath pandeya ,krishna nand singh, james p. stables
Anticonvulsant activity of Schiff bases of isatin derivatives. *Acta Pharm.*
2004; 54:49–56
60. Garden S.J, Torres J.C, Silva L.E, Pinto A.C, *Synth.Commun.*1998;28:1679.
61. Pinto A.C, Silva F. S. Q, Silva R. B, *Tetrahedron Lett.*1994;35,8923.
62. J. K. Kochi. "The Mechanism of the Sandmeyer and Meerwein Reactions". *J. Am. Chem. Soc.*(1957);79 (11): 2942–2948.
63. Erdmann, O. L.; *J. Prakt.Chem.*1840.
64. Anitha G.Amin, Shiva K Angala, Delphi Chatterjee, and Dean C.Crick. Rapid screening of inhibitors of *Mycobacterium tuberculosis* Growth using Tetrazolium salts. *Methods Mol Biol.*2009;465:187-201.
65. Neetu kumra Taneja and Jaya Siwaswami Tyagi. Resazurin reduction assays for screening of anti-tubercular compounds against dormant and actively growing *Mycobacterium tuberculosis*, *Mycobacterium bovis* BCG and *Mycobacterium smegmatis*. *Journal of Antimicrobial Chemotherapy.*2007 Aug;60(2):288-293.
66. Juan-Carlos Palomino, Anandi Martin, Mirtha Camacho, Humberto Guerra, Jean Swings, and Françoise Portaels. Resazurin Microtiter Assay Plate: Simple and Inexpensive Method for Detection Drug Resistance in *Mycobacterium tuberculosis* Antimicrob Agents Chemother. 2002 Aug; 46(8): 2720–2722.
67. Hong.X, Hopfinger.AJ, Construction, molecular modeling, and simulation of mycobacterium Tuberculosis cell walls. *Biomacromolecules* (2004);5:1052–1065.
68. Laurie AT et al. Q-site finder, an energy based method for the prediction of protein-ligand binding sites. *Bioinformatics.* 2005;21(9):1908-1916.
69. Drug design availability from
URL:<http://En.Wikipedia.Org/Wiki/DrugDesign>. Accessed on 12.2.2013.
70. Goodsell, DS .Glutamine synthetase. RCSB Protein Data Bank. Retrieved May 2010.
71. Johan Gising, Mikael T Nilsson et al. Trisubstituted Imidazoles as *Mycobacterium tuberculosis* Glutamine Synthetase Inhibitors. *Journal of Medicinal Chemistry.* 2012;55(6):2894-2898.

72. Amari. S, aizawa.H, Zhang J, Fukuzawa K, Mochizuki.Y, IiwasawaY, Nakata K, ChumanH, Nakano T. "viscana: visualized cluster analysis of protein-ligand interaction based on the ab initio fragment molecular orbital method for virtual ligand screening". J cheminfmodel2006;46(1): 221–230.Doi:10.1021/ci050262q.
73. Madsen, Ulf; Krogsgaard Larsen, Povl; Liljefors, Tommy Text book of Drug Design and Discovery. Washington, DC: Taylor & Francis. (2002); ISBN0415282888.
74. Morris GM, Huey R, Lindstrom W, et al. AutoDock4 and AutoDockTools4: automated docking with selective receptor flexibility. Journal of Computational Chemistry. 2009;30(16):2785-91.
75. <http://www.molinspiration.com/cgi-bin/properties>.Retrieved on7.3.2014.
76. Robert.M.Silverstein, Francis X Webster and David J Kiemle.Spectrometric identification of organic compounds. Seventh edition. John Wiley &Sons,Inc. 2005;19-40:82-108.
77. KalsiP.S. spectroscopy of organic compounds. 6th edition. New Delhi: new age international private ltd.2002:185-186
78. SharmaY.R. elementary organic spectroscopy.5th edition. . New Delhi. S.chand and company pvt ltd.2013:75-159
79. Nakayama GR, Caton MC, Nova MP and Parandoosh Z. Assessment of the Alamar Blue assay for cellular growth and viability in vitro. Journal of Immunological Methods. 1997;204(2):205-208.
80. Ramprasad S N. Multiple applications of Alamar Blue as an indicator of metabolic function and cellular health in cell viability bioassays. Sensors. 2012; 12(12):12347-12360.
81. Miku J, Steverding D. A simple colorimetric method to screen drug cytotoxicity against Leishmania using the dye Alamar Blue. Parasitology International. 2000;48(3):265-269.

ANNEXURE



INDIAN PHARMACEUTICAL ASSOCIATION

NATIONAL CONVENTION 2017 - 18

Theme: Pharma Vision 2030: Planning the Future

Certificate of Participation

This is to certify that

Dr./Prof./Mr./Ms.....K.ALAJARASI.....P.....

participated in the National Convention 2017 - 18 held at B. S. Abdur Rahman

Crescent Institute of Science & Technology, Chennai, 10th - 11th February 2018


Dr. S. Manivannan
Chairman - LOC


J. Jayaseelan
IPA Convention Co-ordinator - LOC


T. Sathish
Hon. Secretary - LOC

*"Skill and Will
to Make and Serve
Quality Pills"*
69th IPC
22nd - 24th December, 2017
CHANDIGARH



Certificate

This is to certify that

Prof./Dr./Mr./Ms.*K.A.L.A.Y.A.R.A.S.I.: P*.....

has participated as Delegate / Volunteer

in the 69th Indian Pharmaceutical Congress

held at Chitkara University, Rajpura from December 22nd to 24th, 2017.

Dr. Mahesh Burande
President - IPCA

Dr. Shailendra Saraf
Chairman - LOC

Dr. Dhirender Kaushik
Organizing Secretary

Dr. Ashish Baldi
Chairman, Registration Committee - LOC

Organised by : Indian Pharmaceutical Congress Association (IPCA)

Hosted by :

Association of Pharmaceutical Teachers of India (APTI)



Workshop on

**INDUSTRIAL PERSPECTIVES IN ANALYTICAL TECHNIQUES
& TRAINING ON CHEMOMETRICS SOFTWARE**

Conducted by

**Department of Pharmaceutical Analysis
C.L. BAID METHA COLLEGE OF PHARMACY**

Thoraipakkam, Chennai - 600 097.

Co - Sponsored by

The Tamil Nadu Dr. M.G.R. Medical University, Chennai

15th & 16th September 2017

CERTIFICATE



This is to certify that Dr/Mr/Ms. KALAYARASI.P

has participated as Resource Person /Chair Person / Delegate in the Workshop

on "Industrial Perspectives in Analytical Techniques and Training

on Chemometrics Software" on 15th and 16th September 2017.

C.N.N.

Dr. C.N. Nalini
Organizing Secretary

Dr. Grace Rathnam
Dr. Grace Rathnam
Convenor





ARULMIGU KALASALINGAM COLLEGE OF PHARMACY

in association with

THE TAMIL NADU DR.M.G.R. MEDICAL UNIVERSITY



Certificate

This is to certify that Mr./Ms./Prof./Dr. P. KALAYARASI of _____

has actively participated in the International

Seminar on "3D - Approaches in Combinatorial Chemistry : Challenges and Perspectives" organized by Department of Pharmaceutical Chemistry on 28.02.2018
at Arulmigu Kalasalingam College of Pharmacy, Krishnankoil, Tamil Nadu.

**5 credit points awarded by The Tamil Nadu Dr.M.G.R. Medical University,
Chennai.**

Dr.R.Rajapandi
Scientific Advisor

Dr.J.Amutha Iswarya Devi
Organizing Secretary

Dr.N.Venkateshan
Convenor

Dr.S.Shasi Anand
Secretary

Dr.K.Sridharan
Chairman



DEPARTMENT OF PHARMACOGNOSY
SRI RAMACHANDRA COLLEGE OF PHARMACY
SRI RAMACHANDRA UNIVERSITY

(Declared under section 3 of the UGC Act, 1956)
Accredited by NAAC with 'A' Grade
Porur, Chennai - 600 116.

in association with
Society for Ethnopharmacology, Chennai Chapter

CERTIFICATE

*This is to certify that Dr./Mr./Ms. P. Kalayarasu.....
has participated as a Resource person / Delegate in the Pre conference workshop on "Recent
Advances in Analytical Techniques - Drugs & Pharmaceuticals" held on 27th June 2017,
organized by Sri Ramachandra University, Porur, Chennai.*

This carries 2 Credits.

A. Chand

DR. D. CHAMUNDEESWARI

Principal &
Co-ordinator, SFE - Chennai Chapter

K. V. Somasundaram

DR. K.V. SOMASUNDARAM

Dean of Faculties



ARULMIGU KALASALINGAM COLLEGE OF PHARMACY

The Seminar is sponsored & accredited by

THE TAMIL NADU DR.M.G.R. MEDICAL UNIVERSITY



Certificate

This is to certify that Mr./Ms./Prof./Dr. K.ALAJYARASI. P of

_____ has actively participated as a Delegate in the National

Seminar on **"Impact of Intellectual Property Rights in Formulation and Development"** organized by **Department of Pharmaceutics** on 23.02.2019 at **Arulmigu Kalasalingam College of Pharmacy, Krishnankoil, TamilNadu.**

5 credit points awarded by The TamilNadu Dr.M.G.R. Medical University, Chennai.



Dr.R.Ramprasad
Organizing Secretary

 S.R. Senthilkumar

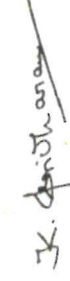
Dr.S.R.Senthilkumar
Convener



Dr.N.Venkateshan
President - LOC



Dr.S.Shasi Anand
Secretary



Dr.K.Sridharan
Chairman